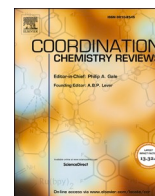




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Review

## Nanozyme-based strategies for efficient theranostics of brain diseases



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## ABSTRACT

Oxidative stress emerges as a critical mechanism underlying the development and progression of various brain diseases, arising from either increased generation of reactive oxygen species (ROS) or dysfunction of the antioxidant defense system. Nanozymes, possessing enzymatic catalytic capabilities alongside the physicochemical characteristics of nanosized materials, have showcased extraordinary promise in regulating the production or scavenging of ROS. Consequently, nanozymes have been utilized to detect the basic level and dynamic changes of neurochemical biomarkers closely related to physiological and pathological brain conditions, as well as to mitigate oxidative damage and facilitate the penetration of the blood-brain barrier. As such, nanozymes have emerged as a promising tool for developing diagnostic and therapeutic approaches for various neurological disorders. This review offers a comprehensive overview of the catalytic mechanisms exhibited by nanozymes, which have demonstrated tremendous potential in the effective diagnosis and treatment of brain disorders. Through summarizing the emerging methods for applying nanozymes, this review aims to provide directions on the rational design of nanozymes for the theranostics of brain diseases. Additionally, this review seeks to address the current challenges and delineate future directions for the advancement of this field.

**Abbreviations:** Ach, acetylcholine; AChE, acetylcholinesterase; AD, Alzheimer's Disease; ARSACS, autosomal recessive spastic ataxia of Charlevoix-Saguenay; AUR, Amplex UltraRed; A $\beta$ , amyloid  $\beta$  protein; BBB, blood-brain barrier; Bor, borneol; BSA, bovine serum albumin; CAs, carbonic anhydrases; CAT, catalase; CD, carbon dot; CDT, chemodynamic therapy; Ce, ceria; Ch, choline; ChOx, choline oxidase; CM, cerebral malaria; CPD, cyclobutane pyrimidine dimers; Cys, cysteine; DA, dopamine; DOPAC, 3,4-dihydroxyphenylacetic acid; ECM, experimental cerebral malaria; ECs, endothelial cells; Eda, edaravone; FAD, flavin adenine dinucleotide; GBM, glioblastoma multiforme; GDH, glucose dehydrogenase; GOx, glucose oxidase; GPx, glutathione peroxidase; GSH, reduced glutathione; GSSG, oxidized glutathione; H<sub>2</sub>O<sub>2</sub>, hydrogen peroxide; HD, Huntington's Disease; HDAC, histone deacetylase; HF<sub>n</sub>, human H-ferritin; HRP, horseradish peroxidase; HTT, huntingtin; IS, ischemic stroke; K<sub>M</sub>, Michaelis-Menten constant; k<sub>cat</sub>, catalytic constant; k<sub>cat</sub>/K<sub>M</sub>, specificity constant; LOx, lactate oxidase; MOF, metal-organic framework; MSN, mesoporous silica nanoparticles; MRI, magnetic resonance imaging; NAs, nano-alloys; NAD (P), nicotinamide adenine dinucleotide (phosphate); NCs, nanoparticle clusters; NLRP3, NLR family pyrin domain containing 3; NPs, nanoparticles; O<sub>2</sub><sup>-</sup>, superoxide anion radical; <sup>1</sup>O<sub>2</sub>, singlet oxygen; OH<sup>-</sup>, hydroxide ions; ONOO<sup>-</sup>, peroxynitrite; OPH, organo-phosphorus hydrolase; OXD, oxidase; PD, Parkinson's Disease; PDT, photodynamic therapy; PDNPs, polydopamine nanoparticles; POD, peroxidase; POM, polyoxometalate; POMD, POM with Wells-Dawson structure; PPO, polyphenol oxidase; PQQ, pyrroloquinoline quinone; PTT, photothermal therapy; RBCs, red blood cells; rGO, reduced graphene oxide; RNS, reactive nitrogen species; RONS, reactive oxygen and nitrogen species; ROS, reactive oxygen species; Se, selenium; Sec, selenocysteine; SERS, surface-enhanced Raman scattering; SNC, Substantia Nigra compact; SOD, superoxide dismutase; TBH, *tert*-butyl hydroperoxide solution; TBI, traumatic brain injury; Tf, transferrin; TMB, 3,3',5,5'-tetramethylbenzidine; TME, tumor microenvironment; TPH2, tryptophan hydroxylase-2; TPx, thiol peroxidase; ZIF, zeolitic imidazolate framework; UV, ultraviolet; V<sub>max</sub>, maximum reaction rate; \*NO, nitric oxide; \*OH, hydroxyl radical; 3,5-DTBC, 3,5-3,5-*Di-tert*-butylcatechol; 4-HNE, 4-hydroxynonenal;  $\alpha$ -syn,  $\alpha$ -synuclein.

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## 1. Introduction

Brain diseases, as a broad term, are used to describe a range of nervous system diseases affecting the structural and functional integrity of the brain,[1] including ischemic stroke (IS),[2] traumatic brain injury (TBI),[3] tumor,[4] and neurodegenerative disease.[5–7] Structurally, the brain is protected by the blood-brain barrier (BBB), which selectively prevents the ingress of deleterious substances, while resulting in low permeability of drugs. It is the Achilles Heel for the poor theranostics of brain diseases.[8,9] Functionally, the brain belongs to the central nervous system, responsible for consciousness and behavior.[10,11] The brain diseases are often accompanied by serious consciousness and behavior disorders. With the continuous growth and aging of the global population, the burden placed upon public healthcare by neurological disorders has increased significantly.[12,13] More importantly, brain diseases affect patients' quality of life seriously.[14,15] So new methods, strategies and technologies are called for the theranostics of brain diseases.

Redox assumes a paramount significance in the initiation and progression of brain diseases.[16] For example, ROS storm generated by reperfusion is the main cause of poor prognosis in IS.[17] Degenerative diseases and depression are always accompanied by increased oxidative stress.[5,18] Oxidative stress is increasingly used as an indicator for brain diseases treatment and prognosis evaluation. The clear core definition of nanozymes is "nanomaterials that catalyze the conversion of enzyme substrates to products and follow enzymatic kinetics (e.g. Michaelis-Menten) under physiologically relevant conditions".[19] However, given that nanozymes can also work under harsh conditions, the boundaries of the definition of nanozymes are fuzzy and will be updated as the field rapidly develops. Since first reported in 2007,[20] nanozymes have attracted considerable attention in redox regulation on account of obvious advantages over natural enzymes, including ease of synthesis in large quantities, cost-effectiveness, exceptional stability, versatile and tunable catalytic capabilities (Table 1).[21,22] Moreover, growing works have reported that nanozymes have the potential to penetrate the BBB by various pathways such as endocytosis and transcytosis.[23,24] Some nanozymes like nanozyme coated by ferritin have unique ability to effectively penetrate the BBB based on the transcytosis mediated by transferrin receptor 1.[25] Additionally, nanozymes can be easily modified by active substances,[26] peptides[27] and receptors [28] to improve their performance in penetrating the BBB. Given these advantages, nanozymes have been designed to effectively treat brain diseases and monitor the basic level and changes of neurochemical biomarkers related to brain physiological and pathological conditions, which confirms the feasibility of nanozymes to intervene in pathological processes.[29–31]

So far, several scholarly reviews have been published on the utilization of nanozymes in neurological disorders, covering IS, TBI and neurodegenerative diseases. However, the systematic consolidation of information concerning other important neurological diseases, such as brain cancer, cerebral malaria (CM), and depression, remains lacking. In

**Table 1**

Advantages and challenges of nanozymes compared with natural enzymes.

Advantages [21,22]	Challenges[32]
<ul style="list-style-type: none"> <li>• simpler preparation process</li> <li>• lower preparation cost</li> <li>• higher stability</li> <li>• more functions in theranostics of diseases</li> <li>• better controllability due to the easy modification</li> </ul>	<ul style="list-style-type: none"> <li>• catalytic activity and selectivity of most nanozymes are expected to improve</li> <li>• researches on more kinds of enzyme-like activities and nanozymes with multienzyme-like activities are needed</li> <li>• more energy to explore catalytic kinetics and mechanism of nanozymes needs to be devoted</li> <li>• more applicable scenarios should be developed</li> <li>• biosafety and potential toxicity still remain challenging</li> </ul>

this review article, we comprehensively summarize the catalytic mechanisms of nanozymes, systematically review their pre-clinical applications across diverse brain diseases (Fig. 1), and discuss the challenges in practical applications. We hope to provide a platform for sparking interests among researchers interested in nanozymes and promoting nanozymes to better serve for human health.

## 2. The enzyme-like activities and mechanisms of nanozymes

The natural enzymes encompass seven distinct classifications, which are oxidoreductases, transferases, hydrolases, lyases, isomerases, ligases, and translocases, responsible for biological reactions in living systems.[33] Among these, oxidoreductases-like, hydrolases-like, lyases-like and isomerases-like nanozymes have been developed (Table 2) (Fig. 2). [34]

Nanozymes typically exhibit a kinetic mechanism consistent with natural enzymes, namely Michaelis-Menten kinetics.[35] As one of the most typical enzyme kinetic equations, the Michaelis-Menten equation (1) represents the quantitative relationship between substrate concentration ( $[S]$ ) and enzyme reaction rate ( $v$ ), which is used to describe the ability of the enzyme to bind substrates, the catalytic reaction rate and the factors affecting reaction rate. Kinetic constants include the maximum reaction rate ( $V_{max}$ ), Michaelis-Menten constant ( $K_M$ ), catalytic constant ( $k_{cat}$ ) and specificity constant ( $k_{cat}/K_M$ ).[36]

$$v = \frac{V_{max}[S]}{K_M + [S]} \quad (1)$$

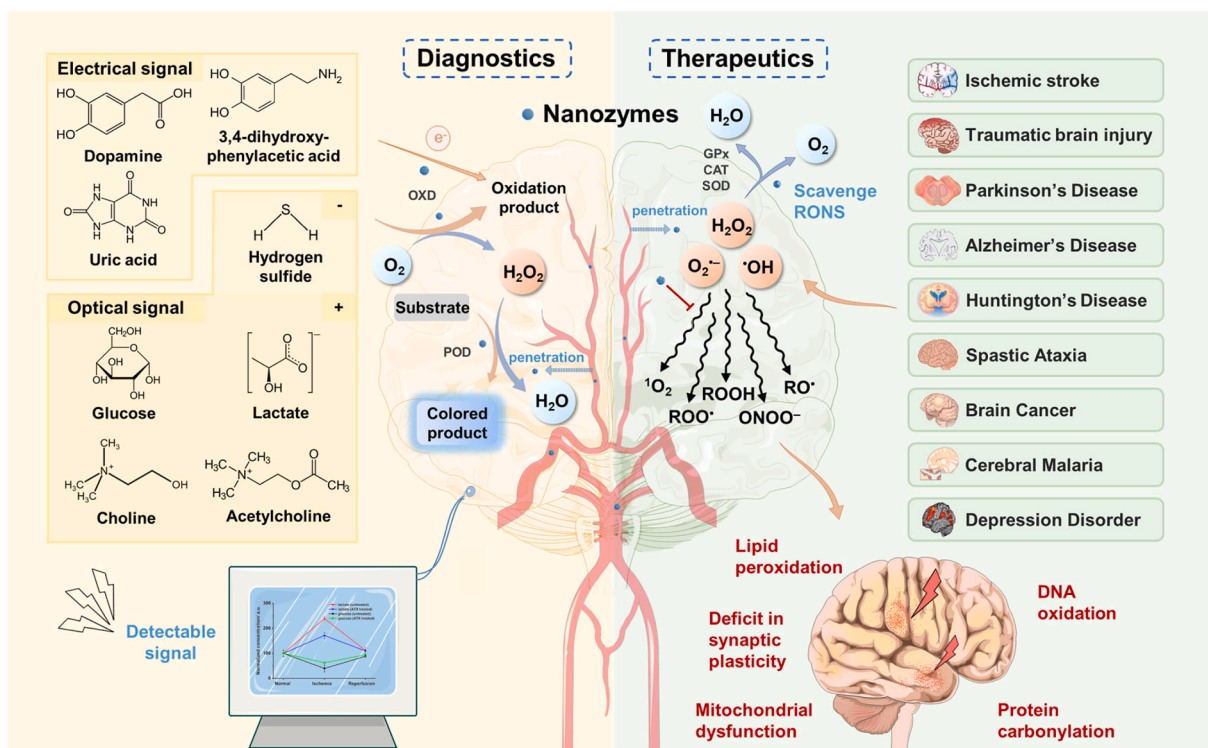
Based on the catalytic mechanisms of natural enzymes, this section will focus on reviewing the biomimetic catalytic mechanisms of various nanozymes. Given the multi-enzyme activity of some nanozymes, the mechanisms with multi-step/series reactions are also described, which can synergistically act on multiple substrates or be regulated by different environments to achieve specific catalysis.

### 2.1. Oxidoreductases

The main type of nanozymes are oxidoreductases, which are also the most studied and applied class. Oxidative stress stands as a negative factor precipitating brain dysfunction. In living organisms, the balance of redox levels is maintained by natural enzymes.[66] However, it may be disrupted under pathological states through the augmentation of ROS and reactive nitrogen species (RNS) or the impairment of antioxidant systems,[67] leading to oxidative stress. Oxidative stress damages biomacromolecules (proteins, lipids, and DNA), and blocks normal signaling pathways.[68,69] Reductases-like nanozymes, such as catalase (CAT), glutathione peroxidase (GPx), and superoxide dismutase (SOD), are considered as potential solutions to the above problems, and are expected to scavenge RONS and mitigate oxidative damage.[21,22,70] In addition, a general application of oxidases (OXD)-like and peroxidase (POD)-like nanozymes has been observed in the treatment of diseases such as wounds and cancers, and construction of multifunctional biosensors that sensitively detect neurochemical biomarkers closely related to brain physiological and pathological conditions.

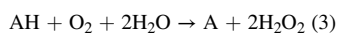
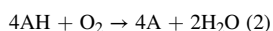
#### 2.1.1. Oxidase

Oxidation reactions are essential for aerobic metabolism, with ROS such as superoxide anion radical ( $O_2^{\bullet -}$ ), hydrogen peroxide ( $H_2O_2$ ), hydroxyl radical ( $\bullet OH$ ), and singlet oxygen ( $^1O_2$ ) as by-products.[71] With  $O_2$ , natural OXD can catalyze substrates to the corresponding oxidation products, while  $O_2$  is incorporated into the reaction products or accepts electrons for conversion into  $H_2O$  or  $H_2O_2$  (2, 3). OXD is usually named according to the substrates, such as glucose oxidase (GOx), etc. At present, extensive experimental evidences have substantiated that nanomaterials exhibit OXD-mimicking activity. For example, Au@Pt mimics ascorbate oxidase-like activity,[43] P-MoO<sub>3-x</sub> nanoparticles (NPs) mimic sulfite oxidase-like activity,[44] and Fe-N-doped graphene mimics

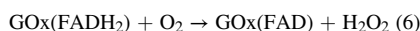
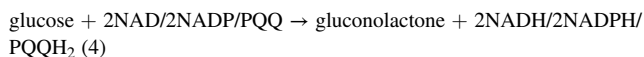


**Fig. 1.** Overview of nanozymes-based theranostics of brain diseases. Nanozymes engineered for enhanced brain targeting readily traverse the BBB. On the one hand, nanozymes can directly or indirectly catalyze neurochemicals to generate detectable signals, allowing for the monitoring of brain physiological or pathological conditions and the diagnosis of brain diseases. On the other hand, numerous brain diseases are intricately associated with the excessive production of ROS. ROS-mediated oxidative stress and excessive inflammatory responses seriously impair neurological functions. Nanozymes possess the remarkable capacity to modulate the generation or elimination of ROS, thereby realizing the efficacious management of brain diseases. Blood vessels and brain structure diagrams are provided by Servier Medical Art (<http://smart.servier.com>) under CC BY 3.0 license.

NADPH oxidase-like activity.[45] Most OXD-like nanozymes have lower  $K_M$  values than natural enzymes, meaning a high affinity for substrates. But their specificity is relatively low and they can catalyze multiple substrates such as 3,3',5,5'-tetramethylbenzidine (TMB), glucose, catechol, and *o*-phenylenediamine.[41]



Glucose oxidizing enzymes are divided into two types: glucose dehydrogenase (GDH) and GOx. GDH uses nicotinamide adenine dinucleotide (phosphate) (NAD(P)) or pyrroloquinoline quinone (PQQ) as the electron acceptor to oxidize glucose in one step (4).[72] While GOx uses a two-step mechanism, flavin adenine dinucleotide (FAD) cofactor assumes the role of an intermediate electron acceptor to oxidize  $\beta$ -D-glucose, leading to the formation of D-glucono- $\delta$ -lactone. Thereafter, the intermediate FADH<sub>2</sub> is oxidized by the terminal electron acceptor O<sub>2</sub> to generate FAD and H<sub>2</sub>O<sub>2</sub> (5, 6).[73] Both reaction types are dehydrogenation reactions, that is, primarily functioning as an electron acceptor, the oxidant exerts its influence without engaging in direct reactivity with the substrates.



In 2004, bare Au NPs were found to have the ability to oxidize glucose, being able to catalyze 21% of glucose into gluconate within 200 s. A series of kinetic experiments characterizing glucose oxidation revealed a linear correlation between catalytic activity and gold

concentration ( $10^{-5}$ - $10^{-6}$  M).[37] Subsequent studies supported this result and revealed how Au NPs catalyzed the oxidation of glucose. After hydroxide ions (OH<sup>-</sup>) as Brønsted base removed the C1 hydroxyl proton of glucose, the formed hydrated glucose anion was adsorbed on Au NPs, and O<sub>2</sub> was reduced to H<sub>2</sub>O<sub>2</sub> by nucleophilic attack (Fig. 3a).[38] This reaction follows typical Michaelis-Menten kinetics and bears a close resemblance to the reaction pathway of natural GOx. The only difference is that OH<sup>-</sup> replaces the His residue of GOx to break the C-H bond.[39] Based on this, Au NPs can be developed as a glucose sensor in combination with POD, or be developed for cancer therapy through depleting glucose in malignant cells.[74]

As a copper-containing enzyme, natural polyphenol oxidase (PPO) exhibits the capacity to facilitate the oxidation of an assortment of phenolic compounds, leading to the formation of highly active *o*-quinones. The oxidation is accomplished in two steps: after the ortho hydrogen of the existing hydroxyl group is hydroxylated, *o*-dihydroxybenzene is oxidized to *o*-benzoquinone (Fig. 3b). According to different substrates and mechanisms of action, PPOs are divided into three categories: catechol oxidase, tyrosinase, and laccase.[75]

Li *et al.* constructed MOF-818 with a trinuclear copper center designed to mimic the active site of natural catechol oxidase. Remarkably, the nanozyme exhibited activity in catalyzing catechol oxidation while demonstrating the absence of POD-like activity. In acidic or alkaline environments, MOF-818 specifically promoted the oxidation of Di-*tert*-butylcatechol (3,5-DTBC), but could not catalyze the oxidation of TMB and ABTS. It was found that the catalytic action of the tricopper(II) center resulted in the rapid conversion of 3,5-DTBC into 3,5-di-*tert*-butyl-*o*-benzoquinone (3,5-DTBQ) center, while O<sub>2</sub> was reduced to H<sub>2</sub>O<sub>2</sub> in the reoxidation catalytic cycle (Fig. 3c). After fixing the MOF-818 concentration, the oxidation kinetics were studied by changing the 3,5-DTBC concentration. Calculated values  $V_{\text{max}}$  ( $3.17 \times 10^{-6}$  M/s),  $K_M$

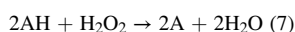
**Table 2**  
The enzyme-like activities of nanozymes.

Catalytic types	Subclasses	Featured Nanozymes	References	
oxidoreductases	oxidase	glucose oxidase	Au NPs [37–39]	
		lactate oxidase	Co <sub>4</sub> N/C [40]	
		catechol oxidase	MOF-818 [41]	
		ascorbate oxidase	CHzyme (Cu-His) [42]	
			Au@Pt [43]	
		sulfite oxidase	P-MoO <sub>3-x</sub> NPs [44]	
		NADPH oxidase	Fe-N-doped graphene [45]	
		peroxidase	peroxidase	AuNPs@CDs [46]
				Fe <sub>3</sub> O <sub>4</sub> [20,47]
			glutathione peroxidase	Se@pDA [48]
		Au NPs (Sec-Arg-Gly-Asp-Cys) [49]		
	catalase	CeO <sub>2</sub> NPs [50,51]		
	superoxide dismutase	Ag, Pd, Pt, and Au and alloys [52]		
	hydrolases	phosphatase	Au <sub>4-x</sub> M <sub>x</sub> (M = Ag, Pd, Pt) [53]	
			Carbon dot nanozyme [54]	
			MOFs [55]	
			Ce-FMA-MOF [56]	
Zn-heptapeptide bionanozymes [56]				
glycosidase		Cu <sub>3</sub> P [57]		
esterase		PtNP [58]		
nuclease		Pt [59]		
lyases		carbonic anhydrase	Amino acid-functionalized fullerene [60]	
			ZIF-8 [61]	
isomerases	topoisomerase I	TiO <sub>2</sub> [62]		
		CeO <sub>2</sub> [63]		
		Carbon dots [64]		
	peroxynitrite isomerase	H-RGO [65]		

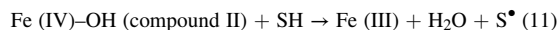
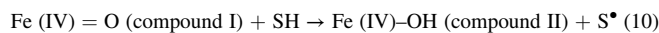
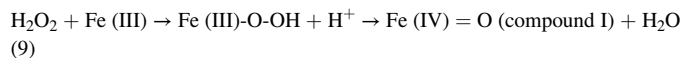
( $8.10 \times 10^{-4}$  M),  $k_{\text{cat}}$  ( $0.383 \text{ s}^{-1}$ ) and  $k_{\text{cat}}/K_{\text{M}}$  ( $4.73 \times 10^2 \text{ M}^{-1}\text{s}^{-1}$ ) indicated that MOF-818 has excellent catechol oxidase-like activity. [41] Li *et al.* devised a strategy to engineer a novel nanozyme (CHzyme) with catechol oxidase-like properties, incorporating copper and histidine as key constituents. Furthermore, they combined the enzyme-catalyzed reaction with Schiff base chemistry to realize a cascade reaction. CHzyme oxidizes catechol to o-quinone, and the carbonyl of o-quinone reacts with the amino group in o-phenylenediamine, forming a fluorescent product. In contrast to the conventional enzyme-linked immunosorbent assay, the detection sensitivity is increased by 15 times. [42] Catecholamines (dopamine (DA), norepinephrine, and epinephrine), as key neurotransmitters, are considered to be important indicators of neuroblastoma, schizophrenia, Parkinson's disease (PD), and other neurological diseases, [76] so the investigation into the PPO-like activity of nanomaterials and elucidation of its distinctive catalytic mechanism hold immense significance.

### 2.1.2. Peroxidase

The POD family is very huge and commonly exists in various types of cells in eukaryotes. It has multiple isoforms, and the major component is heme peroxidase containing Fe (III) protoporphyrin IX as a prosthetic group. It mainly uses peroxide ( $\text{H}_2\text{O}_2/\text{ROOH}$ ) as an electron receptor to catalyze oxidation of various substrates (7, 8). [77] Typical substrates include phenols, amines, polycyclic aromatic hydrocarbons, and sulfonates. [78]



In the catalytic cycle, the first step is the interaction of  $\text{H}_2\text{O}_2$  with the Fe(III) atoms in the heme group to generate the transient intermediate iron hydroperoxide. Subsequently, the distal oxygen of iron hydroperoxide is protonated, promoting the cleavage of the O-O bond, forming compound I containing a Fe(IV) oxoiron base center and a porphyrin radical (9). Then, with the participation of reducing substrate, compound I is converted into compound II, and the porphyrin free radical is eliminated, but iron still exists in the Fe(IV) state (10). Finally, compound II returns to the resting state of the enzyme (11).



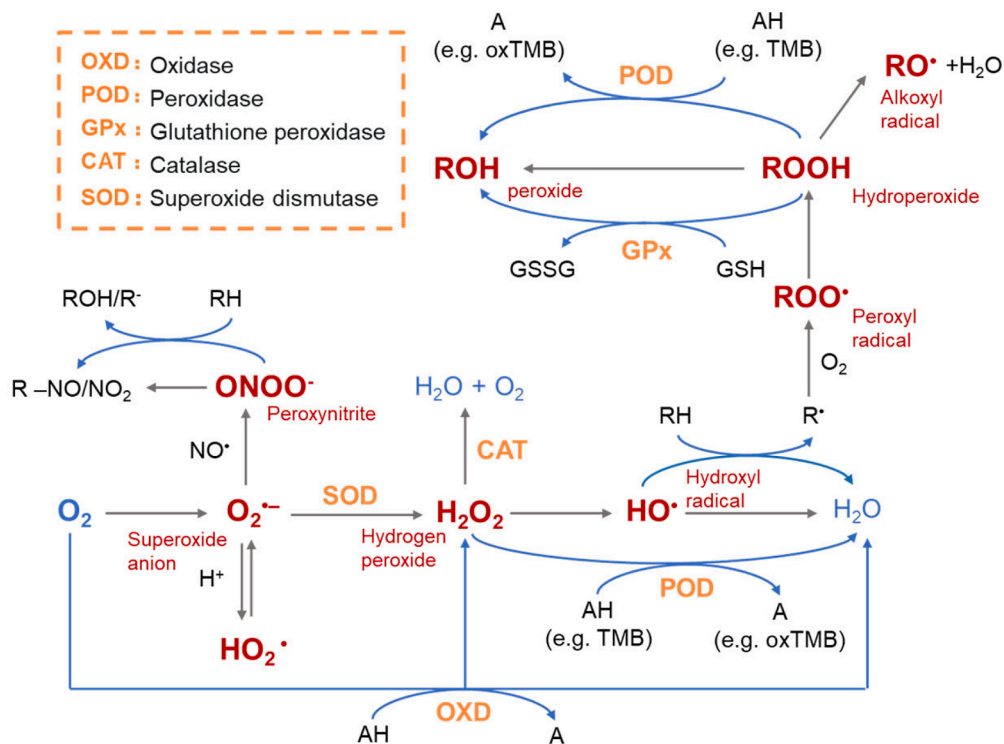
The POD-like activity of nanomaterials is the earliest confirmed activity, and the types of POD-like nanomaterials have gradually expanded from  $\text{Fe}_3\text{O}_4$  to other metals, metal oxides, carbon nanozymes, and MOFs. The inorganic nanomaterial-based nanozymes typically provide intrinsic POD-mimicking activity, which catalyze the oxidation of colorless TMB to blue amine or diamine. This reaction can be used as disease diagnosis strategies. [79] For example, AuNPs@CDs designed by Zheng *et al.* facilitated the electrons transfer from TMB to  $\text{H}_2\text{O}_2$ . [46] PtNPs/GO proposed by Zhang *et al.* exhibited POD-like activity and catalyzed the color reaction of TMB in the presence of  $\text{H}_2\text{O}_2$ . [80] In addition, common chromogenic substrates include Amplex UltraRed (AUR), o-phenyl-enediamine, and di-azo-amino-benzene. For example, Chang *et al.* developed Au/Ag NPs, which can catalyze the reaction of AUR with  $\text{H}_2\text{O}_2$  to form fluorescent products, thereby detecting acetylcholine (ACh) and diagnosing neurodegenerative diseases.

One of the most typical nanozymes with POD-like activity is  $\text{Fe}_3\text{O}_4$  nanozyme. Similar to natural horseradish peroxidase (HRP), there is a ping-pong mechanism and Michaelis–Menten kinetics involved in  $\text{Fe}_3\text{O}_4$  nanozyme. In terms of the catalytic mechanism, the nanozyme first binds and reacts with the corresponding substrate, and releases the product before reacting with the next substrate. Taking the color reaction of  $\text{H}_2\text{O}_2$  and TMB catalyzed by  $\text{Fe}_3\text{O}_4$  nanozyme as an example, the  $K_{\text{M}}$  value of substrate TMB catalyzed by  $\text{Fe}_3\text{O}_4$  is smaller than that of natural HRP, but the  $K_{\text{M}}$  value of another substrate  $\text{H}_2\text{O}_2$  catalyzed by  $\text{Fe}_3\text{O}_4$  is larger than that of natural HRP. Furthermore, the  $k_{\text{cat}}$  value of  $\text{Fe}_3\text{O}_4$  is larger than that of natural HRP due to the large specific surface area and enriched active sites. [20] Previous studies on the catalytic mechanism of  $\text{Fe}_3\text{O}_4$  or other POD-like nanozymes have led to the conclusion of Fenton or Fenton-like reactions, as well as surface-mediated electron transfer processes. [81] Recently, Dong *et al.* reported that  $\text{Fe}_3\text{O}_4$  nanozymes possess self-depletion properties due to electron transfer within the internal atoms. [47] The structure and intramolecular charge transfer can greatly affect the catalytic performance of materials, [82] indicating that surface and internal atoms should be considered simultaneously in the design, modulation, and application of nanozymes. More specifically, Zhao *et al.* modulated the Co number near N in the  $\text{Co}_x\text{-N}$  nanocomposite, engineering the electronic properties at the N center to mimic lactate oxidase (LOx). [40] Ji *et al.* controlled the electronic configuration of the monatomic iron active site by the precise coordination of phosphorus and nitrogen to achieve POD-mimicking catalytic activity and kinetics on par with that of natural enzymes. [83] Those researches remind us to pay attention to the geometric structure and electronic coordination between active centers and nonmetals in the process of simulating natural enzymes.

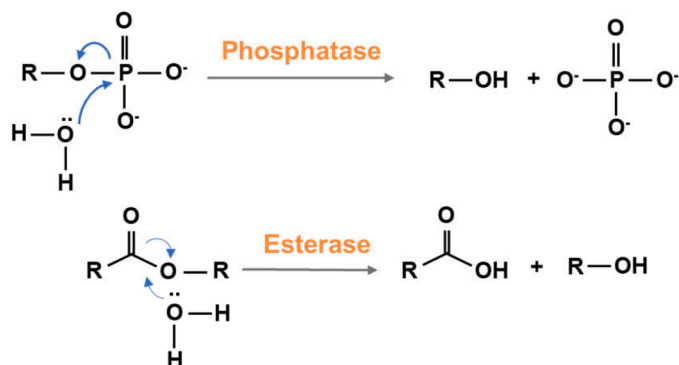
### 2.1.3. Glutathione peroxidase

GPx isozymes are mainly composed of 8 subfamilies, which fulfill a crucial function in antioxidant defense due to their inherent ability to facilitate the reduction of ROS. Except for GPx5, GPx7, and GPx8, the

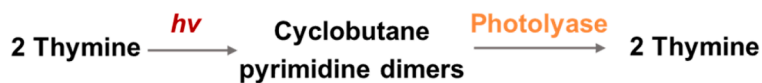
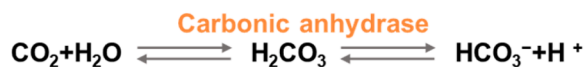
## 1. Oxidoreductases



## 2. Hydrolases



## 3. Lyases



## 4. Isomerases

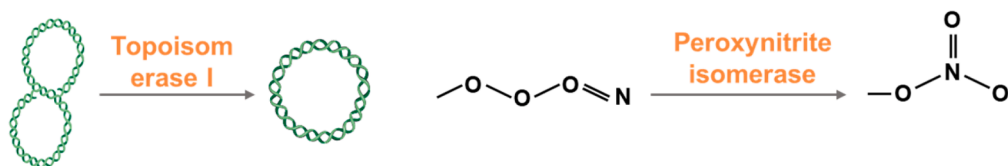
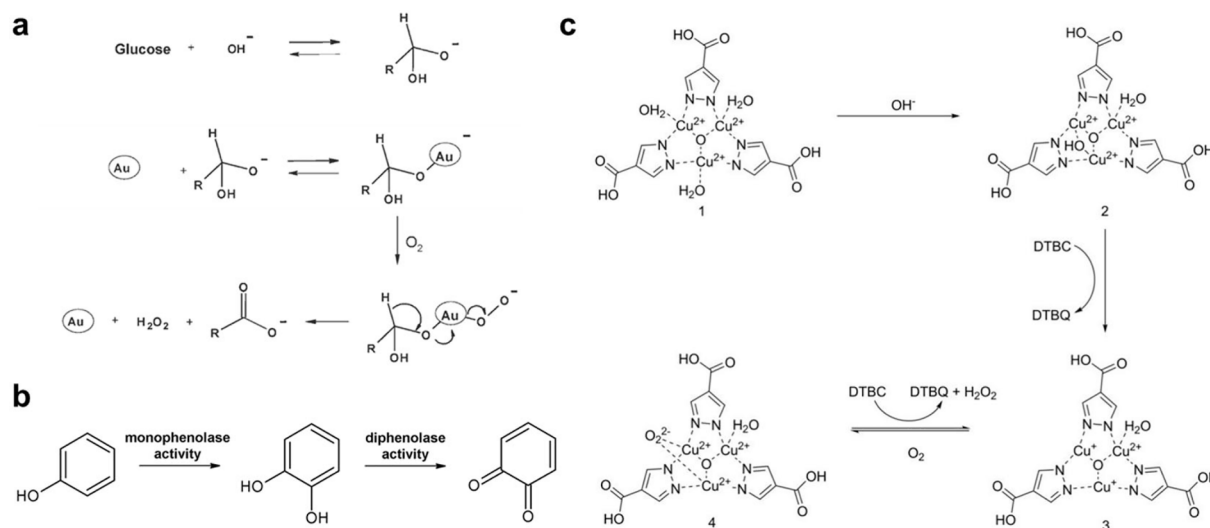
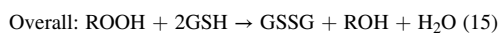
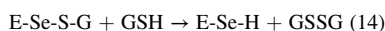
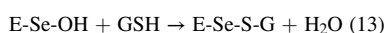
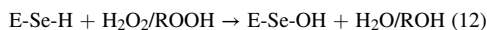


Fig. 2. The chemical basis of reactive oxygen and nitrogen species (RONS) generation, and the examples of chemical reactions involving oxidoreductases-like, hydrolases-like, lyases-like and isomerases-like nanozymes. Adapted from Ref. [16]. Copyright 2019 MDPI.



**Fig. 3.** The diagrammatic representation elucidating the intricate enzymatic mechanism of oxidase or oxidase mimics. (a) Catalytic pathway of Au NPs mimicking GOx. (b) Catalytic process of PPO. (c) Proposed mechanism of catechol oxidase activity of metal-organic framework (MOF)-818. Reprinted with permission from Refs. [38,75] and [41]. Copyright 2006 Wiley-VCH, 2020 Springer Nature and 2020 American Chemical Society.

catalytic sites are all selenocysteine (Sec) residues, and selenium (Se) mediates the redox cycle. In the widely accepted GPx catalytic cycle, selenol (E-Se-H) first reacts with peroxide ( $\text{H}_2\text{O}_2/\text{ROOH}$ ) to be oxidized to selenate (E-Se-OH) (12). E-Se-OH is considered as the key intermediate, reacting with reduced glutathione (GSH) to yield the formation of selenyl sulfide adduct (E-Se-S-G) (13). Then, another GSH reacts with E-Se-S-G to restore the enzyme to its functional state, forming oxidized glutathione (GSSG) (14). [84] Overall, two equivalent amounts of GSH are oxidized to disulfide and water, while  $\text{H}_2\text{O}_2/\text{ROOH}$  is reduced to the corresponding  $\text{H}_2\text{O}/\text{ROH}$  (15). Glutathione reductase, in conjunction with its corresponding coenzyme NADPH, can reduce GSSG back to GSH again. After this cycle, E-Se-H will react with the next  $\text{H}_2\text{O}_2/\text{ROOH}$ .

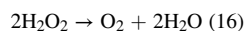


Inspired by the above catalytic mechanism, Huang *et al.* designed a composite nanozyme (Se@pDA) composed of selenium and polydopamine. Similar to natural GPx, the mechanism of Se nanocomponents catalyzing  $\text{H}_2\text{O}_2$  decomposition may follow the ping-pong mechanism. The Se component first reacts with  $\text{H}_2\text{O}_2$  to produce selenium oxide. Then, the intermediate product, selenium oxide, reacts with GSH to restore the Se form, and GSH is oxidized to GSSG. Moreover, the excellent GPx-mimicking ability of Se and the reducing property of polydopamine can exert synergistic antioxidant effects to protect cellular components from oxidative stress. [48] Zhang *et al.* prepared Au nanozymes modified with Se pentapeptide. The N-terminal of the polypeptide is a catalytic group Sec with GPx-like activity, and the C-terminal is a binding group cysteine (Cys). Kinetic analysis results show that the addition of Au NPs changes the catalytic mechanism of the peptide from a ping-pong mechanism to an ordered mechanism. Moreover, the  $k_{\text{cat}}/K_M$  of GSH and  $\text{H}_2\text{O}_2$  by selenopeptide-functionalized AuNPs is one order of magnitude higher than that of free selenopeptides. Based on this, it can be speculated that Au NPs can enhance the catalytic activity by restricting the mobility and conformation of the polypeptide, exposing most of the Sec groups to the environment in the presence of substrates GSH and  $\text{H}_2\text{O}_2$ . It is worth mentioning that the  $K_M$  value of the substrate  $\text{H}_2\text{O}_2$  is smaller than that of GSH. Similar to natural GPx, the selenopeptide-functionalized AuNPs only show specific

recognition of GSH. [49]

#### 2.1.4. Catalase

The CAT plays an important role in defending against  $\text{H}_2\text{O}_2$ , which is one of the origins of radicals through Fenton reaction or analogous pathways. [85] In the body, CAT found in virtually all aerobic organisms can effectively remove  $\text{H}_2\text{O}_2$  by decomposing  $\text{H}_2\text{O}_2$  into  $\text{O}_2$  and  $\text{H}_2\text{O}$ , thereby defending cells against oxidative stress (16): [86]

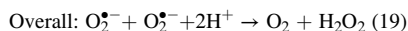
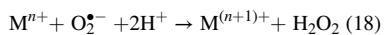
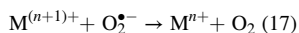


As mentioned above, the CAT-mimicking nanozymes have better stability and lower costs than natural CAT. Nowadays, there is a growing anticipation for CAT-mimicking nanozymes to emerge as viable alternatives to natural CAT in many application fields. Dedicated efforts have been undertaken by scientists to investigate the catalytic mechanism of CAT-mimicking nanozymes to achieve this goal. Hitherto, various nanomaterials have been recognized as CAT-mimicking nanozymes, including metal/metal oxides, Prussian blue, MOFs, and N-doped NPs. [87] These nanomaterials also follow Michaelis-Menten kinetic reactions and exhibit high binding affinities for  $\text{H}_2\text{O}_2$ . One of the simple catalytic mechanisms represented by part of metal-based nanozymes is based on the redox reaction by multiple valence switches. Patel *et al.* synthesized ceria (Ce) NPs, which utilized regenerative redox switching between  $\text{Ce}^{3+}$  and  $\text{Ce}^{4+}$  ions to selectively remove ROS. [50] However, the catalytic mechanism is not clear for coupled electron transfer, and emphasis should be placed on exploring not only the reaction conditions but also the structural and electronic characteristics of nanozymes in the forthcoming research endeavors. Other metal-based nanozymes like Au [37], Ag [88], and Pd [89] NPs are likely to change their enzyme-like activity in different pH conditions, which is attributed to the different adsorption energies of the molecules under various pH polarizations. Considering the structural and electronic characteristics of  $\text{CeO}_2$  NPs, Wang *et al.* demonstrated that oxygen vacancies have a significant impact on the catalytic performance by altering the redox state of Ce, [51] based on the perpetual motion model of  $\text{CeO}_2$  catalytic mechanism. [90] Xu *et al.* precisely concluded that there are two major types of mechanisms of CAT-like nanozymes. The difference is whether  $\text{H}_2\text{O}_2$  chooses to break H-O or the O-O bond preferentially when it gets absorbed. [87]

#### 2.1.5. Superoxide dismutase

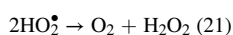
Natural SOD is usually composed of protein and metal cofactors, and

can be divided into four types according to different cofactors: CuZn-SOD, Mn-SOD, Fe-SOD, and Ni-SOD.[91] Accompanied by the alternate redox of metal ions in the active site, SOD possesses the ability to catalyze the disproportionation of  $O_2^{\bullet-}$ , resulting in the production of  $O_2$  and  $H_2O_2$  (19),[92] and the catalytic reaction formula is as follows:

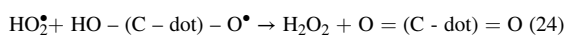
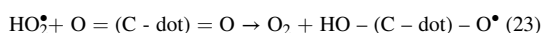


Among them, the metal ion in the oxidized state accepts an electron from  $O_2^{\bullet-}$  to convert into a reduced state, and the  $O_2^{\bullet-}$  that loses an electron to form  $O_2$  (17). Next, the metal ion in the reduced state transfers an electron to another  $O_2^{\bullet-}$ , and the  $O_2^{\bullet-}$  that has gained the electron combines with two protons to form  $H_2O_2$  (18).

The kinetics of SOD-like nanozymes also follow this ping-pong mechanism, which can effectively eliminate  $O_2^{\bullet-}$ . [24] Since the SOD-like activity of C60 was reported in the 1990s,[93] SOD-like activity has been found in a variety of metal-doped nanozymes (such as Au, Ag, Pd, Pt, Mn, V, and other metals and their oxides) and carbon dot (CD). For SOD-like nanozymes of metal and alloy, the mechanism predominantly encompasses two parts, namely the protonation of  $O_2^{\bullet-}$  and the adsorption rearrangement of  $HO_2^{\bullet}$  on the metal surface.



As a Brønsted base,  $O_2^{\bullet-}$  reacts readily with  $H_2O$  to form  $HO_2^{\bullet}$  and  $OH^-$  (20). In light of the pronounced exothermicity exhibited by the adsorption of  $HO_2^{\bullet}$  onto the nanozyme surface, this easy adsorption shifts the equilibrium towards more  $HO_2^{\bullet}$  generation. At the same time, the very low  $E_{act}$  means that the adsorbed  $HO_2^{\bullet}$  groups are easily converted into  $O_2$  and  $H_2O_2$  (21). [52] For CD nanozymes, the catalytic mechanism was revealed by Gao *et al.* through surface structure adjustment and theoretical calculations. The hydroxyl and carboxyl groups of CDs serve as binding sites, while the carbonyl moieties of CDs serve as the catalytic sites.



Consistent with the above process,  $O_2^{\bullet-}$  existing in an alkaline environment readily captures a proton from aqueous solution to form  $HO_2^{\bullet}$  and  $OH^-$  (22). The carbonyl group within the CD transforms the hydroxyl group through the oxidation of  $HO_2^{\bullet}$  to  $O_2$  (23). Afterward,  $\pi$ -OH is converted into carbonyl by reducing  $HO_2^{\bullet}$  free radicals to  $H_2O_2$  (24). [53]

$O_2^{\bullet-}$  is highly oxidative and can be further metabolized into other ROS, causing oxidative stress. Therefore, as an important antioxidant, SOD-like nanozymes hold immense promise in the realm of preventing and treating a multitude of ROS-related diseases.

## 2.2. Hydrolases

Hydrolase is a general class of enzyme catalyzing the hydrolysis reaction, or a distinctive type of transferase using water as the receptor for transferred group. Hydrolases are involved in 13 different bio-transformation, including esterases, phosphoesterases, amidases, proteases, and glycosidases.[55] Thus, hydrolases have a wide range of substrates.

Given that phosphate bonds exhibit the highest activity among the aforementioned hydrolytic bonds, most studies on hydrolase mimics are centered around this. Elena *et al.* entrapped the organo-phosphorus hydrolase (OPH) protein within a poly ion complex, with a size below

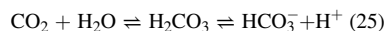
100 nm, to propose hydrolytic nanozymes which have demonstrated remarkable efficacy in safeguarding animals against the detrimental effects of pesticide exposure, specifically paraoxon, as well as the toxic impact of warfare agent known as VX.[94] In addition to specific modification of natural enzymes to enhance their performance, integrating natural metal ion catalytic centers moieties within/onto nano-materials have been deeply studied, following the discovery of the structural motif featuring a binuclear metal center within the amido-hydrolase superfamily. Xia *et al.* directed their attention towards systematically assembling the functional components found in OPH catalytic centers within MOFs including bimetallic  $Zn^{2+}$  centers.[54] Oxygen vacancy is also a key factor in catalytic hydrolysis reactions. Lin *et al.* fabricated ceria NPs with abundant and regenerable oxygen vacancy, which exhibit the ability to dephosphorylate phosphor-tyrosine. [95] Except for inorganic-derived nanomaterials, biomolecule-sourced nanozymes (bionanozymes) have been reported by Liang *et al.* Drawing inspiration from the fundamental structure of natural hydrolases, they successfully engineered three hydrolase-mimicking bionanozymes based on oligopeptide, which were able to hydrolyze substrates with side-chains through the self-assembly of histidine-rich heptapeptides induced by Zn.[56] Misfolding and aggregation of proteins leading to forming toxic substances is a common feature of many neurodegenerative diseases covering Alzheimer's disease (AD) and PD. Nanozymes that mimic protease-like activities hold promise for the elimination of amyloid  $\beta$  protein ( $A\beta$ ),  $\alpha$ -synuclein ( $\alpha$ -syn) and tau.[96,97] Moreover, the ligand is another essential factor in the design of hydrolytic nanozymes. Different lengths of ligands can yield nanozymes with different active site densities.[98]

However, the extensive substrates range of hydrolases and limited knowledge of their catalytic mechanisms present challenges in the development of new hydrolytic nanozymes. Li *et al.* designed a hydrolytic nanozyme based on a Ce-FMA-MOF structure through integration and induction of previous data.[55] That reminds us of an effective way to design and discover new nanozymes based on machine learning or chemoinformatic models. Hydrolytic nanozymes are expected to be a new field in urgent need of extensive research.

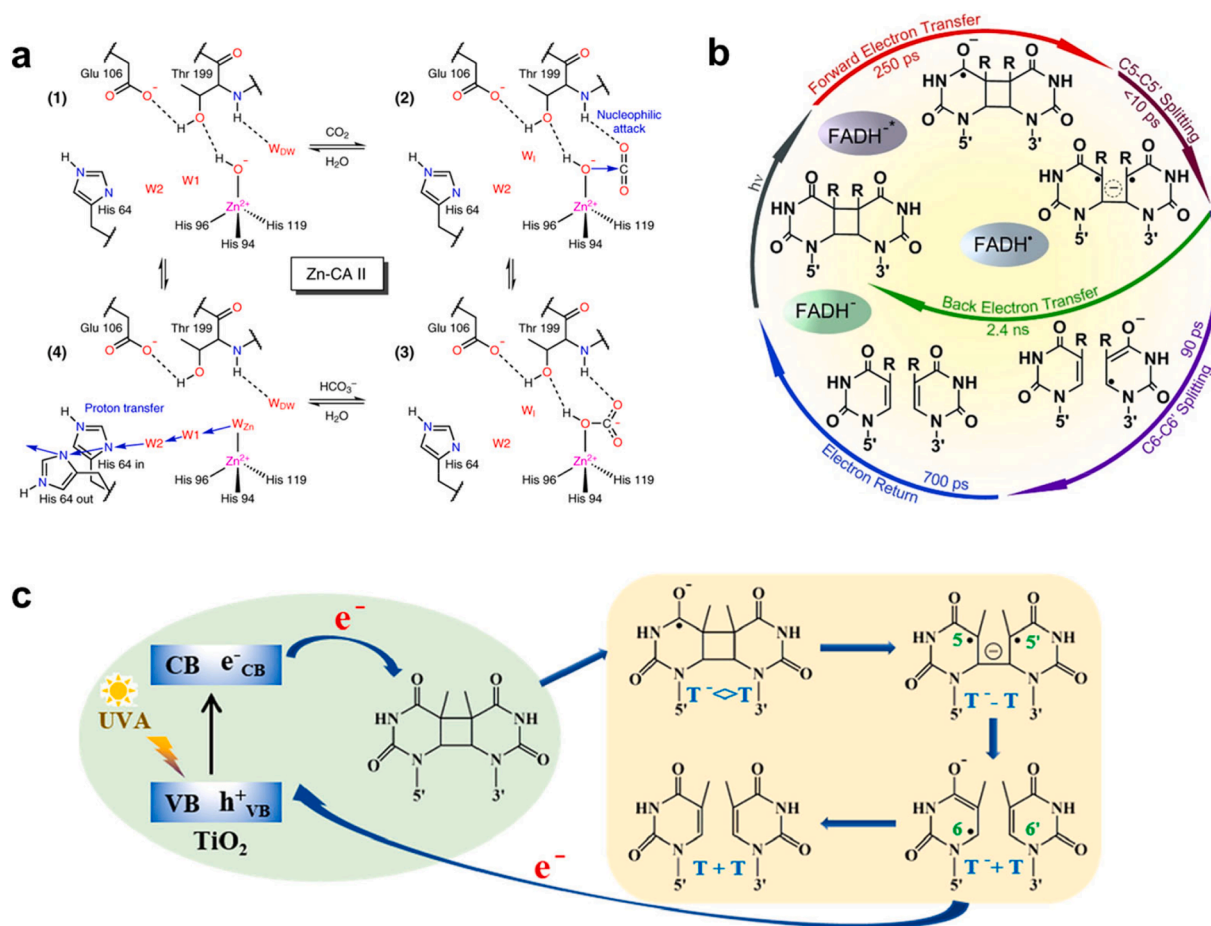
## 2.3. Lyases

The catalytic mechanism of lyases is to break the chemical bonds such as C-C, C-O, C-N, and C-S. It often participates in reversible reactions, requiring only one substrate for the forward reaction and two substrates for the reverse reaction. Through their cleavage capabilities, lyases change the molecular architecture, and the products often form new double bonds or ring structures. At the same time, the reaction does not require the participation of water or oxygen.

Lyases, including aldolase, hydratase, and deaminase, participate in many different types of biochemical reactions. For example, carbonic anhydrases (CAs) serve as catalysts for the fundamental process of life: the hydration of  $CO_2$  to form bicarbonate and proton. As a subclass of metalloenzymes, the catalytic site within the majority of CAs contains  $Zn^{2+}$ . [99] The catalytic mechanism consists of several steps (25):



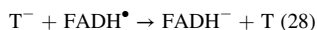
First,  $CO_2$  binds to the active site, and the  $OH^-$  bound by  $Zn^{2+}$  carries out a nucleophilic attack on  $CO_2$  to form an enzyme- $HCO_3^-$  adduct. Subsequently, due to the weak interaction between  $HCO_3^-$  and  $Zn^{2+}$  ions,  $HCO_3^-$  is replaced by  $H_2O$  at the active site. Finally, proton is transferred through the pathway of  $W_{Zn-H_2O} \rightarrow W1 \rightarrow W2 \rightarrow His64$ , regenerating  $OH^-$  bound to  $Zn^{2+}$  (Fig. 4a).[100] The  $CO_2$  hydration/  $H_2CO_3$  dehydration reaction is very slow, while the dissociation or ionization of  $H_2CO_3$  is very rapid and always in equilibrium under physiological conditions.[101] In recent years, to mitigate the greenhouse effect, capturing and sequestering  $CO_2$  by simulating the activity of natural CAs has emerged as a prominent subject of research.[102] Demirsoy *et al.*



**Fig. 4.** The diagrammatic representation of the catalytic mechanisms of lyase or lyase mimics. (a) The specific mechanism of CO<sub>2</sub> hydration reaction catalyzed by Zn-CA II. (b) Complete photocatalytic cycle for repairing UV-induced DNA damage by DNA photolyase. (c) The repair mechanism of CPD by TiO<sub>2</sub>. Reprinted with permission from Refs. [100,103] and [62]. Copyright 2023 Springer Nature, 2011 National Academy of Science, and 2022 American Chemical Society.

used amino acid-functionalized fullerene nanostructures to mimic different enzymes. It can self-assemble in an aqueous environment to form multiple active sites. The histidine- and threonine-functionalized fullerene derivative HT, which is the most similar nanocatalyst to the  $\alpha$ -CA active site containing histidine, threonine, and glutamic acid. Histidine is used to coordinate Zn<sup>2+</sup> ions to achieve the purpose of water activation, and threonine is responsible for increasing the nucleophilicity of OH<sup>-</sup>. [60] A novel zeolitic imidazolate framework (ZIF-8)-based nanozyme was proposed by Chen *et al.* Zn<sup>2+</sup> with less imidazolium coordination on the outer surface can mimic the active site of human CAII, promoting the generation of OH<sup>-</sup> bound to Zn<sup>2+</sup>, and thus have catalytic properties similar to human CAII. [61]

Additionally, the nanozyme alternatives of DNA photolyases have also been developed. Ultraviolet (UV) irradiation induces the formation of cyclobutane pyrimidine dimers (CPDs) through the linking of two neighboring pyrimidine bases in a DNA strand, thereby blocking the process of replication and transcription. [103] This damage can be repaired by DNA photolyase with FADH<sup>-</sup> molecule as an active cofactor.



In brief, the photorepair mechanism is that the cofactor FADH<sup>-</sup> is excited by absorbing blue light directly or transferring energy in antenna chromophores (26). Excited FADH<sup>-\*</sup> delivers an electron to CPD, which splits into two thymine bases (27). Finally, the electron returns to the

cofactor, restoring enzyme activity (28) (Fig. 4b). [104] Zhou *et al.* found that TiO<sub>2</sub> can be used as a photolyase mimic to repair CPD damage of DNA. Anatase/rutile mixed phase TiO<sub>2</sub> (P25) has excellent photochemical properties due to high blue light transmittance and electron mobility. The spectral result revealed the maximum repair rate of P25 was 1.59  $\mu\text{M}/\text{min}$ . Moreover, the transient spectra and kinetic data indicated that the repair of CPD by P25 is based on the electron transfer mechanism. The photogenerated electrons originating from P25 undergo transfer transition to CPD, resulting in the instability of CPD structure, and then sequentially cracking C5-C5' and C6-C6' of cyclobutane. After the CPD is split into T and T<sup>-</sup>, the electrons of T<sup>-</sup> return to the holes of TiO<sub>2</sub> to complete the photocatalytic cycle and repair the damaged DNA (Fig. 4c). In addition to excellent photocatalytic properties, small-sized TiO<sub>2</sub> NPs are easily taken up by cells and have good biocompatibility. [62] Similar to TiO<sub>2</sub>, CeO<sub>2</sub> is also a semiconductor and can generate photoelectrons. Under visible light irradiation, CeO<sub>2</sub> can successfully cleave 71.9 % of CPD into monomers, while only 2.4 % of CPD in the control group was cleaved, revealing the photolyase-like activity of CeO<sub>2</sub>. Moreover, CeO<sub>2</sub>, an antioxidant, can inhibit the generation of ROS, and thus inhibit ROS to completely decompose CPD into fragments and improve the photocatalytic selectivity. In addition, the  $K_M$  and  $k_{\text{cat}}$  indicate that the photocatalytic kinetics of CeO<sub>2</sub> has strong surface-dependent reactivity, and surface defects will effectively enhance the affinity between CeO<sub>2</sub> and CPD. [63]

#### 2.4. Isomerases

Isomerases promote intramolecular rearrangements, which can

catalyze the transformation of substrates into compounds with the same molecular formula but different atomic spatial arrangements. Most reactions are from a single substrate to a single product.[105] Depending on the specific reaction, it can be subdivided into epimerase, cis–trans isomerase, tautomerase, etc. Isomers widely exist in basic substances such as amino acids, sugars, and nucleic acids in nature. Isomerases can realize the interconversion between isomers, for example, alanine racemases catalyze the chiral conversion of L- and D-alanine,[106] glucose isomerases catalyze the interconversion of glucose and fructose.[107]

Unlike natural enzymes, most of the reported nanozymes do not possess the enantioselective ability. Recently, Qu *et al.* grafted multitudinous D- or L-amino acids on the surface of ceria NPs as selectors for chiral recognition, which demonstrated a promising approach for designing stereoselective nanozymes.[108] Carbon-based materials have attracted widespread interest in the development of isomerase artificial mimics due to their easy functionalization.

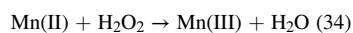
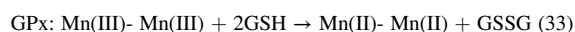
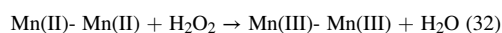
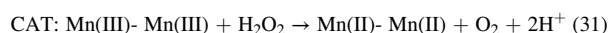
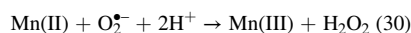
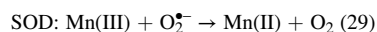
Li *et al.* designed a chiral CD derived from Cys, which exhibits a resemblance to topoisomerase I and can bind to DNA intercalation via an extended graphitic structure. By generating  $\cdot\text{OH}$  to cleave the DNA phosphate backbone, it mediates DNA single-strand breaks, thereby regulating DNA topology. Similar to the regulation of gene expression by topoisomerase I, chiral CD can promote gene expression.[64] A heme-functionalized reduced graphene oxide (H-rGO) nanosheet was constructed by Vernekar *et al.* Under physiological conditions, the hybrid nanosheets exhibited significant peroxynitrite isomerization and reduction capabilities. The reduction/isomerization mechanism is similar to that of the iron porphyrin complex.[65]

## 2.5. Multi-enzyme system

In organisms, a complete biochemical process is often not completed by one enzyme alone, but by several biocatalytic reactions mediated by multi-enzyme complexes that occur simultaneously or sequentially.[109] Although various nanozymes with oxidoreductases-like, hydrolases-like, lyases-like, or isomerases-like properties have been prepared and applied, in order to bridge the gap between nanozyme catalysis and whole-cell catalysis, the development of multi-nanozyme systems that perform multi-step/tandem reactions is indispensable.[110] Currently, the multi-enzyme activities of nanozymes mainly include dual-enzyme activities such as OXD-POD, POD-CAT, and CAT-SOD. In addition, three or more enzyme-like activities are also gradually being developed.

The diagnosis and treatment of brain diseases will greatly benefit from multi-nanozyme systems. As mentioned previously, brain diseases are often caused by the exacerbation of oxidative stress, a process mediated by excessive production of ROS. Antioxidant enzymes possess the capability to scavenge ROS, maintain redox balance, and reduce cell damage and local inflammation caused by oxidative stress. In addition, pro-oxidative enzymes play a crucial role in the detection of neurochemical biomarkers and the treatment of brain tumors.[111] However,

when traditional pro-oxidative enzymes and antioxidant enzymes work independently, their functions are relatively simple, and may consequently fall short of the demands for effective diagnosis and treatment of brain diseases. If nanozymes with diverse enzymatic activities are developed, it becomes possible to precisely modulate the levels of ROS within brain cells through synergistic effects and cascade reactions.[110] For example, Singh *et al.* found that  $\text{Mn}_3\text{O}_4$  NPs can simulate the activities of three types of antioxidant enzymes: SOD, CAT, and GPx. Similar to natural Mn-SOD,  $\text{Mn}_3\text{O}_4$  NPs first convert  $\text{O}_2^{\cdot-}$  into  $\text{O}_2$  by reducing the metal center (29), and then convert  $\text{O}_2^{\cdot-}$  into  $\text{H}_2\text{O}_2$  by oxidizing the metal center (30). The product of the SOD catalytic reaction,  $\text{H}_2\text{O}_2$ , can subsequently serve as a substrate for CAT and GPx catalysis reactions. CAT directly acts on  $\text{H}_2\text{O}_2$  to produce  $\text{H}_2\text{O}$  and  $\text{O}_2$  (31, 32), while GPx uses GSH as a cofactor to convert  $\text{H}_2\text{O}_2$  into  $\text{H}_2\text{O}$  without producing  $\text{O}_2$  (33, 34).[112]



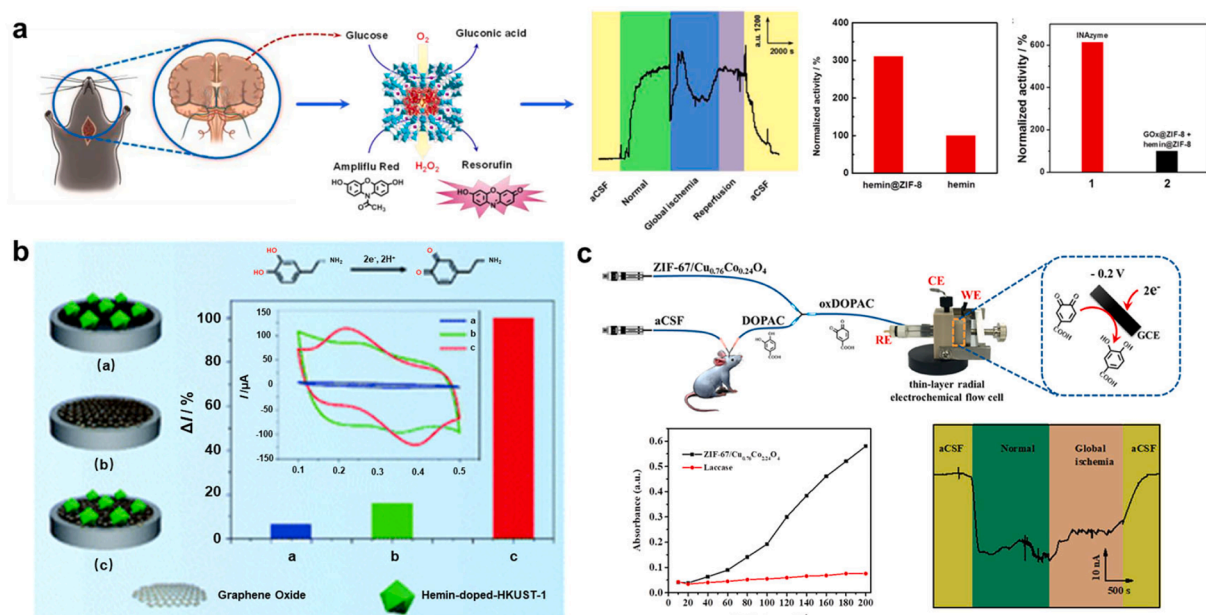
Furthermore, under the catalysis of POD,  $\text{H}_2\text{O}_2$  generates more toxic free radicals, such as  $^1\text{O}_2$  and  $\cdot\text{OH}$ , which can increase oxidative damage to biological macromolecules. Wei *et al.* synthesized a multi-enzyme nanoreactor (IrRu-GOx@PEG NPs) composed of natural GOx, ultra-small IrRu alloy NPs with CAT-like and POD-like dual enzyme activities, and surface-modified PEG. First, natural GOx degrades glucose into  $\text{H}_2\text{O}_2$  and gluconic acid, accomplishing two critical roles: 1) increasing the concentration of  $\text{H}_2\text{O}_2$  in the tumor microenvironment (TME) and providing substrates for subsequent reactions; 2) cutting off the tumor's nutritional source and promoting tumor cell apoptosis through starvation therapy. Secondly, IrRu NPs catalyze the upstream  $\text{H}_2\text{O}_2$  to generate  $\text{O}_2$  and highly toxic  $^1\text{O}_2$ . Among them,  $\text{O}_2$  can relieve hypoxia in the TME, thereby lifting the efficiency limit of GOx-based starvation therapy and forming a virtuous cycle.  $^1\text{O}_2$  can directly kill tumor cells and inhibit tumor growth.[113]

Designing nanozyme systems with multiple types of enzyme activities will help further advance the detection of analytes. This approach not only enables the fabrication of cascade sensing systems, but also expands the range of analytes and improves overall detection performance.[114] For example, Wu *et al.* synthesized FeCo co-doped carbon spheres (FeCo@C) with intrinsic POD and OXD-like activities. The multi-enzyme activity of FeCo@C is affected by external environments such as pH. After catalyzing for 6 min in a pH 3.6 environment, FeCo@C showed high OXD-like activity and can oxidize TMB to oxTMB, resulting

**Table 3**

A summary of nanozymes for brain disease diagnosis.

Nanozymes	Activities	Neurochemical biomarkers detected	Detection range ( $\mu\text{M}$ )	Detection limit ( $\mu\text{M}$ )	References
INzyme (GOx/hemin@ZIF-8)	POD	Glucose	100–1000	1.7	[120]
AuNPs@MIL-101@GOx	POD	Glucose & Lactate	10–200	4.2 5.0	[121]
$\text{V}_2\text{O}_5$ nanobelts	POD, GOx	Glucose	200–5000	0.33	[122]
Hemin-doped HKUST-1/rGO	POD	Dopamine	0.03–10	0.0327	[125]
ZIF-67/ $\text{Cu}_{0.76}\text{Co}_{2.24}$ O <sub>4</sub> NSs	POD, SOD, GPx, Laccase	3,4-dihydroxyphenylacetic acid	0.5–20	0.15	[126]
ZIF-L-Co-10 mg Cys	Ascorbate oxidase, Laccase, Glutathione oxidase	Uric acid	0.2–50	0.067	[128]
PBA NC	LOx	Hydrogen sulfide	0.1–20	0.033	[106]
MIL-101(Fe)	POD	ACh & Ch	0.1–10 0.01–100	0.020 8.9	[132]



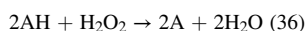
**Fig. 5.** Nanozymes for the detection of neurochemicals to diagnose brain diseases. (a) Uninterrupted real-time monitoring of cerebral glucose by an integrated sensing platform utilizing INAzyme. (b) Detection of DA by an ultrasensitive electrochemical sensor based on Hemin-doped-HKUST-1/rGO. (c) Monitoring of DOPAC by an online electrochemical detection platform based on the ZIF-67/Cu<sub>0.76</sub>Co<sub>2.24</sub>O<sub>4</sub> NSs with laccase-like activity. Reproduced with permission from refs [120,125] and [126]. Copyright 2021 Royal Society of Chemistry and 2016, 2020 American Chemical Society.

in color changes. However, FeCo@C showed higher POD-like activity after catalyzing for 3 min in a pH 4.4 environment. By controlling the pH value to avoid mutual interference of the dual enzyme activities, dual detection of hydroquinone and H<sub>2</sub>O<sub>2</sub> is achieved.[115] Therefore, the emergence of multi-nanozyme systems brings great hope to the further development of the field of diagnosis and treatment of brain diseases.

### 3. Nanozyme-based diagnosis of brain diseases

Monitoring neurochemical biomarkers assumes paramount significance in unraveling the functions of these substances within the physiological and pathological phenomena of the brain, as well as for the diagnosis of various brain diseases. Therefore, online *in vivo* detection systems based on electrochemical or optical detection are current research hotspots. Based on OXD/POD-mimicking activities, nanozymes have been explored for monitoring single or multiple brain neurochemicals (Table 3).

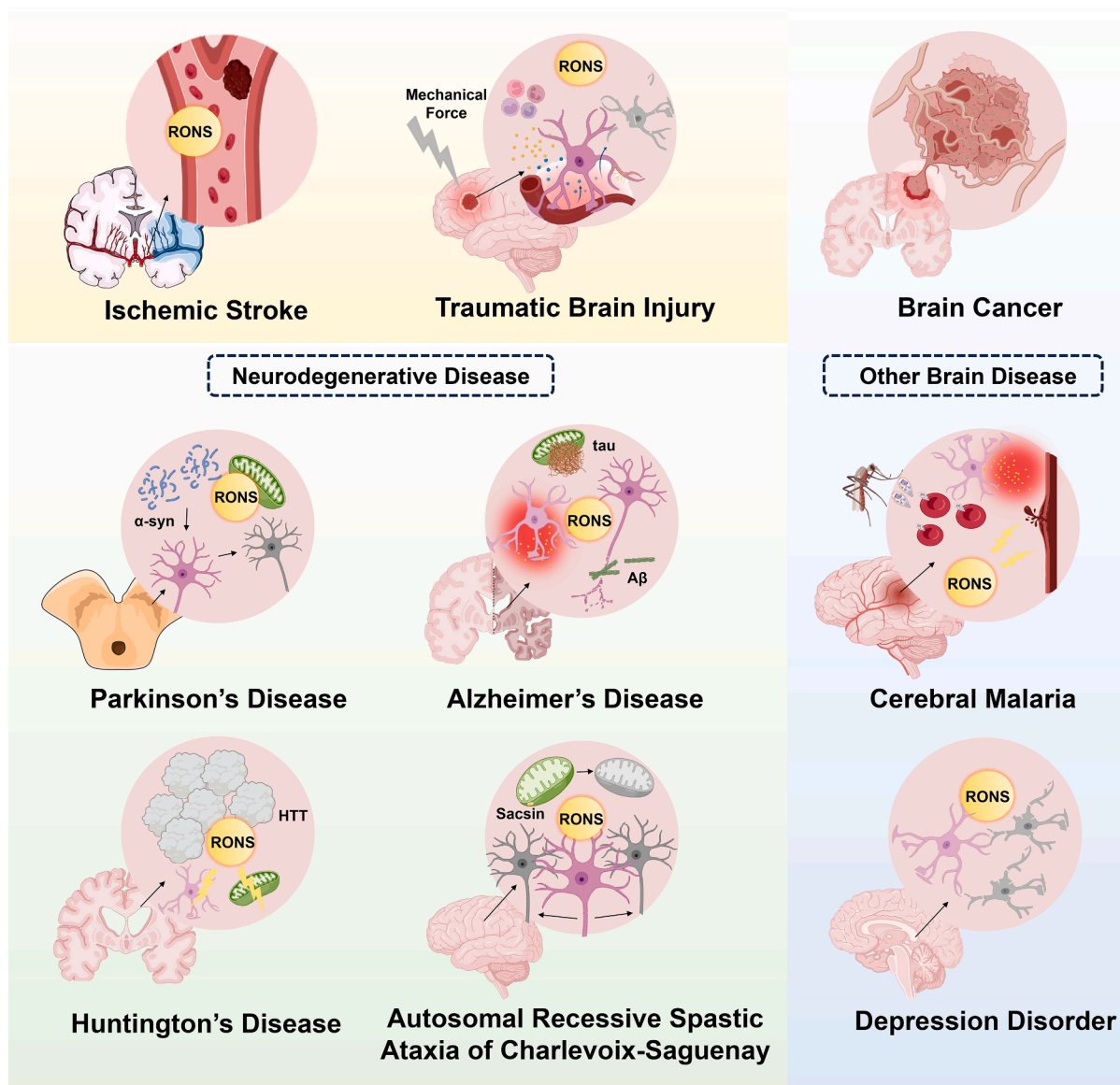
The human brain, a mere 2 % of body mass, consumes approximately 20 % of the energy harnessed from glucose metabolism.[116,117] In addition to being the main energy source, glucose also participates in significant activities such as the synthesis of neurotransmitters, learning and memory building, as well as pathologic conditions including oxidative stress.[118] Thus, disturbances in glucose levels may lead to cognitive impairment, memory impairment, and other disorders. Previous studies have designed biosensors based on GOx and HRP. In this biosensing system, glucose is oxidized to H<sub>2</sub>O<sub>2</sub> by GOx (35), and then TMB or other organic substrates react with H<sub>2</sub>O<sub>2</sub> under the catalysis of HRP to generate color products (36).[119].



Nanozymes with POD-like activity have been used as HRP substitutes due to their remarkable advantages. By embedding heme and GOx in ZIF-8 nanostructures, Cheng *et al.* prepared integrated nanozymes (GOx/hemin@ZIF-8). In contrast to free heme, the POD-mimicking activity of hemin@ZIF-8 was increased by more than 3 times. This was because free heme tended to form aggregates, whereas the ZIF-8

maintained heme as a monomer. Moreover, the significant enhancement of the enzyme-mimicking properties is attributed to the elevated specific surface area and the enrichment effect conferred by the ZIF-8. Notably, the co-encapsulation of GOx and heme within the ZIF-8 mimicked multiple enzymes confined in subcellular compartments in biological systems. This nanoscale proximity effect significantly improved the enzyme stability and synergistically enhanced the catalytic activity by lowering diffusion hindrance and reducing the degradation of highly reactive intermediates. In addition, the online detection system composed of integrated nanozymes and microdialysis technology can continuously monitor the dynamic fluctuations in glucose levels within the striatal region of the brain after ischemia/reperfusion (Fig. 5a). Other enzymes may also be applied in this strategy, such as LOx.[120] By combining the surface-enhanced Raman scattering (SERS) activity and catalytic properties of Au NPs, the same team designed Au NPs@MIL-101-based nanozymes. Due to the POD-like activity, it can catalyze the conversion of Raman inactive reporter molecules (i.e., leucomalachite green) into active reporter molecules (i.e., malachite green). Then, the nanozymes were assembled with GOx and LOx to further construct integrated nanozymes, which demonstrated remarkable efficacy in the real-time monitoring of glucose and lactate dynamics closely related to normal and abnormal states within the brains of living animals.[121] The multi-enzyme activity exhibited by many nanozymes provides a new idea for glucose monitoring which requires the participation of natural GOx. Ding *et al.* found that V<sub>2</sub>O<sub>5</sub> nanoribbons have bifunctional enzyme-mimicking capabilities (GOx and POD), and utilized the tandem enzymatic activity of V<sub>2</sub>O<sub>5</sub> nanoribbons to continuously monitor glucose in the rat brain. This method has exceptional selectivity, heightened sensitivity, high stability, and a broad dynamic detection range (0.2 to 5 mM), [122] which further breaks the traditional colorimetric detection process and truly realizes glucose detection without natural enzymes.

Furthermore, through rational design and regulation of the activity of nanozymes, the types of neurochemicals detected online are being continuously expanded. As a prominent catecholamine neurotransmitter, DA plays an important role in neuromodulation encompassing cognition, motor coordination, motivation, and reward.[123] It has



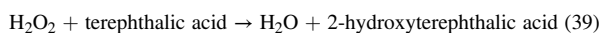
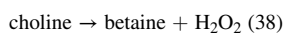
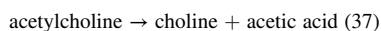
**Fig. 6.** Schematic diagram summarizing the pathological features of different brain disorders, including IS, TBI, neurodegenerative diseases (PD, AD, Huntington's disease (HD), and autosomal recessive spastic ataxia of Charlevoix-Saguenay (ARSACS) and other brain diseases (brain cancer, CM, and depression disorder). This Figure created with BioRend.com and Servier Medical Art (<http://smart.servier.com>).

been implicated in various brain diseases including PD, schizophrenia, depression, and attention deficit hyperactivity disorder. For example, PD ensues from the gradual degeneration and loss of dopaminergic neurons.[124] Therefore, achieving sensitive, accurate, and low-cost detection of DA assumes paramount significance in the clinical diagnosis of brain diseases. Kang *et al.* synthesized novel multifunctional MOF nanozymes doped with hemin, and further composited them with reduced graphene oxide (rGO) to fabricate an ultrasensitive electrochemical sensor for DA detection. The catalytic current of HKUST-1/rGO/GCE doped with hemin exhibited a remarkable enhancement, surpassing that of rGO/GCE and Hemin-doped-HKUST-1/GCE by approximately 6.3 times and 16 times, signifying the synergistic catalytic ability (Fig. 5b). Based on the improved catalytic activity, the sensor has a higher sensitivity ( $1.224 \mu\text{A} \mu\text{M}^{-1}$ ) and a lower detection limit ( $3.27 \times 10^{-8} \text{M}$ ). [125] As an important metabolite of DA, 3,4-dihydroxyphenylacetic acid (DOPAC) within the cerebrospinal fluid of PD patients is markedly lower than that of normal controls, which is consistent with severe striatal DA depletion.[126,127] Liu *et al.* prepared ZIF-67/Cu<sub>0.76</sub>Co<sub>0.24</sub>O<sub>4</sub> NSs by a one-step alcohol-hydrothermal

method using ZIF-67 as a template and achieved continuous and near real-time response DOPAC measurement. Different from the series of studies above which mainly used the POD-like activity of nanozymes to realize real-time detection of neurochemicals *in vivo*, this study was the basis of the laccase-mimicking activity to monitor DOPAC. The combination of online electrochemical system with microdialysis technology enabled the acquisition of DOPAC concentration in cerebral dialysate both pre- and post-ischemia. The results showed that when the rats were in ischemia for 30 min, the current dropped notably, which corresponded to a decrease in DOPAC concentration from  $12.3 \pm 1.8 \mu\text{M}$  ( $n = 3$ ) before ischemia to  $9 \pm 1.2 \mu\text{M}$  ( $n = 3$ ) (Fig. 5c).[126] The online electrochemical system and optical detection platform built by this team have also realized the online continuous detection of brain uric acid and hydrogen sulfide,[128,129] which provided new ideas for the diagnosis of brain diseases, including afflictions like injury and neurodegenerative diseases.

Another important neurotransmitter, ACh, is closely related to neurotransmission and modulation.[130] Its precursor choline (Ch) is not only an essential nutrient for the body, but also a neuroprotective

agent for maintaining the normal development of the brain, which is beneficial to alleviate age-related memory decline and cognitive impairment.[131] Abnormal levels of Ch and ACh can induce a variety of brain diseases, including Down syndrome, PD, AD, and schizophrenia. Therefore, the quantitative analysis of Ch and ACh holds immense importance in the diagnosis of related brain diseases. A fluorescent biosensor utilizing MIL-101(Fe) nanozyme as its core component was developed by Gou *et al.* MIL-101(Fe) with outstanding POD-mimicking activity can be used to detect Ch and ACh after coupling with acetylcholinesterase (AChE) and choline oxidase (ChOx). The cascade catalytic reaction is mainly divided into three steps. AChE catalyzes the hydrolysis of ACh into acetic acid and Ch (37). Subsequently, ChOx catalyzes the cleavage of Ch to produce H<sub>2</sub>O<sub>2</sub> (38). Finally, MIL-101(Fe) catalyzes the decomposition of H<sub>2</sub>O<sub>2</sub> to generate highly active  $\cdot$ OH, and oxidizes the non-fluorescent substrate terephthalic acid to the highly fluorescent product 2-hydroxyterephthalic acid (39).[132]



In the precise analysis of biomolecules, enhancing the catalytic efficiency of nanozymes remains necessary. Therefore, some studies have combined defect engineering with nanozymes. Defects have long been considered undesirable features of crystals because disruptions in the periodic arrangement of atoms lead to disturbances in the geometry and electronic structure. However, with progressively better understanding of catalytic processes, defect engineering has emerged as a practical approach for adjusting the structure and properties of nanomaterials. In most cases, the defects directly assume the role of active centers or co-catalytic sites. Moreover, the embeddedness of defects may activate nearby atoms to become active sites through the deformation of local geometry and electronic structure, which effectively reduces the gap in enzymatic activity between nanozymes and natural enzymes.[133] By doping Cys, Ren *et al.* introduced structural defects into ZIF-L-Co. When doped with 10 mg Cys, the multi-enzyme activities of ZIF-L-Co were increased several times. The researchers conducted multiple experiments to explore the origins of high activity, and the conclusions can be divided into the following aspects: 1) Cys with high affinity for Co<sup>2+</sup> disrupts the equilibrium between Co<sup>2+</sup> and N within ZIF, causing lattice distortion, and bringing abundant coordinative unsaturated sites. 2) The surface area increases, which promotes the adsorption of substrates on the surface of the material. The detection system utilizing ZIF-L-Co-10 mg Cys was able to monitor UA levels in the striatum of mouse brains after ischemia–reperfusion injury.[128]

#### 4. Nanozyme-based treatment of brain diseases

Thanks to their scalability and designability, nanozymes show great potential in biomedicine. Researchers designed different types of nanozymes to effectively treat brain diseases by improving brain targeting, enhancing the activity of antioxidant enzymes, inhibiting inflammatory reactions, or achieving combined reactions (Fig. 6).

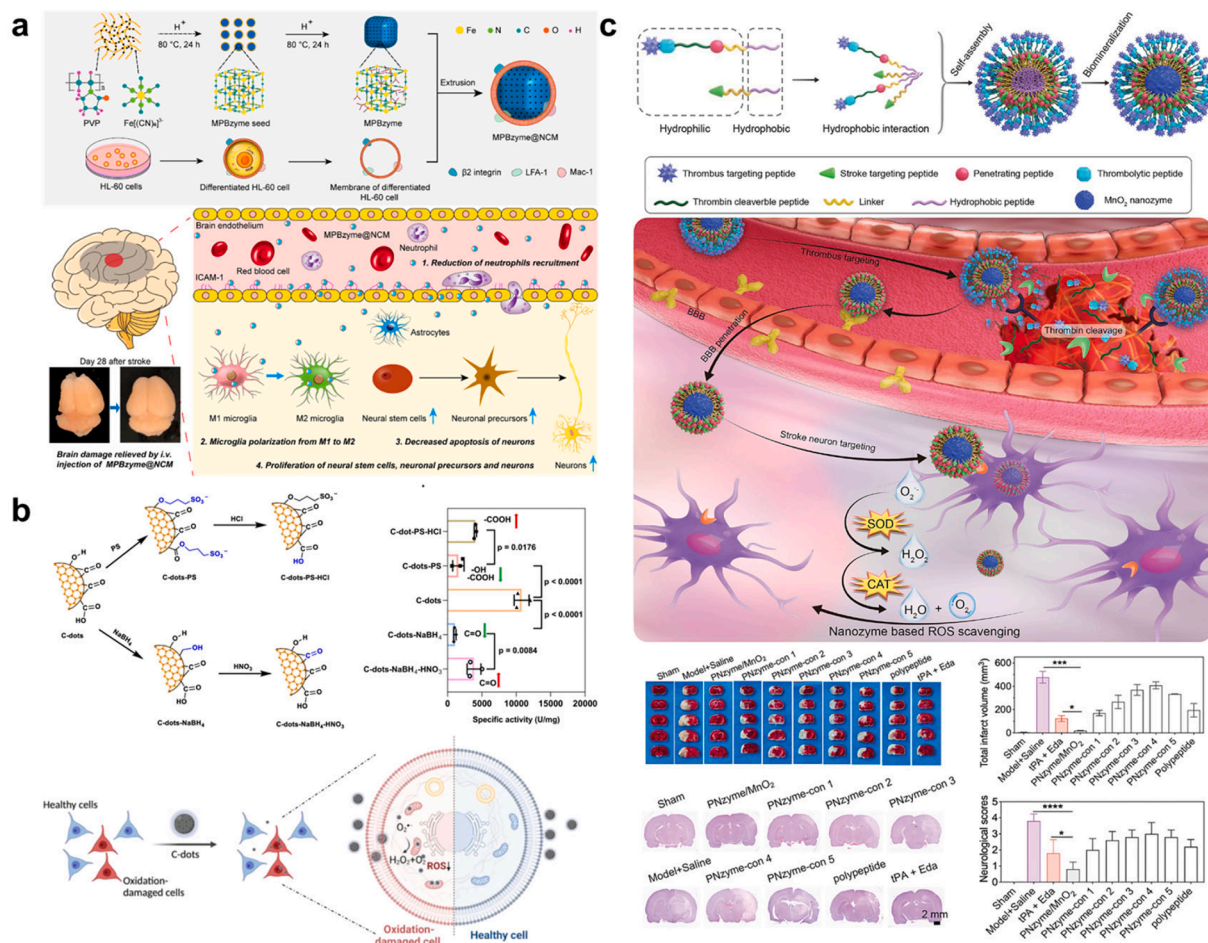
##### 4.1. Ischemic stroke

As the first leading cause of disability and the second leading cause of death worldwide, stroke affects over 101 million people currently.[134] It can be broadly divided into IS (71 %) and hemorrhagic stroke (29 %). [2] IS generally refers to cerebral ischemia and hypoxia caused by embolism or thrombus blocking cerebral blood vessels.[135,136] Clinically, vascular recanalization is often achieved through intravenous thrombolytic therapy or mechanical thrombectomy to save the ischemic penumbra.[137] However, after ischemia/reperfusion, numerous RONS are produced, including O<sub>2</sub><sup>•-</sup>, H<sub>2</sub>O<sub>2</sub>,  $\cdot$ OH, nitric oxide ( $\cdot$ NO), and

peroxynitrite (ONOO<sup>-</sup>).[138] They mediate oxidative stress and instigate multiple immoderate inflammatory reactions, which impair neurological function, and thus seriously affect the prognosis of patients.[139] Specifically, RONS lead to irreversible structural damage and dysfunction of brain tissues by strongly oxidizing proteins, nucleic acids, and lipids.[140] Besides, RONS activate microglia and stimulate inflammatory cells to secrete interleukin-1 $\beta$ , interleukin-6, and other cytokines, thereby mediating inflammation and immune response.[141] Endogenous antioxidant enzymes are excessively consumed during disease progression and are difficult to adequately supplement externally due to the low stability and high production costs of natural enzymes.[138] For this reason, a series of free radical scavengers, such as edaravone (Eda), have been used in IS as neuroprotective agents. Despite their beneficial effects, low bioavailability, poor selectivity, poor BBB permeability, and severe side effects have greatly limited their development.[142,143] Compared with these typical antioxidants, nanozymes have higher catalytic activity, bioavailability, and stability, resulting in extended residence time in tissues. Therefore, nanozymes have emerged as a viable strategy for treating IS.

Oxygen vacancies on the surface of the ultrasmall ceria nanozymes allow reduced state (Ce<sup>3+</sup>) and oxidized state (Ce<sup>4+</sup>) to coexist, thereby scavenging RONS through electron transfer between Ce<sup>4+</sup> and Ce<sup>3+</sup>. [144,145] In 2012, ceria nanozymes with SOD and CAT mimetic activities were prepared by Kim *et al.* to prevent ischemia by scavenging ROS and reducing apoptosis.[146] However, ultra-small ceria nanozymes exhibit a short half-life in blood circulation, and easily agglomerate. In addition, the accumulation of nanozymes in the brain may disrupt BBB and cause neurological dysfunction. Hence, it is of great significance to accomplish BBB penetration and protection simultaneously. Bao *et al.* designed EA/P-CeO<sub>2</sub> to enhance brain accumulation through receptor-mediated transcytosis. EA/P-CeO<sub>2</sub> relied on the enzyme-mimicking activity of CeO<sub>2</sub> and loaded Eda to synergistically eliminate ROS for effective treatment of IS.[147] He *et al.* adopted another strategy. They encapsulated CeO<sub>2</sub> NPs with ZIF-8 to change the surface properties of CeO<sub>2</sub> and prevent its aggregation. Moreover, ZIF-8 demonstrates notable efficacy in prolonging blood circulation time and improving the penetration of BBB. CeO<sub>2</sub>@ZIF-8 effectively inhibited oxidative damage to brain neurons as well as damage caused by inflammation.[148]

The ratio of Ce<sup>3+</sup> to Ce<sup>4+</sup> and the particle size greatly limited the catalytic activity of CeO<sub>2</sub> NPs.[149] Therefore, other nano-antioxidants (such as MnO<sub>2</sub>, Prussian blue, Fe<sub>3</sub>O<sub>4</sub>, CDs, and melanin) have been successively developed for IS treatment.[138,150,151] Using appropriate strategies to modify nanozymes to improve their brain targeting is one of the research directions. Zhao *et al.* designed a manganese dioxide nanozyme (Eda-MnO<sub>2</sub>@Tf, EMT) targeting transferrin (Tf) and loading with Eda. EMT exhibited excellent brain targeting and strong ROS scavenging capability. Besides, the Mn<sup>2+</sup> released in the weakly acidic environment could serve as a means of monitoring therapeutic progress through magnetic resonance imaging (MRI).[30] Mesoporous Prussian blue nanozymes coated on neutrophil-like cell membranes (MPBzyme@NCM) were designed by Feng *et al.* One of the pathological hallmarks after stroke is the interplay between leukocytes and inflamed microvascular endothelial cells (ECs) in the brain. Inspired by this, the transfer of neutrophil membranes to the surface of nanozymes could potentially enhance their specific targeting towards inflamed microvascular ECs in the brain, thereby enabling their entry into the damaged brain and subsequent phagocytosis by microglia. MPBzyme@NCM mediated the long-term treatment of IS by driving microglial polarization to M2, reducing neutrophil recruitment, and inhibiting neuronal apoptosis (Fig. 7a). This research work explored a new method for nanozymes to break through the BBB. However, how to further improve the delivery efficiency of nanozymes into the damaged brain through this entry method still needs to be studied.[152] Another research direction is to design nanozymes with elevated antioxidant activity. Liu *et al.* designed Co-Fe<sub>3</sub>O<sub>4</sub> co-doped nanozymes to target RONS in the



**Fig. 7.** Strategies aimed at enhancing the therapeutic efficacy of nanozymes for IS. (a) Improvement in brain targeting. MPBzyme@NCM targeting inflamed brain microvascular ECs. (b) Enhancement of antioxidant activity. C-dots with high SOD-mimicking activity obtained by adjusting surface functional groups. (c) Combination or cascade therapy. PNzyme/ MnO<sub>2</sub> with the ability to both dissolve thrombus and scavenge ROS. Reprinted with permission from Refs. [152,53] and [154]. Copyright 2021 American Chemical Society, 2023 Springer Nature, and 2021 Wiley-VCH.

ischemic brain. When compared with Fe<sub>3</sub>O<sub>4</sub>, the defect-engineered Co-Fe<sub>3</sub>O<sub>4</sub> showcased a 100-fold increase in affinity towards H<sub>2</sub>O<sub>2</sub>. [153] Gao *et al.* prepared CD nanozymes, which exhibited SOD-like activity greater than 10,000 U/mg under appropriate reaction conditions and raw material conditions. The results of surface structure adjustment combined with theoretical calculations show that enhancing the abundance of oxygen-containing functional groups including hydroxyl, carboxyl, and carbonyl on the surface of CD is beneficial for the improvement of SOD-like activity. In addition, CD nanozymes can target the interior of oxidative damage cells and effectively scavenge intracellular ROS. In a mouse model of IS, CD nanozymes can specifically accumulate in damaged brain regions and reduce stroke-induced oxidative damage (Fig. 7b). [53]

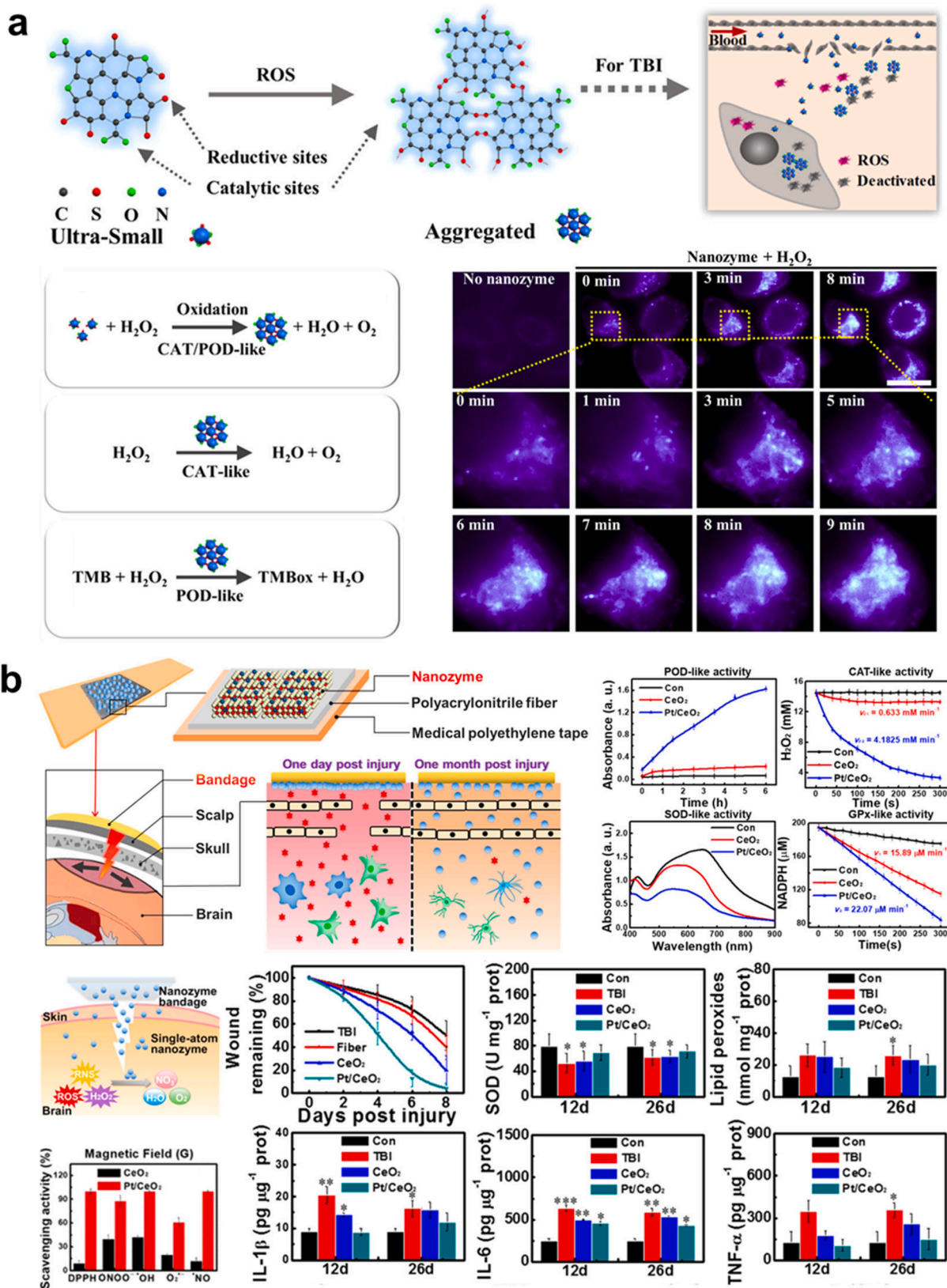
The above-mentioned nanozymes mainly play a neuroprotective role in alleviating the secondary injury caused by massive RONS generated by reperfusion. It is worth mentioning that Wang *et al.* reported innovative peptide-templated manganese dioxide nanozymes (PNzyme/MnO<sub>2</sub>), which have both neuroprotective and thrombolytic abilities. Self-assembling polypeptides contain a variety of functional motifs. The fibrin-binding sequence specifically targets thrombi in ischemic brain regions, and thrombin activates the thrombin-recognized, and cleaved sequence as a functional switch initiates the activity of the thrombolytic sequence, followed by secondary and tertiary targeting (Tf-mediated BBB targeting sequence and apoptotic neuron targeting sequence) to guide the further localization of IS tissue to scavenge ROS. Subsequently, the nanozyme that possesses the cascade catalytic activity of SOD and

CAT can effectively decompose ROS into nontoxic O<sub>2</sub> and H<sub>2</sub>O, thus protecting the organisms from excessive ROS (Fig. 7c). [154]

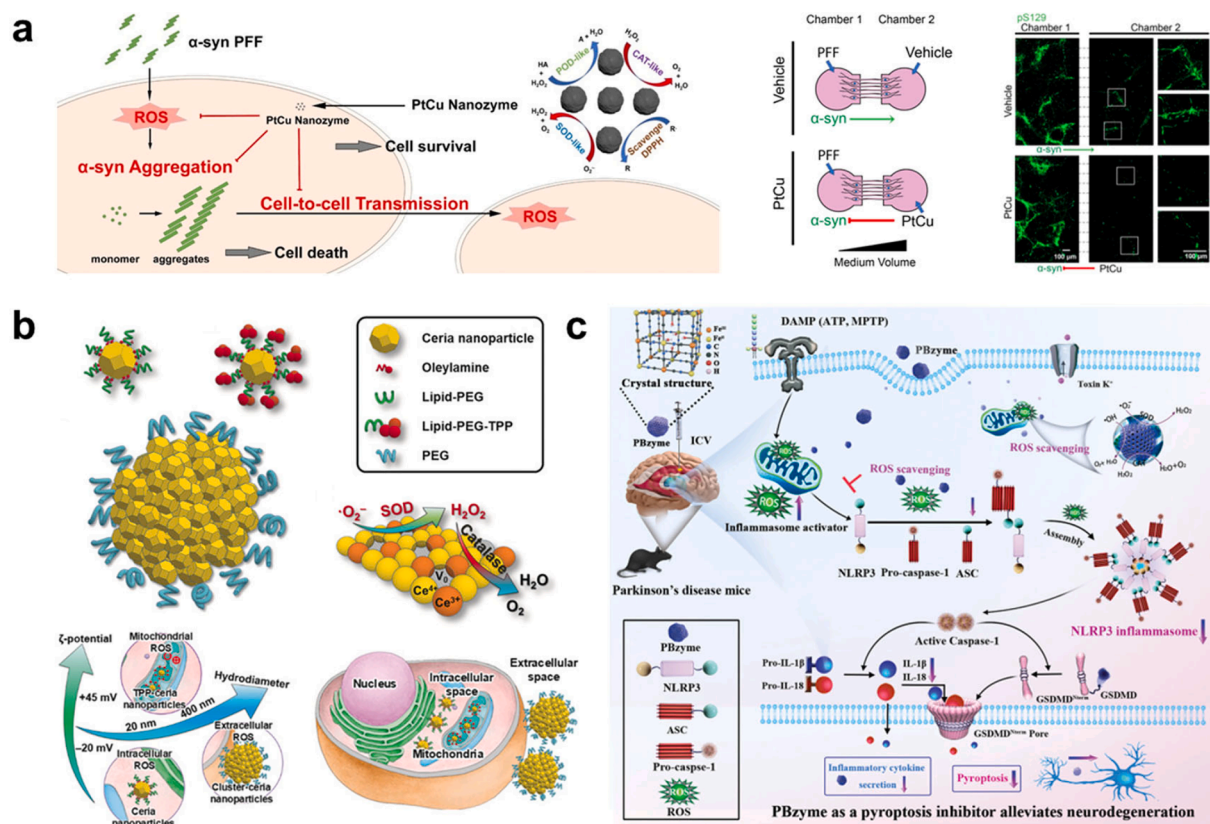
#### 4.2. Traumatic brain injury

TBI refers to temporary or even permanent damage to brain function or structure caused by external mechanical force. The pathophysiology of TBI is divided into two phases, primary and secondary injury. When the brain suffers a primary mechanical injury, neurons and glial cells respond to the injury, triggering various physiological and biochemical processes within the organism, which further leads to lasting secondary damage, including BBB disruption, neuroinflammation, cerebral edema, and ischemia hypoxia. In the complex cascade of secondary damage mechanisms, oxidative and nitrosative stress which arises from the accumulation of RONS is a key factor. [155] Mitochondrial dysfunction leads to O<sub>2</sub> leakage, which generates ROS under the catalysis of NADPH oxidase. In addition, hyperactivation of N-methyl-d-aspartate receptors and Ca<sup>2+</sup> influx lead to the excessive generation of RNS. [156] RONS spread into the surrounding normal brain, oxidizing nucleic acids, proteins, and lipids.

As possessing powerful antioxidant activity, nanozymes have demonstrated encouraging potential for utilization in the treatment of TBI. The excretion of inorganic nanozymes is slow, while nanozymes with larger sizes tend to accumulate non-specifically in healthy organs. Therefore, some researchers have turned to exploring alternative solutions of small organic nanozymes for brain damage. Mu *et al.* developed



**Fig. 8.** Nanozymes for the treatment of TBI. (a) Aggregation of ROS-induced nanozymes for the treatment of TBI. (b) A non-invasive bandage for treating brain trauma utilizing single-atom catalytic nanozymes. Reprinted with permission from Refs. [158] and [159]. Copyright 2019 American Chemical Society.



**Fig. 9.** Nanozymes for the treatment of PD. (a) Inhibition of  $\alpha$ -syn pathology. An overview of PtCu nanozymes scavenging ROS and mitigating the pathological effects induced by  $\alpha$ -syn. (b) ROS scavenging. Schematic illustration depicting ceria NPs scavenge ROS in response to their cellular distribution. (c) Inhibition of neuroinflammation. Mechanism of PBzyme as a pyroptosis antagonist to inhibit neuroinflammation and protect PD neurons. Reprinted with permission from Refs. [165,169] and [170]. Copyright 2021 Elsevier, 2018 and 2022 Wiley-VCH.

ultra-small carbogenic nanozymes. After intravenous injection, nanozymes exhibited efficient elimination of over-produced RONS in acute TBI-induced brain tissue. Unlike the above-mentioned ultrasmall ceria nanozymes that easily aggregate *in vivo*, the ultrasmall carbogenic nanozymes showed effective renal clearance at high doses.[157] He *et al.* successfully synthesized an ultrasmall ( $\sim 3$  nm) organic nanozyme using the microwave-heating method of lysine and GSH. Upon exposure to free radicals, it can gradually aggregate to 75–100 times its original size, and undergo disintegration into smaller entities through the action of NADPH-glutathione reductase (Fig. 8a). This aggregation reversibility was beneficial to promote excretion following the recovery from TBI.[158]

Compared with the previous intravenous route of nanozyme, the nanozyme-based wound dressing provides a new non-invasive treatment for TBI. Traditional bandages are mainly used to relieve pain, pressurize to stop bleeding, and protect wounds from environmental influences. However, after long-term exposure at room temperature, the antibiotics, antioxidants, and enzymatic debriding agents on the surface of the bandage are prone to failure, and it is challenging to promote wound healing. Yan *et al.* ingeniously devised a non-invasive treatment approach for trauma by developing a single-atom Pt/CeO<sub>2</sub>-rich bandage. Dispersed Pt single atoms enhanced the multienzyme activity of CeO<sub>2</sub> clusters by 3–10 times and the RONS-scavenging activity by 2–10 times because Pt single atoms with oxygen vacancies can provide active catalytic sites. Experiments showed that the bandage can reduce oxidative stress and inflammatory response indicators. Compared with traditional intravenous administration for brain injury, Pt/CeO<sub>2</sub> nanozyme-based bandages seem to show great potential due to their non-toxic and sustained multi-catalytic processes (Fig. 8b).[159] Subsequently, the research team kept this idea and further designed an enhanced catalytic

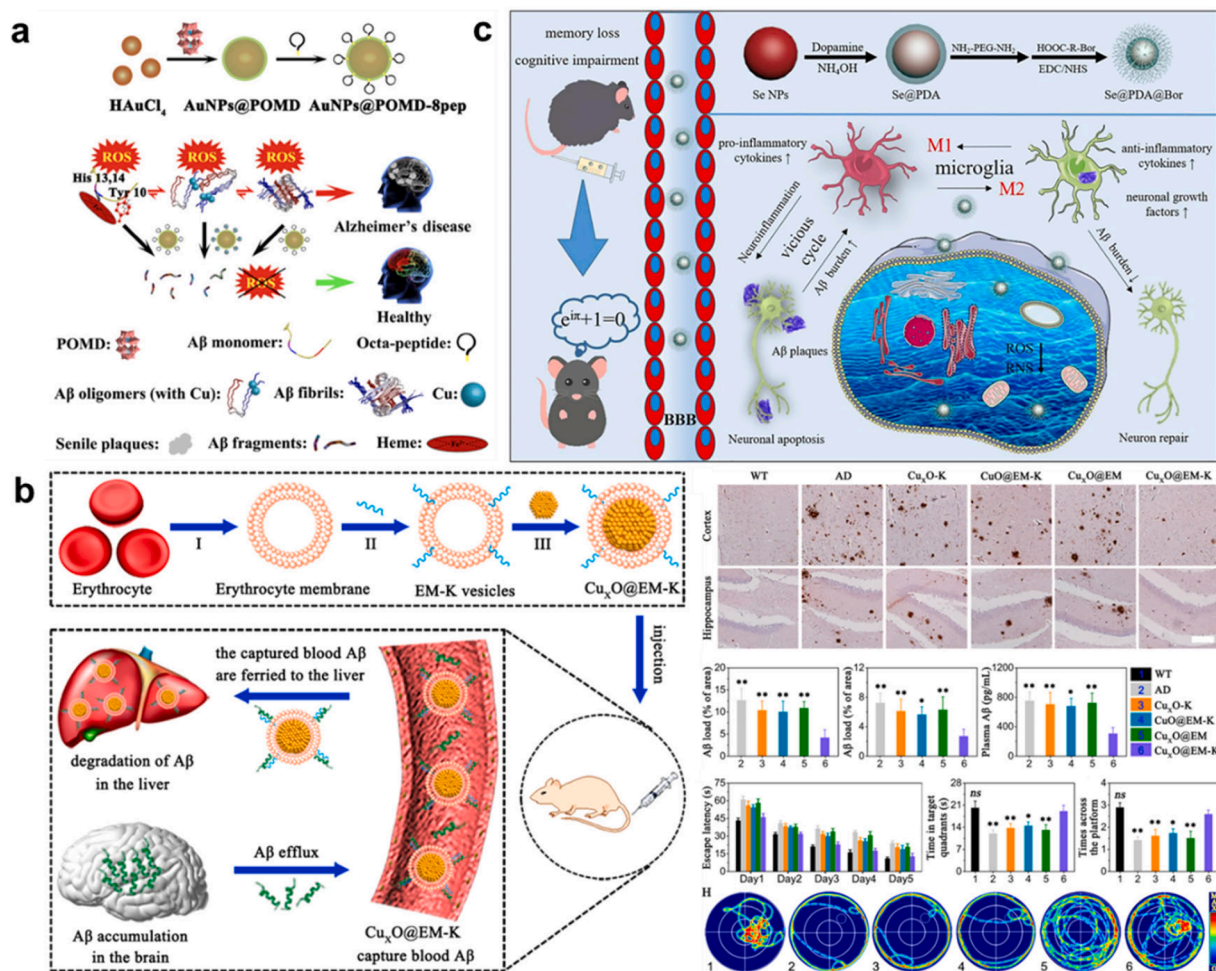
patch utilizing Cr-doped CeO<sub>2</sub> nanozymes.[160] Compared with the aforementioned research, the preparation procedure of nanozyme was changed from a hydrothermal process to a coprecipitation process. Among CeO<sub>2</sub> nanozymes doped with various metallic elements, the Cr/CeO<sub>2</sub> nanozymes showed the best catalytic activity.

The catalytic process *in vivo* is complex, so nanozymes with physiological environment preference are particularly important for treating TBI. Nonetheless, the majority of nanozymes exhibit suboptimal catalytic activity in the neutral physiological environment. The aforementioned Pt/CeO<sub>2</sub> nanozyme has an acidic environment preference, exhibiting POD-mimicking activity in an acidic environment while CAT-mimicking activity in neutral and alkaline environments. Mu *et al.* demonstrated a trimetallic nanozyme that exhibited optimal catalytic performance in a physiological environment attributed to the lattice distortion and exposure of catalytic centers. At the same time, the multi-enzyme mimetic activity endowed it with multiple antioxidant properties, which can improve mice's post-brain injury survival percentage and significantly alleviate neuroinflammation.[161]

### 4.3. Neurodegenerative disease

#### 4.3.1. Parkinson's disease

PD, ranking as the second most prevalent neurodegenerative disease, presents with abnormal motor symptoms (tremor, motor retardation, stiffness, abnormal gait, and postural instability) and a series of non-motor complications (cognitive, psychiatric, sleep, and sensory disturbances).[162] PD involves different types of neurons populations distributed across multiple brain regions, the most important one being the DA neurons in the Substantia Nigra compact (SNc). The  $\alpha$ -syn, a characteristic constituent of the Lewy body in DA neurons, is misfolded



**Fig. 10.** Nanozymes for the treatment of AD. (a) Simultaneous consideration of multiple factors including A $\beta$ , tau, and ROS. AuNPs@POMD-8pep exhibits dual functionality, being proficient in both depleting A $\beta$  aggregates and scavenging ROS. (b) Targeted clearance of A $\beta$  from peripheral organs. Cu<sub>x</sub>O@EM-K efficiently captures A $\beta$  present in the bloodstream, leading to a reduction in brain A $\beta$  burden. (c) Ameliorating neuroinflammation. Se@PDA@Bor promotes the conversion of microglia from M1 to M2 by scavenging RONS, thus reducing neuroinflammation. Reprinted with permission from Refs. [175,176] and [178]. Copyright 2016 Springer Nature, 2020 American Chemical Society, and 2021 Elsevier.

and accumulated, which produces toxicity and promotes the gradual death of dopaminergic neurons, thereby causing dysfunction.[163] In addition to the abnormal accumulation of  $\alpha$ -syn, the degeneration of DA neurons is heavily influenced by oxidative stress, which arises from an overproduction of ROS. Oxidative stress, in which mitochondrial dysfunction participates, leads to the accumulation of oxidized DA, ultimately leading to a decline in glucocerebrosidase activity and lysosomal dysfunction, causing neuronal degeneration.[164] As a pathogenic factor of prion-like spread, oxidative stress also leads to the inevitable spread of prion protein  $\alpha$ -syn. [165] Additionally, oxidative stress can be mediated by neuroinflammation and microglia activation. In the inflammatory environment, arachidonic acid generates ROS under the action of lipid oxidase. Microglia can be activated by increased nitric oxide synthase-induced up-regulation of major histocompatibility complexes and glycoproteins, and expression of related inflammatory factors (tumor necrosis factor- $\alpha$ , interleukin-1 $\beta$  and -6) and molecules (adhesion molecules, cyclooxygenase-2, and  $\bullet$ NO), which exacerbate the irreversible damage to DA neurons in SNc.[166]

As mentioned above,  $\alpha$ -syn accumulation contributes to the progressive degeneration and eventual demise of DA neurons. Liu *et al.* found that the anti-oxidation nanozyme PtCu nano-alloys (NAs) significantly inhibited the diffusion of  $\alpha$ -syn modeled by an intrastriatal injection of pre-made fibers. In addition, PtCu NAs significantly inhibited  $\alpha$ -syn pathology, neurotoxicity, neuronal diffusion *in vitro*, and striatal to

substantia nigra diffusion *in vivo* by scavenging ROS in primary neuron culture (Fig. 9a).[165] This study showed that antioxidant nanozymes may be used to combat the pathological diffusion of  $\alpha$ -syn in PD. But unknown factors such as the biological safety of metal NPs in the brain, the permeability of BBB, and the long-term effect still need to be explored. Given the close association between PD and ROS, effective removal of ROS can protect tissues from OS, which contributes to the treatment of PD. The Cu<sub>x</sub>O nanoparticle clusters (NCs) which exhibit analogical properties to CAT, GPx, and SOD studied by Hao *et al.* scavenged ROS and protected PD cells from neurotoxicity induced by oxidative stress. The nanozyme also showed good biocompatibility and therapeutic effect on 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-induced PD mice with oxidative stress-mediated neurological disorders.[167] This research may provide an opportunity for applying inorganic nanostructures with enzyme-like activity in biomedical and other bioengineering fields. ROS at different positions in cells plays different roles in diseases, so it is very necessary to develop cell-based localization nanozymes for ROS removal. To this end, Li *et al.* engineered lactoferrin-modified Au-Bi<sub>2</sub>Se<sub>3</sub> nanodots located near the mitochondria, which significantly scavenge ROS in 1-methyl-4-phenylpyridinium<sup>+</sup> cells, thereby maintaining the normal mitochondrial membrane potential, and also showed a strong BBB penetration ability. [168] Ceria NPs designed and synthesized by Kwon *et al.* utilized regenerative redox switching between Ce<sup>3+</sup> and Ce<sup>4+</sup> ions to selectively

remove ROS. This study showed that the clearance of intracellular ROS through ceria NPs or the clearance of mitochondrial ROS through TPP conjugated-ceria NPs are able to effectively inhibit neuroinflammation and lipid peroxidation. Furthermore, this approach helped maintain the tyrosine hydroxylase (TH) level in the mouse striatum. However, the use of cluster-ceria NPs for the removal of extracellular ROS showed no therapeutic effect on maintaining TH level and inhibiting lipid peroxidation, which suggested reducing intracellular and mitochondrial rather than extracellular oxidative stress is essential for the treatment of PD (Fig. 9b). [169].

Neuroinflammation is also a detrimental factor in PD. Intraventricular administration of Prussian Blue nanozyme (PBzyme) in a PD mouse model can reduce DA degeneration and inhibit neuroinflammation. By scavenging ROS, PBzyme inhibited the activation of NLR family pyrin domain containing 3 (NLRP3) inflammasomes, down-regulating the cleavage of gasdermin D and the production of inflammatory factors, inhibiting microglia pyroptosis ultimately (Fig. 9c). [170] This study provided helpful insight into the mechanism of action and therapeutic strategies for treating PD through the use of pyroptosis inhibitors. Through adding an anti-inflammatory small molecule compound (quercetin), Li *et al.* made up for the shortcomings of inorganic nanozymes (Ptzyme) in alleviating the brain's inflammatory microenvironment. [171].

#### 4.3.2. Alzheimer's disease

Nowadays, at least 50 million people worldwide suffer from AD, which is the most common among old people. Features of AD are the progressive decline of memory, cognitive, and behavioral functions. The brain of AD patients has moderate cortical atrophy, most notably of the multimodal limbic lobe structures and association cortices. [172] The accumulation of A $\beta$  induces microglia polarization and activates toll-like receptors, leading to the release of multitudinous pro-inflammatory cytokines. A $\beta$  overload also leads to the destruction of lysosomal degradation defects of A $\beta$  fragment, triggering the release of cathepsin B, and prompting the assembly of NLRP3 inflammasome to amplify the inflammatory response. [173] In addition, tau that is involved in axonal transport of organelles is hyperphosphorylated and aggregated to form neurofibrillary tangles, [145] resulting in mitochondrial dysfunction and oxidative stress. Mitochondrial morphology is altered in the brain of AD subjects, with decreased activity of enzymes involved in oxidative phosphorylation. [174]

Since factors such as A $\beta$ , tau, and ROS have intrinsic connections in the pathogenesis of AD, optimal therapeutic approaches necessitate simultaneous consideration of multiple factors. Gao *et al.* reported a reasonable design of nanozymes based on AuNPs. The aggregation of A $\beta$  was impeded through the interaction between polyoxometalate (POM) featuring Wells-Dawson structure (POMD) and the cationic domain of His13 to Lys16. AuNPs easily grafted POMD onto its surface and also showed excellent ability in redox performance and BBB penetration. This nanozyme not only had protease-mimicking activity to consume A $\beta$  aggregates, but also had SOD-mimicking activity to remove ROS, and exhibited potential for Cu removal as a metal chelator (Fig. 10a). [175] This study presented a compelling approach for the multifunctional treatment of AD by specifically modifying nanozymes. Guan *et al.* engineered a synthetic nanozyme, namely Ceria/Polymerases Hybrid (CeONP@POMs), which encompassed both proteolytic and SOD-mimicking activities. The findings showed that CeONP@POMs effectively hydrolyzed A $\beta$ 40 and reduced intracellular ROS. CeONP@POMs also crossed BBB, regulated microglia, and protected neurons from A $\beta$ -related cytotoxicity *in vivo*. [31] Furthermore, recognizing the crucial involvement of peripheral organs in A $\beta$  clearance within the brain, Ma *et al.* used Cu $_x$ O nanozyme embedded into XTg-AD mouse erythrocyte membrane, incorporating the A $\beta$ -targeted pentapeptide KLVFF3 to prepare Cu $_x$ O@EM-K. The surface layer of the erythrocyte membrane can prevent the formation of protein corona, so as to retain its ability to target and clear A $\beta$  in biological fluids and achieve the effect of rapidly

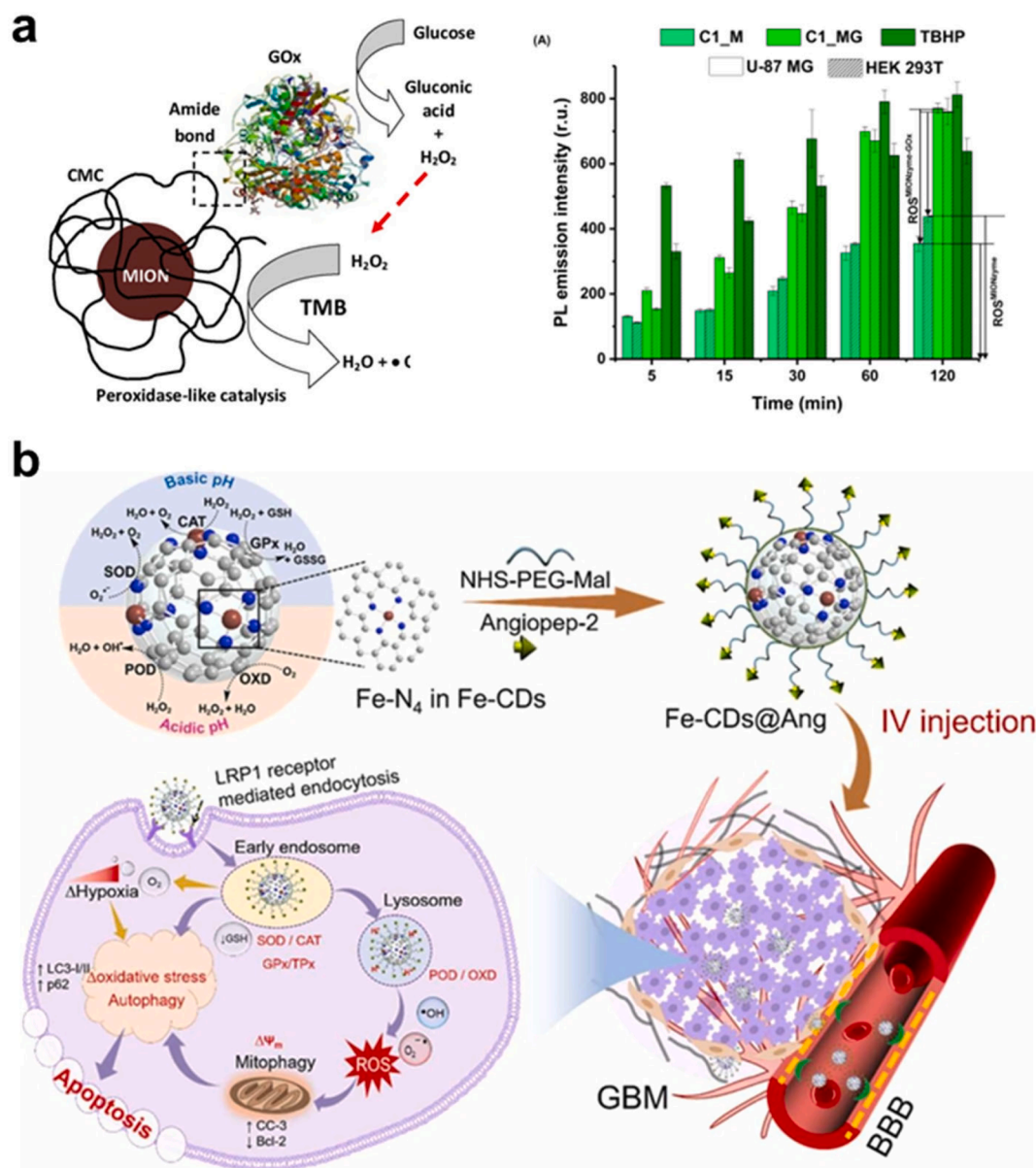
reducing the A $\beta$  level in the brain. The Cu $_x$ O also had a variety of antioxidant enzyme-mimicking activities, which can effectively inhibit the oxidative damage of cell membrane induced by A $\beta$  and stabilize the cell membrane (Fig. 10b). [176] This research innovatively transferred the action site of the nanozyme, presenting a valuable thought for improving the targeting of nanozymes. Due to tau hyperphosphorylation and tau aggregates playing an essential role in AD, Chen *et al.* rationally assembled mesoporous silica nanoparticles (MSN) with ceria and iron oxide nanocrystallines, and subsequently modified nanoassembly with Ga labeled Amino-T807 and loaded them with methylene blue MB to fabricate a hyperphosphorylated tau-targeted multifunctional nanocomposite. Ceria nanocrystals with SOD-like activity effectively eliminate excessive ROS, and thus inhibit tau hyperphosphorylation through alleviating the oxidative stress. At the same time, loaded MB can restrain the aggregation of hyperphosphorylated tau, thereby inhibiting neuronal apoptosis. [177]

Neuroinflammation assumes a significant part in the pathological process of AD as well. Gong *et al.* developed a Selenium-polydopamine nanozyme (Se@PDA@Bor) showing remarkable curative effect on AD. Studies have shown that the overproduction of pro-inflammatory cytokine fosters A $\beta$  generation and additionally stimulates microglia proliferation and activation, resulting in a mutual cycle of A $\beta$  aggregation and neuroinflammation. Se is an important component of selenoprotein systems, including glutathione and thioredoxin, which exhibit the capability to effectively scavenge ROS. Se@PDA@Bor exerted inhibitory effects on A $\beta$  accumulation, preventing neuroinflammation reactivation, and breaking the vicious circle. Moreover, borneol (Bor) packaging made it easier to pass through BBB. Se@PDA@Bor restored its capacity to engulf A $\beta$  and promote neuronal restoration by activating microglia from M1 to M2 phenotype. Experiments in APP/PS1 transgenic AD mice showed that nanozymes effectively regulated the neuroprotective capabilities of astrocytes and microglia within a brief timeframe. This intervention led to improvements in neuroinflammation, reduction of A $\beta$  accumulation, as well as amelioration of memory decline and cognitive disorders (Fig. 10c). [178] Compared with the traditional single-target nanozyme, better targeting and biocompatibility can be achieved by the modified nanozyme. In this way, nanozymes offer notable advantages for treating AD.

#### 4.3.3. Huntington's disease

HD is a dominant genetic disease which occurs on the autosome. Patients with HD initially exhibit changes in personality, mood, and mental status, sometimes accompanied by decreases in cognitive ability, followed by involuntary dance movements, bradykinesia, dystonia, stiffness, and dementia. Histone acetylation is widely recognized as a pivotal mechanism influencing cognitive functions. In HD, the aggregation of mutated huntingtin (HTT) proteins may decrease histone acetylation degree, resulting in neuronal damage and loss. [179] Oxidative stress also represents a significant contributor to the HD pathogenesis. Prolonged exposure to oxidative stress can damage DNA repair system, destroy cell structure, and cause mitochondrial dysfunction. The mutated HTT protein may inhibit the adaptive transcription process of oxidative stress. [180]

As mentioned, reducing intracellular aggregation of mutant HTT proteins would be a promising approach for treating HD. In the research carried out by Cong *et al.*, the efficacy of Se NPs in the treatment of HD based on the transgenic HA759 *C. elegans* HD model. 2  $\mu$ M Se NPs significantly reduced ASH neuron death, axonal degeneration, and behavioral impairments. Furthermore, Se NPs provided protective effects against stress in *C. elegans*, thereby effectively restoring the normal response of *C. elegans* to external stimuli. As mentioned above, Se NPs that have strong antioxidant activity can reduce the ROS levels in HD nematodes. Its surface was easy to form protein coronas, which can inhibit the aggregation of HTT proteins in nematodes. In addition, down-regulating the expressions of histone deacetylase (HDAC)1, HDAC4, and sirtuin 1 can inhibit HD pathology in model organisms. The



**Fig. 11.** Nanozymes for the treatment of brain cancer. (a) Hybrid nanostructure based on MIONzyme-GOx effectively mediates the accumulation of ROS by the cascade reactions of GOx enzyme and POD-like activity of nanozyme, and efficiently induces brain cancer cell death. (b) ROS-regulated enzyme-linked reaction induced by angiopep-2-functionalized Fe-CDs nanozymes to induce autophagy-lysosome pathway to treat GBM. Reprinted with permission from Refs. [187] and [29]. Copyright 2022 Elsevier.

mRNA levels of wild-type and HA759 nematodes treated with Se NPs were compared with those of the untreated group. After Se NPs treatment in HA759 nematodes, a significant decrease was observed in the expression levels of the associated genes including *sir2.1*, *hda-1* and *hda-4*. However, the mRNA levels of *hda-2*, *3*, *6*, *10* remained unchanged. [181] This study found that Se NPs protected and repaired neurological function under stress conditions, which may be conducive to the treatment of HD disease and has guiding significance for the rational design of Se NPs. However, since not all mRNA levels of HDAC members were decreased after treatment with Se NPs, we need to strengthen the epigenetic research on the effect of Se NPs on HDAC expression. This study also reminds us that the applied doses of other Se compounds should be considered when applying the nanodrug to treat HD, so as to ensure the safety of drug administration.

#### 4.3.4. Autosomal recessive spastic ataxia of Charlevoix-Saguenay

ARSACS is an uncommon inherited neurodegenerative disorder that

is primarily attributed to mutations in the *SACS* gene. [182] Sacsin encoded by the *SACS* gene is a protein on the surface of mitochondria, and its functional destruction makes the mitochondria show high perfusion balloon-like, thus impairing mitochondrial function. In the sacsin-knockout mouse model, mitochondria are accumulated in the cell body and proximal dendrites of the sacsin-knockout neurons, which may reduce mitochondrial transport to the dendrites, resulting in abnormal dendritic morphology and eventually neuronal cell death. [183] *SACS* mutations exacerbate oxidative stress and affect bioenergy metabolism. Consequently, this may amplify bioenergy deficits, establish a detrimental circle, and possibly promote cell death during neurodegenerative processes. Increased ROS production leads to mitochondrial bioenergy dysfunction, which is common in many neurodegenerative diseases. [184] Therefore, antioxidants have a great prospect in the treatment of ARSACS.

Polydopamine nanoparticles (PDNPs) contain abundant functional groups including imines and catechins, and they confer upon PDNPs the

capability to effectively clear substantial quantities of ROS. To evaluate the antioxidant properties of PDNPs, Battaglini *et al.* treated fibroblasts from healthy individuals and ARSACS patients with 100  $\mu\text{g}/\text{mL}$  PDNPs. The results revealed that PDNPs significantly reduced basal ROS levels of these two types of cells, and were more potent than other antioxidant compounds, including nanostructured lipid carriers, tannic acid, and benzophenone. Apoptotic cells and necrotic cells increased significantly after treatment with 2.5 mM *tert*-butyl hydroperoxide solution (TBH), and decreased the number of living cells, while PDNPs treatment improved this situation. PDNPs also had a protective effect on mitochondria by recovering the mitochondrial elongation loss caused by TBH treatment. In the environment of 2.5 mM TBH and 6 mM oligomycin (a respiratory chain inhibitor), PDNPs played a role in preventing mitochondrial membrane potential loss. Furthermore, the PDNPs also showed good BBB permeability in the cross-porous BBB model which is established through the coculture of endotheliocytes and astrocytes. [185] This study demonstrated the outstanding antioxidant and mitochondrial protection capabilities of PDNPs, providing a promising basis for the utilization of PDNPs as a prospective therapeutic approach to ARSACS. However, further studies *in vivo* are needed to analyze the signaling pathways affected by PDNPs, which would be imperative for advancing the clinical applications of PDNPs.

#### 4.4. Other diseases

##### 4.4.1. Brain cancer

Brain cancer is widely recognized as one of the most lethal diseases affecting the nervous system. Among brain cancers, glioblastoma multiforme (GBM) continues to prevail as the most prevalent and deadly form. In addition, brain metastases are the most common neurological complication of systemic cancers, with a significantly higher prevalence than primary brain tumors. Traditional cancer treatments, including surgical resection, radiotherapy, and chemotherapy, have shown the disadvantages, such as poor prognosis. [186] However, the intracellular pathways and metabolic patterns of tumor cells are quite different from those of healthy cells, including local high concentration of  $\text{H}_2\text{O}_2$ , hypoxia, uneven blood perfusion, and extracellular acidic pH caused by glycolytic metabolism of cancer cells, etc., which offer a multitude of prospects for developing prospective anticancer therapies and minimize the adverse reactions on healthy cells. [187] It is noteworthy that in addition to antioxidant nanozymes mitigating the development of most brain diseases by consuming excessive RONS, pro-oxidative nanozymes with OXD and POD-like activities also hold significant therapeutic value, particularly in brain tumor therapy. [188]

On the one hand, chemodynamic therapy (CDT) mediated by POD-like active nanozymes converts endogenous  $\text{H}_2\text{O}_2$  into highly harmful  $\cdot\text{OH}$  through Fenton or Fenton-like reactions, thus capable of directly inducing massive cell death. However, in the TME, the concentration of  $\text{H}_2\text{O}_2$  is insufficient to sustain  $\cdot\text{OH}$  production. [189] Therefore, enhancing the local concentration of  $\text{H}_2\text{O}_2$  within TME is needed to accelerate the Fenton or Fenton-like reactions, thereby improving the efficacy of CDT. Mansur *et al.* designed and fabricated a hybrid nanostructure composed of superparamagnetic iron oxide. This innovative nanostructure was subjected to functionalization and stabilization using carboxymethyl cellulose biopolymer. Additionally, it underwent a covalent bioconjugation process with GOx, resulting in the formation of MIONzyme-GOx. MIONzyme-GOx consumed glucose nutrients through GOx-catalyzed biological reaction and generated  $\cdot\text{OH}$  through  $\text{H}_2\text{O}_2$  catalyzed by a Fenton-like reaction under the acidic pH of TME, leading to cell death. Compared with “pure” MIONzyme, MIONzyme-GOx had higher toxicity to glioblastoma cells (Fig. 11a). [187] Besides, Mansur *et al.* synthesized Au NPs stabilized by trisodium citrate (AuNPs@TSC) to mimic OXD. Additionally, they coupled cobalt-doped superparamagnetic iron oxide NPs stabilized by carboxymethylcellulose ligands (Co-MION@CMC) to mimic POD. Here, TSC mimicked GOx, catalyzing the production of  $\text{H}_2\text{O}_2$  to accelerate the Fenton reaction.

$\text{Co}^{2+}$  contributed to an elevation in the magnetocrystalline anisotropy. Co-MION@CMC can improve the thermal therapy performance of magnetite NPs, which can also be used for magnetic thermal therapy. The nanosystem was connected with an iRGD sequence (9-amino acid cyclic peptide, sequence: CRGDKGPDC) as a targeting peptide to recognize  $\alpha\text{v}\beta 3$  integrin which is often overexpressed in brain tumor cell membranes to positively target GBM cancer cells. [190] In this study, the double inorganic nanozymes system was used to compensate for the instability of natural enzymes, which induced biocatalytic cascade reactions in TME, and achieved the combined magnetothermal-chemical therapy, providing an effective strategy for the design of hybrid nanozyme systems. However, further studies *in vivo* are necessary to design the features of nano-systems to improve the biodistribution and clearance/excretion of nano-preparations, making nanozymes beneficial for the clinical treatment of cancer.

On the other hand, apart from stand-alone applications, nanozymes can also improve TME, enabling their synergistic application alongside other treatment methods such as chemotherapy, radiotherapy, photothermal therapy (PTT), photodynamic therapy (PDT), sonodynamic therapy, and immunotherapy. This combination serves to enhance therapeutic efficacy while mitigating adverse reactions. For example, malignant GBM cells that are resistant to chemotherapy exhibit increased vulnerability to autophagic cell death pathways. [191] The CDs synthesized by Muhammad *et al.* supported iron monatomic nanozyme (Fe-CDs) to show multiple enzyme activities of OXD, POD, CAT, SOD, GPx, and thiol peroxidase (TPx). To effectively target low-density lipoprotein receptor-related protein-1, which is highly expressed in brain capillary ECs and tumor cells, the Fe-CDs underwent chemical modification by Angiopep-2 (Fe-CDs@Ang). Fe-CDs@Ang which exhibited inherent OXD/POD-like activity accumulated within the acidic *endo*-lysosomes (pH 4–5), impairing lysosomal degradation capacity and inducing autophagy, thereby overcoming drug resistance in solid GBM. Furthermore, SOD, CAT, and GPx-like activities can enhance autophagy and lysosome-based apoptosis, and alleviate hypoxia (Fig. 11b). [29] Yin *et al.* developed a  $\text{Gd}_2\text{O}_3/\text{Ir}/\text{TMB-RVG29}$  (G@IT-R) nanomachine that can achieve tumor-specific PTT. Under the activation of acidic TMB and high concentration of  $\text{H}_2\text{O}_2$ , Ir nanozyme exhibited POD-like activity, enhancing the color reaction of the photothermal agent TMB, ultimately enabling tumor-specific PTT. [27] In addition to the above-mentioned nanozymes synergizing a single therapy, synergizing two or more complex therapeutic systems is also a current research trend. Sunil *et al.* reported nanozymes (BIONs) with photodynamic and immunological activities. The  $\text{CeO}_2$  nanozyme exhibited a pro-oxidative effect in tumor sites to enhance the efficacy of PDT, while exerting an antioxidant effect in healthy tissues like the kidneys. [192]

##### 4.4.2. Cerebral malaria

Malaria is a parasitic disorder resulting from malaria parasites transmitted by mosquitoes, among which *P. falciparum* can cause CM, a life-threatening neurological complication. The parasitized red blood cell expresses *Plasmodium falciparum* erythrocyte membrane protein-1 on the surface, enabling its interaction with membrane proteins of ECs including intercellular adhesion molecule-1. ECs phagocytize merozoites and presents malaria antigen to  $\text{CD8}^+$  T cells. This interaction will trigger BBB destruction mediated by interferon- $\gamma$  and perforin, [193] causing fluid and protein to leak from the circulatory system to the intercellular matrix, thereby forming angioedema. [194] A research report of live mice utilizing whole brain covered high-field MRI showed that inflammatory destruction and vasogenic edema of the BBB began with the olfactory bulb and then spread deep into the brain along a distinct pathway known as the rostral migratory flow. Ultimately, this pathological process extended toward the brain stem, leading to coma and death. [195] The release of free heme from parasitized red blood cells (RBCs) may generate excessive ROS and damage ECs and BBB. [196]

*Plasmodium* infection may eventually result in the inflammatory

Table 4

A summary of nanozymes for brain disease treatment.

Brain Diseases	Nanozymes	Activities	Mechanisms	References
Ischemic Stroke	HPBZs	CAT, POD, SOD	ROS scavenging, inhibition of apoptosis, anti-inflammatory	[138]
	CeO <sub>2</sub>	CAT, SOD	ROS scavenging, inhibition of apoptosis	[146]
	EA/P-CeO <sub>2</sub>	CAT, SOD	ROS scavenging, BBB protection	[147]
	CeO <sub>2</sub> @ZIF-8 NPs	CAT, POD	ROS scavenging, inhibiting lipid peroxidation, reducing the oxidative damage and death of neurons, anti-inflammatory	[148]
	PEG-Fe <sub>3</sub> O <sub>4</sub>	CAT, POD, SOD	ROS scavenging, facilitating BBB reconstruction	[150]
	MeNPs	SOD	multi-antioxidative, anti-inflammatory	[151]
	Eda-MnO <sub>2</sub> @Tf	CAT, SOD	ROS scavenging, anti-inflammatory	[30]
	Co-Fe <sub>3</sub> O <sub>4</sub>	CAT, POD	RONS scavenging	[153]
	MPBzyme @NCM	CAT, SOD	ROS scavenging, enhanced differentiation of microglia polarization from M1 to M2, reducing recruitment of neutrophils, inhibition of neuronal apoptosis, increasing the proliferation of neurocytes	[152]
	Traumatic Brain Injury	Carbon dot	SOD	intracellular excess ROS scavenging
PNzyme/MnO <sub>2</sub>		CAT, SOD	thrombolysis multilevel, precise targeting, ROS scavenging, anti-inflammatory	[154]
O-NZ		SOD, GPx	RONS scavenging, improving neurocognition and memory by up-regulation of heme oxygenase-1	[158]
Carbogenic nanozyme		CAT	ROS scavenging, immune suppression	[157]
TriM nanozyme		CAT, POD	RONS scavenging	[161]
Parkinson's Disease	Pt/CeO <sub>2</sub>	CAT, GPx, POD, SOD	RONS scavenging, anti-neuroinflammation	[159]
	Cr/CeO <sub>2</sub>	CAT, GPx, SOD	RONS scavenging, anti-neuroinflammation	[160]
	PtCu NAs	POD, CAT, SOD	inhibiting $\alpha$ -syn pathology and spreading, reducing cell apoptosis and interneuronal transmission	[165]
	Cu <sub>x</sub> O NCs	CAT, GPx, SOD	ROS scavenging	[167]
	Lf-Au-Bi <sub>2</sub> Se <sub>3</sub> NDs	POD, SOD, CAT, GPx	protecting the mitochondria from oxidative stress, improvement of behavioral performance and neuronal damage	[168]
	Ceria/TPP-Ceria NPs	SOD, CAT	intracellular or mitochondrial ROS scavenging, inhibition of neuroinflammation and lipid peroxidation	[169]
	PBzyme	CAT, POD, SOD	reducing dopamine degeneration, anti-neuroinflammation	[170]
Alzheimer's Disease	Que/Ptzyme@Man-PLGA	SOD, CAT	ROS scavenging, anti-neuroinflammation, promoting microglial polarization toward an anti-inflammatory M2 phenotype	[171]
	AuNPs@POMD-8pep	Protease, SOD	A $\beta$ aggregates depleting, A $\beta$ -mediated ROS scavenging, removing Cu from Cu-induced A $\beta$ oligomers	[175]
	CeONP@POMs	Protease, SOD	A $\beta$ aggregates depleting, intracellular ROS scavenging	[31]
	Cu <sub>x</sub> O@EM-K	CAT, SOD, GPx	peripheral A $\beta$ aggregates depleting	[176]
	CeNC/IONC/MSN-T807-MB	SOD	ROS scavenging, inhibition of tau hyperphosphorylation and aggregation	[177]
Huntington's Disease	Se@PDA@Bor	SOD, CAT	RONS scavenging, restoration of mitochondrial homeostasis	[178]
	Se nanoparticles		ROS scavenging, suppression of proteins aggregation, down-regulating mRNA levels of HDAC family members	[181]
ARSACS	PDNPs		recovering the mitochondrial elongation loss, intracellular ROS scavenging	[187]
Brain Cancer	MIONzyme-GOx	POD, GOx	direct destruction of brain cancer cells	[191]
	AuNP@TSC-Co-MION@CMC	OXD, POD	direct destruction of brain cancer cells	[193]
	Fe-CDs@Ang	POD, OXD, SOD, CAT, TPx GPx	regressing the GME tumor, overcoming the drug resistance	[194]
	Gd <sub>2</sub> O <sub>3</sub> @Ir/TMB-RVG29	POD	tumor specific PTT	[27]
Cerebral Malaria	BIONs		enhancing PDT curative effect, initiation of adaptive immune response	[192]
	Fenozyme	CAT, POD	ROS scavenging, anti-inflammatory	[18]
Major Depression Disorder	CeO <sub>2</sub> @BSA	CAT, SOD	ROS scavenging, improvement of depression-like behaviors, anti-neuroinflammation	[200]

destruction of BBB, and the excessive ROS. Therefore, the treatment of CM by improving BBB damage and eliminating ROS has a significant effect. Zhao *et al.* developed a novel type of ferritin nanozyme (Fenozyme), which was constructed from recombinant human ferritin (HF<sub>n</sub>) shell that selectively targeted the BBB ECs, and the internal Fe<sub>3</sub>O<sub>4</sub> nanozyme core that had CAT-like activity to achieve ROS clearance. The results showed that Fenozyme significantly increased the activity score of experimental cerebral malaria (ECM) mice. By combining HF<sub>n</sub> receptors, Fenozyme was targeted to BBB ECs, therefore protecting the BBB barrier's integrity by scavenging ROS. Treatment with Fenozyme or HF<sub>n</sub> protein can reduce the RBCs percentage infected by parasites. Moreover, Fenozyme and HF<sub>n</sub> resulted in a substantial elevation in the population of macrophages within the liver of ECM mice, notably stimulated the proliferation of macrophage Raw264.7, and enhanced the phagocytosis of bone marrow-derived macrophages to pRBCs<sup>GFP</sup> and nRBCs<sup>GFP</sup>. This result showed that Fenozyme alleviated parasitemia by

enhancing the proliferation and phagocytosis of macrophages. In addition, Fenozyme also effectively alleviated neurological sequelae in ECM mice treated with artemether.[25] This study has provided a strong theoretical basis for strengthening the targeting of nanozyme therapy and its joint application with other drugs, as well as valuable ideas for studying how to treat the injuries of other tissues in CM.

#### 4.4.3. Depression disorder

Depression disorder is an emotional dysfunction caused by genetic system abnormalities or acquired environmental changes. Although current treatment options encompass medication, psychotherapy, and physical therapy, their efficacy remains constrained. The pathology of depression disorder is complex and diverse, involving genetic and psychosocial stress, stress hormones and cytokines, monoamine synthesis, glutamate, and aminobutyric acid neurotransmission.[197] Studies have shown that excessive ROS accumulation stands out as a prominent

factor contributing to the pathological characteristics of depression.[18] In the depressed state, the imbalance of redox homeostasis of cells aggravates the depressive behavior, reduces the ability of neurogenesis in the hippocampus, increases the loss of astrocytes, and leads to brain dysfunction and abnormal signal transmission process of neurons.[198]  $O_2^{\bullet-}$  in the mouse brain peroxisome increases during oxidative stress, leading to the inactivation of CAT. This inactivation results in elevated intracellular  $H_2O_2$  levels increases and tryptophan hydroxylase-2 (TPH2) is further oxidized. A decrease in that level of TPH2 leads to dysfunction of the 5-hydroxytryptamine system in the brain, ultimately contributing to depression disorder.[199]

Fu *et al.* employed the bovine serum albumin (BSA) culture method to synthesize a new antidepressant nanodrug  $CeO_2@BSA$ , which showed significant scavenging ability to  $O_2^{\bullet-}$  and  $\cdot OH$  at 0.025  $\mu g/mL$  and 2.5  $\mu g/mL$  Ce concentration respectively, and significantly reduced the proportion of  $ROS^+$  cells. In contrast to the group treated with PBS, the administration of  $CeO_2@BSA$  nanocluster to mice led to a significant reduction in ROS levels of the mouse brain.  $CeO_2@BSA$  therapy restrained the increase of  $IBA1^+$  activated microglia induced by chronic restraint stress, saved the down-regulation of a brain-derived neurotrophic factor, and reversed the down-regulation of protein synaptophysin and PSD95, which indicated that  $CeO_2@BSA$  had anti-inflammatory and neuroprotective effects in depression.[200] This study proved that  $CeO_2@BSA$  could effectively modulate pathological changes and ameliorate depression-like behaviors, confirming ROS as a therapeutic target for depression and offering inspiration for future research.

## 5. Challenges and perspectives

Oxidative balance is key for maintaining homeostasis. Excessive ROS will attack biomacromolecules directly and disturb normal signaling pathways, resulting in severe brain diseases. Nanozymes, as a new antioxidant, show great potential in regulating oxidation pressure (Table 4). As a new star, nanozymes have been concerned widely and developed rapidly, but also faced with general problems and challenges that need to be solved.

- (1) Comprehension and regulation of the *in vivo* biological behavior of nanozymes. Nanozymes have shown good catalytic activity *in vitro*, but their distribution, activity, and metabolism *in vivo* still need further in-depth investigation. Thus, a more systematic and comprehensive bioeffect of nanozymes is essential to translational applications.
- (2) Design and development of novel catalytic types of nanozymes. There are seven types of natural enzymes, which are oxidoreductase, transferases, hydrolases, lyases, isomerases, ligases, and translocases. Among these, oxidoreductases, hydrolases, lyases, and isomerases have been realized in nanozymes, and there are still transferases and ligases cannot do. So, developing novel catalytic types of nanozymes and multienzyme mimetic activity of nanozymes represent a crucial direction. This is especially relevant for brain diseases, which often involve complex biochemical processes.
- (3) Deciphering the catalytic mechanism of nanozymes, and achieving activity controllability *in vivo*. Different brain diseases may require different catalytic reactions, and these are quite complicated. But the universal conclusions or rules of nanozymes are few. The exploration of catalytic mechanism serves as a guiding force for the rational design as well as activity controllability of nanozymes with desired. The physicochemical properties of nanozymes, encompassing constituent atoms, crystal forms, sizes, and surface modifications, can be customized to optimize their catalytic activity, making them more suitable for diagnosing and treating particular brain diseases.

- (4) Expanding the application scenarios of nanozymes. The versatile attributes of nanozymes, including superparamagnetism, fluorescence, photothermal capabilities, BBB penetration, and enzymatic activity, coupled with their facile functionalization, position them as highly promising tools in the realm of therapeutic applications in brain diseases. The further promotion of nanozymes depends on the matched biomedical application scenario. How to combine different characteristics of nanozymes with brain diseases, making the best use of advantages and bypassing the disadvantages will promote the nanozymes from bench to bedside.

## Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

## Data availability

Data will be made available on request.

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## Appendix A. Supplementary data

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## References

- [1] W. Zhang, W. Wang, D.X. Yu, Z. Xiao, Z. He, Application of nanodiagnostics and nanotherapy to CNS diseases, *Nanomedicine (Lond.)* 13 (2018) 2341–2371, <https://doi.org/10.2217/nmm-2018-0163>.
- [2] B.C.V. Campbell, D.A. De Silva, M.R. Macleod, S.B. Coutts, L.H. Schwamm, S. M. Davis, G.A. Donnan, Ischaemic stroke, *Nat. Rev. Dis. Primers* 5 (2019) 70, <https://doi.org/10.1038/s41572-019-0118-8>.
- [3] A. Khellaf, D.Z. Khan, A. Helmy, Recent advances in traumatic brain injury, *J. Neurol.* 266 (2019) 2878–2889, <https://doi.org/10.1007/s00415-019-09541-4>.
- [4] L.M. DeAngelis, Brain tumors, *N. Engl. J. Med.* 344 (2001) 114–123, <https://doi.org/10.1056/NEJM200101113440207>.
- [5] S. Voet, S. Srinivasan, M. Lamkanfi, G. van Loo, Inflammasomes in neuroinflammatory and neurodegenerative diseases, *EMBO Mol. Med.* 11 (2019), e10248, <https://doi.org/10.15252/emmm.201810248>.
- [6] M.J. Armstrong, M.S. Okun, Diagnosis and treatment of Parkinson disease: a review, *J. Am. Med. Assoc.* 323 (2020) 548–560, <https://doi.org/10.1001/jama.2019.22360>.
- [7] F. Leng, P. Edison, Neuroinflammation and microglial activation in Alzheimer disease: where do we go from here? *Nat. Rev. Neurol.* 17 (2021) 157–172, <https://doi.org/10.1038/s41582-020-00435-y>.
- [8] Z. Gao, Y. Chen, X. Cai, R. Xu, Predict drug permeability to blood–brain-barrier from clinical phenotypes: drug side effects and drug indications, *Bioinformatics* 33 (2017) 901–908, <https://doi.org/10.1093/bioinformatics/btw713>.
- [9] Y. Song, C. Hu, Y. Fu, H. Gao, Modulating the blood–brain tumor barrier for improving drug delivery efficiency and efficacy, *VIEW.* 3 (2022) 20200129, <https://doi.org/10.1002/VIW.20200129>.
- [10] B.L. Edlow, J. Claassen, N.D. Schiff, D.M. Greer, Recovery from disorders of consciousness: mechanisms, prognosis and emerging therapies, *Nat. Rev. Neurol.* 17 (2021) 135–156, <https://doi.org/10.1038/s41582-020-00428-x>.
- [11] C. Aubinet, L. Murphy, M.A. Bahri, S.K. Larroque, H. Cassol, J. Annen, M. Carrière, S. Wannez, A. Thibaut, S. Laureys, Brain, behavior, and cognitive interplay in disorders of consciousness: a multiple case study, *Front. Neurol.* 9 (2018) 665, <https://doi.org/10.3389/fneur.2018.00665>.
- [12] V.L. Feigin, B.A. Stark, C.O. Johnson, G.A. Roth, C. Bisignano, G.G. Abady, M. Abbasifard, M. Abbasi-Kangevari, F. Abd-Allah, V. Abedi, A. Abualhasan, N. ME. Abu-Rmeileh, A.I. Abushouk, O.M. Adebayo, G. Agarwal, P. Agasthi, B. O. Ahinkorah, S. Ahmad, S. Ahmadi, Y. Ahmed Salih, B. Aji, S. Akbarpour, R. O. Akinyemi, H. Al Hamad, F. Alahdab, S.M. Alif, V. Alipour, S.M. Aljunid, S. Almustanyir, R.M. Al-Raddadi, R. Al-Shahi Salman, N. Alvis-Guzman,

- R. Ancuceanu, D. Anderlini, J.A. Anderson, A. Ansari, I.C. Antonazzo, J. Arabloo, J. Årnlöv, K.D. Artanti, Z. Aryan, S. Asgari, T. Ashraf, M. Athar, A. Atreya, M. Ausloos, A.A. Baig, O.C. Baltatu, M. Banach, M.A. Barboza, S.L. Barker-Collo, T.W. Bärnighausen, M.T.U. Barone, S. Basu, G. Bazmandegan, E. Beghi, M. Beheshti, Y. Béjot, A.W. Bell, D.A. Bennett, I.M. Bensenor, W.M. Bezabhe, Y. M. Bezabih, A.S. Bhagavathula, P. Bhardwaj, K. Bhattacharyya, A. Bijani, B. Bikbov, M.M. Birhanu, A. Boloor, A. Bonny, M. Brauer, H. Brenner, D. Bryazka, Z.A. Butt, F.L. Caetano dos Santos, I.R. Campos-Nonato, C. Cantu-Brito, J. J. Carrero, C.A. Castañeda-Orjuela, A.L. Catapano, P.A. Chakraborty, J. Charan, S. G. Choudhari, E.K. Chowdhury, D.-T. Chu, S.-C. Chung, D. Colozza, V.M. Costa, S. Costanzo, M.H. Criqui, O. Dadrás, B. Dagnew, X. Dai, K. Dalal, A.A. M. Damasceno, E. D'Amico, L. Dandona, R. Dandona, J. Darega Gela, K. Davletov, V. De la Cruz-Góngora, R. Desai, D. Dharmetiya, S.D. Dharmaratne, M.L. Dhimial, M. Dhimial, D. Diaz, M. Dichgans, K. Dokova, R. Doshi, A. Douiri, B.B. Duncan, S. Eftekhazadeh, M. Ekholuentele, N. El Nahas, I.Y. Elgendy, M. Elhadi, S.I. El-Jaafari, M. Endres, A.Y. Endries, D.A. Erku, E.J.A. Faraon, U. Farrow, F. Farzadfar, A.H. Feroze, I. Filip, F. Fischer, D. Flood, M.M. Gad, S. Gaidhane, R. Ghanei Gheslugh, A. Ghashghaei, N. Ghith, G. Ghozali, S. Ghozy, A. Gialluisi, S. Giampaoli, S.A. Gilani, P.S. Gill, E.V. Gnedovskaya, M. Golechha, A.C. Goulart, Y. Guo, R. Gupta, V.B. Gupta, V.K. Gupta, P. Gyanwali, N. Hafezi-Nejad, S. Hamidi, A. Hanif, G.J. Hankey, A. Hargono, A. Hashi, T.S. Hassan, H.Y. Hassan, R.J. Havmoeller, S.I. Hay, K. Hayat, M.I. Hegazy, C. Herteliu, R. Holla, S. Hosticus, M. Househ, J. Huang, A. Humayun, B.-F. Hwang, L. Iacoviello, I. Iavicoli, S. E. Ibitoye, O.S. Ilesanmi, I.M. Ilic, M.D. Ilic, U. Iqbal, S.S.N. Irvani, S.M.S. Islam, N.E. Ismail, H. Iso, G. Isola, M. Iwagami, L. Jacob, V. Jain, S.-I. Jang, S.K. Jayapal, S. Jayaram, R. Jayawardena, P. Jeemon, R.P. Jha, W.D. Johnson, J.B. Jonas, N. Joseph, J.J. Jozwiak, M. Jürisson, R. Kalani, R. Kalhor, Y. Kalkonde, A. Kamath, Z. Kamiab, T. Kanchan, H. Kandel, A. Karch, P.D. Katoto, G. A. Kayode, P. Keshavarz, Y.S. Khader, E.A. Khan, I.A. Khan, M. Khan, M. AB. Khan, M.N. Khatib, J. Khubchandani, G.R. Kim, M.S. Kim, Y.J. Kim, A. Kisa, S. Kisa, M. Kivimäki, D. Kolte, A. Koolivand, S.L. Koulmane Laxminarayana, A. i. Koyanagi, K. Krishan, V. Krishnamoorthy, R.V. Krishnamurthi, G.A. Kumar, D. Kusuma, C. La Vecchia, B. Lacey, H.M. Lak, T. Lallukka, S. Lasrado, P. M. Lavados, M. Leonardi, B. Li, S. Li, H. Lin, R.-T. Lin, X. Liu, W.D. Lo, S. Lorkowski, G. Lucchetti, R. Lutzky Saute, H. Magdy Abd El Razek, F. G. Magnani, P.B. Mahajan, A. Majeed, A. Makki, R. Malekzadeh, A.A. Malik, N. Manafi, M.A. Mansournia, L.G. Mantovani, S. Martini, G. Mazzaglia, M. M. Mehndiratta, R.G. Menezes, A. Meretoja, A.G. Mersha, J. Miao Jonasson, B. Miazgowski, T. Miazgowski, I.M. Michalek, E.M. Mirzakhimov, Y. Mohammad, A. Mohammadian-Hafshejani, S. Mohammed, A.H. Mokdad, Y. Mokhtari, M. Molokhia, M.A. Moni, A.A. Montasir, R. Moradzadeh, L. Morawska, J. Morze, W. Muruet, K.I. Musa, A.J. Nagarajan, M. Naghavi, S. Narasimha Swamy, B. R. Nascimento, R.I. Negoi, S. Neupane Kandel, T.H. Nguyen, B.O. Norrving, J. J. Noubiap, V.E. Nwatah, B. Oancea, O.O. Odukoya, A.T. Olagunju, H. Orru, M. O. Owolabi, J.R. Padubidri, A. Pana, T. Parekh, E.-C. Park, F. Pashazadeh Kan, M. Pathak, M.F.P. Peres, A. Perianayagam, T.-M. Pham, M.A. Piradov, V. Podder, S. Polinder, M.J. Postma, A. Pourshams, A. Radfar, A. Rafiei, A. Raggi, F. Rahim, V. Rahimi-Movaghar, M. Rahman, M.A. Rahman, A.M. Rahmani, N. Rajai, P. Ranasinghe, C.R. Rao, S.J. Rao, P. Rathi, D.L. Rawaf, S. Rawaf, M.B. Reitsma, V. Renjith, A.M.N. Renzaho, A. Rezapour, J.A.B. Rodriguez, L. Roever, M. Romoli, A. Rynkiewicz, S. Sacco, M. Sadeghi, S. Saeedi Moghaddam, A. Sahebkar, K. M. Saif-Ur-Rahman, R. Salah, M. Samaei, A.M. Samy, I.S. Santos, M.M. Santric-Milicevic, N. Sarrafzadegan, B. Sathian, D. Sattin, S. Schiavolin, M.P. Schlaich, M. I. Schmidt, A.E. Schutte, S.G. Sepanlou, A. Seylani, F. Sha, S. Shahabi, M. A. Shaikh, M. Shannawaz, M.S.R. Shawon, A. Sheikh, S. Sheikhbahaie, K. Shibuya, S. Siabani, D.A.S. Silva, J.A. Singh, J.K. Singh, V.Y. Skryabin, A. A. Skryabina, B.H. Sobaih, S. Stortecy, S. Stranges, E.G. Tadesse, I.U. Tarigan, M.-H. Temsah, Y. Teuschl, A.G. Thrift, M. Tonelli, M.R. Tovani-Palone, B.X. Tran, M. Tripathi, G.W. Tsegaye, A. Ullah, B. Unim, B. Unnikrishnan, A. Vakilian, S. Valadan Tahbaz, T.J. Vasankari, N. Venketasubramanian, D. Vervoort, B. Vo, V. Volovici, K. Vosoughi, G.T. Vu, L.G. Vu, H.A. Wafa, Y. Waheed, Y. Wang, T. Wijeratne, A.S. Winkler, C.D.A. Wolfe, M. Woodward, J.H. Wu, S. Wulf Hanson, X. Xu, L. Yadav, A. Yadollahpour, S.H. Yahyazadeh Jabbari, K. Yamagishi, H. Yatsuya, N. Yonemoto, C. Yu, I. Yunusa, M.S. Zaman, S. B. Zaman, M. Zamanian, R. Zand, A. Zandifar, M.S. Zastrozhin, A. Zastrozhina, Y. Zhang, Z.-J. Zhang, C. Zhong, Y.M.H. Zuniga, C.J.L. Murray, Global, regional, and national burden of stroke and its risk factors, 1990–2019: a systematic analysis for the Global Burden of Disease Study 2019, *Lancet Neurol.* 20 (10) (2021) 795–820, [https://doi.org/10.1016/S1474-4422\(21\)00252-0](https://doi.org/10.1016/S1474-4422(21)00252-0).
- [13] V.L. Feigin, T. Vos, F. Alahdab, A.M.L. Amit, T.W. Bärnighausen, E. Beghi, M. Beheshti, P.P. Chavan, M.H. Criqui, R. Desai, S. Dhamminda Dharmaratne, E. R. Dorsey, A. Wilder Eagan, I.Y. Elgendy, I. Filip, S. Giampaoli, G. Giussani, N. Hafezi-Nejad, M.K. Hole, T. Ikeda, C. Owens Johnson, R. Kalani, K. Khatib, J. Khubchandani, D. Kim, W.J. Koroshetz, V. Krishnamoorthy, R. V. Krishnamurthi, X. Liu, W.D. Lo, G. Logroscino, G.A. Mensah, T.R. Miller, S. Mohammed, A.H. Mokdad, M. Moradi-Lakeh, S.D. Morrison, V.K. N. Shivamurthy, M. Naghavi, E. Nichols, B.O. Norrving, C.M. Odell, E. Pupillo, A. Radfar, G.A. Roth, A. Shafieesabet, A. Sheikh, S. Sheikhbahaie, J.I. Shin, J. A. Singh, T.J. Steiner, L.J. Stovner, M.T. Wallin, J. Weiss, C. Wu, J.R. Zunt, J. D. Adelson, C.J.L. Murray, Burden of neurological disorders across the US from 1990–2017: a global burden of disease study, *JAMA Neurol.* 78 (2) (2021) 165, <https://doi.org/10.1001/jamaneurol.2020.4152>.
- [14] V.L. Feigin, A.A. Abajobir, K.H. Abate, F. Abd-Allah, A.M. Abdulle, S.F. Abera, G. Y. Abyu, M.B. Ahmed, A.N. Aichour, I. Aichour, M.T.E. Aichour, R.O. Akinymeni, S. Alabed, R. Al-Raddadi, N. Alvis-Guzman, A.T. Amare, H. Ansari, P. Anwari, J. Årnlöv, H. Asayesh, S.W. Asgedom, T.M. Atey, L. Avila-Burgos, E. Frinell, G. A. Avokpaho, M.R. Azarpazhooh, A. Barac, M. Barboza, S.L. Barker-Collo, T. Bärnighausen, N. Bedi, E. Beghi, D.A. Bennett, I.M. Bensenor, A. Berhane, B. D. Betu, S. Bhaumik, S.M. Birlik, S. Biryukov, D.J. Boneya, L.N.B. Bulto, H. Carabin, D. Casey, C.A. Castañeda-Orjuela, F. Catalá-López, H. Chen, A. A. Chitheer, R. Chowdhury, H. Christensen, L. Dandona, R. Dandona, G.A. de Veber, S.D. Dharmaratne, H.P. Do, K. Dokova, E.R. Dorsey, R.G. Ellenbogen, S. Eskandarieh, M.S. Farvid, S.-M. Fereshtehnejad, F. Fischer, K.J. Foreman, J. M. Geleijnse, R.F. Gillum, G. Giussani, E.M. Goldberg, P.N. Gona, A.C. Goulart, H. C. Gugnani, R. Gupta, V. Hachinski, R. Gupta, R.R. Hamadeh, M. Hambisa, G. J. Hankey, H.A. Hareri, R. Havmoeller, S.I. Hay, P. Heydarpour, P.J. Hotez, M.C. B. Jakovljevic, M. Javanbakht, P. Jeemon, J.B. Jonas, Y. Kalkonde, A. Kandel, A. Karch, A. Kasaieian, A. Kaster, P.N. Keiyoro, Y.S. Khader, I.A. Khalil, E.A. Khan, Y.-H. Khang, A. Tawfih, A. Khoja, J. Khubchandani, C. Kulkarni, D. Kim, Y.J. Kim, M. Kivimaki, Y. Kokubo, S. Kosen, M. Kravchenko, R.V. Krishnamurthi, B.K. Defo, G.A. Kumar, R. Kumar, H.H. Kyu, A. Larsson, P.M. Lavados, Y. Li, X. Liang, M. L. Liben, W.D. Lo, G. Logroscino, P.A. Lotufo, C.T. Loy, M.T. Mackay, H.M.A. El Razek, M.M.A. El Razek, A. Majeed, R. Malekzadeh, T. Manhertz, L.G. Mantovani, J. Massano, M. Mazidi, C. McAlinden, S. Mehata, M.M. Mehndiratta, Z. A. Emish, V. Mendoza, M.A. Mengistie, G.A. Mensah, A. Meretoja, I.S. Santos, B. Mezgebe, T.R. Miller, S.R. Mishra, N.M. Ibrahim, A. Mohammadi, K. E. Mohammed, S. Mohammed, A.H. Mokdad, M. Moradi-Lakeh, I.M. Velasquez, K.I. Musa, M. Naghavi, J.W. Ngunjiri, C.T. Nguyen, C. Le Nguyen, Q. Le Nguyen, T. H. Nguyen, E. Nichols, D.N.A. Ningrum, V.M. Nong, B.O. Norrving, J.J. N. Noubiap, F.A. Ogblo, M.O. Owolabi, J.D. Pandian, P.G. Parmar, D.M. Pereira, M. Pertzold, M.R. Phillips, M.A. Piradov, R.G. Poulton, F. Pourmalek, M. Qorbani, A. Rafay, M. Rahman, M.H. Rahman, R.K. Rai, S. Rajscik, A. Ranta, S. Rawaf, A.M. N. Renzaho, M.S. Rezaei, G.A. Roth, G. Roshandel, E. Rubagotti, P. Sachdev, S. Safiri, R. Sahathevan, M.A. Sahraian, A.M. Samy, P. Santalucia, I.S. Santos, B. Sartorius, M. Satpathy, M. Sawhney, M.I. Saylan, S.G. Sepanlou, M.A. Shaikh, R. Shakir, M. Shamsizadeh, K.N. Sheth, M. Shigematsu, H. Shoman, D.A.S. Silva, M. Smith, E. Sobngwi, L.A. Sposato, J.D. Stanaway, D.J. Stein, T.J. Steiner, L. J. Stovner, R.S. Abdulkader, C. El Szeoke, R. Tabarés-Seisdedos, D. Tanne, A. M. Theadom, A.G. Thrift, D.L. Tirschwell, R. Topor-Madry, B.X. Tran, T. Truelsan, K.B. Tuem, K.N. Ukwaja, O.A. Uthman, Y.Y. Varakin, T. Vasankari, N. Venketasubramanian, V.V. Vlassov, F. Wadilo, T. Wakayo, M.T. Wallin, E. Weiderpass, R. Westerman, T. Wijeratne, C.S. Wiysong, M.A. Woldu, C.D. A. Wolfe, D. Xavier, G. Xu, Y. Yano, H.H. Yimam, N. Yonemoto, C. Yu, Z. Zaidi, M. El Sayed Zaki, J.R. Zunt, C.J.L. Murray, T. Vos, Global, regional, and national burden of neurological disorders during 1990–2015: a systematic analysis for the Global Burden of Disease Study 2015, *Lancet Neurol.* 16 (11) (2017) 877–897, [https://doi.org/10.1016/S1474-4422\(17\)30299-5](https://doi.org/10.1016/S1474-4422(17)30299-5).
- [15] V.L. Feigin, E. Nichols, T. Alam, M.S. Bannick, E. Beghi, N. Blake, W.J. Culpepper, E.R. Dorsey, A. Elbaz, R.G. Ellenbogen, J.L. Fisher, C. Fitzmaurice, G. Giussani, L. Glennie, S.L. James, C.O. Johnson, N.J. Kassebaum, G. Logroscino, B. Marin, W.C. Mountjoy-Venning, M. Nguyen, R. Ofori-Asenso, A.P. Patel, M. Piccininni, G.A. Roth, T.J. Steiner, L.J. Stovner, C.E.I. Szeoke, A. Theadom, S.E. Vollset, M. T. Wallin, C. Wright, J.R. Zunt, N. Abbasi, F. Abd-Allah, A. Abdelalim, I. Abdollahpour, V. Aboyans, H.N. Abraha, D. Acharya, A.A. Adamu, O. M. Adebayo, A.M. Adeoye, J.C. Adusu, M. Afarideh, S. Agrawal, A. Ahmadi, M. B. Ahmed, A.N. Aichour, I. Aichour, M.T.E. Aichour, R.O. Akinymeni, N. Akseer, A. Al-Eyadhy, R. Al-Shahi Salman, F. Alahdab, K.A. Alene, S.M. Aljuni, K. Altirkawi, N. Alvis-Guzman, N.H. Anber, C.A.T. Antonio, J. Arabloo, O. Aremu, J. Årnlöv, H. Asayesh, R.J. Asghar, H.T. Atalay, A. Awasthi, B.P. Ayala Quintanilla, T.B. Ayuk, A. Badawi, M. Banach, J.A.M. Banoub, M.A. Barboza, S. L. Barker-Collo, T.W. Bärnighausen, B.T. Baune, N. Bedi, M. Behzadifar, M. Behzadifar, Y. Béjot, B.B. Bekele, A.B. Belachew, D.A. Bennett, I.M. Bensenor, A. Berhane, M. Beuran, K. Bhattacharyya, Z.A. Bhutta, B. Biadgo, A. Bijani, N. Billig, M.S. Bin Sayeed, C.K. Blazes, C. Brayne, Z.A. Butt, I.R. Campos-Nonato, C. Cantu-Brito, M. Car, R. Cárdenas, J.J. Carrero, F. Carvalho, C. A. Castañeda-Orjuela, F. Castro, F. Catalá-López, E. Cerin, Y. Chaiah, J.-C. Chang, I. Chatziralli, P.-C. Chiang, H. Christensen, D.J. Christopher, C. Cooper, P. A. Cortesi, V.M. Costa, M.H. Criqui, C.S. Crowe, A.A.M. Damasceno, A. Daryani, V. De la Cruz-Góngora, F.P. De la Hoz, D. De Leo, G.T. Demoz, K. Deribe, S. D. Dharmaratne, D. Diaz, M.T. Dinberu, S. Djalalinia, D.T. Doku, M. Dubey, E. Dubljanin, E.E. Duken, D. Edvardsson, Z. El-Khatib, M. Endres, A.Y. Endries, S. Eskandarieh, A. Esteghamati, S. Esteghamati, F. Farhadi, A. Faro, F. Farzadfar, M.H. Farzaei, B. Fatima, S.-M. Fereshtehnejad, E. Fernandes, G.T. Feyissa, I. Filip, F. Fischer, T. Fukumoto, M. Ganji, F.G. Gankpe, M.A. Garcia-Gordillo, A.K. Gebre, T.G. Gebremichael, B.K. Gelaw, J.M. Geleijnse, D. Geremew, K.E. Gezae, M. Ghasemi-Kasman, M.Y. Gidey, P.S. Gill, T.K. Gill, E.T. Girma, E. V. Gnedovskaya, A.C. Goulart, A. Grada, G. Grosso, Y. Guo, R. Gupta, R. Gupta, J. A. Haagsma, T.B. Hagos, A. Haj-Mirzaian, A. Haj-Mirzaian, R.R. Hamadeh, S. Hamidi, G.J. Hankey, Y. Hao, J.M. Haro, H. Hassankhani, H.Y. Hassen, R. Havmoeller, S.I. Hay, M.I. Hegazy, B. Heidari, A. Henok, F. Heydarpour, C. L. Hoang, M.K. Hole, E. Homaie Rad, S.M. Hosseini, G. Hu, E.U. Igumbor, O. S. Ilesanmi, S.S.N. Irvani, S.M.S. Islam, M. Jakovljevic, M. Javanbakht, R.P. Jha, Y.B. Jobanputra, J.B. Jonas, J.J. Jozwiak, M. Jürisson, A. Kahsay, R. Kalani, Y. Kalkonde, T.A. Kamil, T. Kanchan, M. Karami, A. Karch, N. Karimi, A. Kasaieian, T.D. Kassa, Z.Y. Kassa, A. Kaul, A.T. Kefale, P.N. Keiyoro, Y. S. Khader, M.A. Khafaie, I.A. Khalil, E.A. Khan, Y.-H. Khang, H. Khaizee, A. A. Kiadali, D.N. Kiirithio, A.S. Kim, D. Kim, Y.-E. Kim, Y.J. Kim, A. Kisa, Y. Kokubo, A.i. Koyanagi, R.V. Krishnamurthi, B. Kuate Defo, B. Kucuk Bicer, M. Kumar, B. Lacey, A. Lafranconi, V.C. Lansingh, A. Latifi, C.T. Leshargie, S. Li, Y. Li, Y. Liao, S. Linn, W.D. Lo, J.C.F. Lopez, S. Lorkowski, P.A. Lotufo, R.M. Lucas, R. Lunevicius, M.T. Mackay, N.B. Mahotra, M. Majdan, R. Majdzadeh, A. Majeed, R. Malekzadeh, D.C. Malta, N. Manafi, M.A. Mansournia, L.G. Mantovani,

- W. März, T.P. Mashamba-Thompson, B.B. Massenburg, K.K.V. Mate, C. McAlinden, J.J. McGrath, V. Mehta, T. Meier, H.G. Meles, A. Melese, P.T. N. Memiah, Z.A. Memish, W. Mendoza, D.T. Mengistu, G. Mengistu, A. Meretoja, T.J. Mietzowa, T. Mestrovic, B. Miazgowski, T.R. Miller, G.K. Mini, E.M. Mirzakhimov, B. Moazen, B. Mohajer, N. Mohammad Gholi Mezerji, M. Mohammadi, M. Mohammadi-Khanaposhtani, R. Mohammadibakhsh, M. Mohammadnia-Afrouzi, S. Mohammad, F. Mohebi, A.H. Mokdad, L. Monasta, S. Mondello, Y. Moodley, M. Moosazadeh, G. Moradi, M. Moradi-Lakeh, M. Moradinazar, P. Moraga, I. Moreno Velásquez, S.D. Morrison, S.M. Mousavi, O.S. Muhammed, W. Muruet, K.I. Musa, G. Mustafa, M. Naderi, G. Nagel, A. Naheed, G. Naik, F. Najafi, V. Nangia, I. Negoi, R.I. Negoi, C.R.J. Newton, J. W. Ngunjiri, C.T. Nguyen, L.H. Nguyen, D.N.A. Ningrum, Y.L. Nirayo, M. R. Nixon, B.o. Norrving, J.J. Noubiap, M. Nourollahpour Shiadeh, P.S. Nyasulu, O.S. Ogah, I.-H. Oh, A.T. Olagunju, T.O. Olagunju, P.R. Olivares, O. E. Onwujekwe, E. Oren, M.O. Owolabi, M. Pa, A.H. Pakpour, W.-H. Pan, S. Panda-Jonas, J.D. Pandian, S.K. Patel, D.M. Pereira, F.M. Petzold, J.D. Pillay, M. A. Piradov, G.V. Polaczkyk, S. Polinder, M.J. Postma, R. Poulton, H. Poustchi, S. Prakash, V. Prakash, M. Qorbani, A. Radfar, A. Rafay, A. Rafiei, F. Rahim, V. Rahimi-Movaghar, M. Rahman, M.H.U. Rahman, M.A. Razhan, P. Rajati, U. Ram, A. Ranta, D.L. Rawaf, S. Rawaf, N. Reinig, C. Reis, A.M.N. Renzaho, S. Resnikoff, S. Rezaeian, M.S. Rezaei, C.M. Rios González, N.L.S. Roberts, L. Roever, L. Ronfani, E.M. Roro, G. Roshandel, A. Rostami, P. Sabbagh, R. L. Sacco, P.S. Sachdev, B. Saddik, H. Safari, R. Safari-Faramani, S. Safi, S. Safiri, R. Sagar, R. Sahathevan, A. Sahebkar, M.A. Sahraian, P. Salamati, S. Salehi Zahabi, Y. Salimi, A.M. Samy, J. Sanabria, I.S. Santos, M.M. Santric Milicevic, N. Sarrafzadegan, B. Sartorius, S. Sarvi, B. Sathian, M. Satpathy, A.R. Sawant, M. Sawhney, I.J.C. Schneider, B. Schöttker, D.C. Schwebel, S. Seedat, S. G. Sepanlou, H. Shabaninejad, A. Shafieesabet, M.A. Shaikh, R.A. Shakir, M. Shams-Beyranvand, M. Shamsizadeh, M. Sharif, M. Sharif-Alhoseini, J. She, A. Sheikh, K.N. Sheth, M. Shigematsu, R. Shiri, R. Shirkoobi, I. Shiu, S. Siabani, T.J. Siddiqi, I.D. Sigfusdottir, R. Sigurvinsson, D.H. Silberberg, J.P. Silva, D.G. A. Silveira, J.A. Singh, D.N. Sinha, E. Skiadaresi, M. Smith, B.H. Sobaih, S. Sobhani, M. Soofi, I.N. Soyiri, L.A. Spasato, D.J. Stein, M.B. Stein, M.A. Stokes, M.B. Sufiyan, B.L. Sykes, P.N. Sylaja, R. Tabarés-Seisdedos, B.J. Te Ao, A. Tehrani-Banihashemi, M.-H. Temsah, O. Temsah, J.S. Thakur, A.G. Thrift, R. Topor-Madry, M. Tortajada-Gurbés, M.R. Tovani-Palane, B.X. Tran, K.B. Tran, T.C. Truelsen, A.G. Tsadik, L. Tudor Car, K.N. Ukwajia, I. Ullah, M.S. Usman, O. A. Uthman, P.R. Valdez, T.J. Vasankari, R. Vasanathan, Y. Veisani, N. Venketasubramanian, F.S. Violante, V. Vlassov, K. Vosoughi, G.T. Vu, I. S. Vujcic, F.S. Wagnew, Y. Waheed, Y.-P. Wang, E. Weiderpass, J. Weiss, H. A. Whiteford, T. Wijeratne, A.S. Winkler, C.S. Wiysonge, C.D.A. Wolfe, G. Xu, A. Yadollahpour, T. Yamada, Y. Yano, M. Yaseri, H. Yatsuya, E.M. Yimer, P. Yip, E. Yisma, N. Yonemoto, M. Youseffard, C. Yu, Z. Zaidi, S.B. Zaman, M. Zamani, H. Zandian, Z. Zare, Y. Zhang, S. Zodepy, M. Naghavi, C.J.L. Murray, T. Vos, Global, regional, and national burden of neurological disorders, 1990–2016: a systematic analysis for the Global Burden of Disease Study 2016, *Lancet Neurol.* 18 (5) (2019) 459–480, [https://doi.org/10.1016/S1474-4422\(18\)30499-X](https://doi.org/10.1016/S1474-4422(18)30499-X).
- [16] F. Collin, Chemical basis of reactive oxygen species reactivity and involvement in neurodegenerative diseases, *Int. J. Mol. Sci.* 20 (10) (2019) 2407, <https://doi.org/10.3390/ijms20102407>.
- [17] A. Jurcau, A.I. Ardelean, Oxidative stress in ischemia/reperfusion injuries following acute ischemic stroke, *Biomedicines.* 10 (2022) 574, <https://doi.org/10.3390/biomedicines10030574>.
- [18] S. Bhatt, A.N. Nagappa, C.R. Patil, Role of oxidative stress in depression, *Drug Discov. Today* 25 (2020) 1270–1276, <https://doi.org/10.1016/j.drudis.2020.05.001>.
- [19] H. Wei, L. Gao, K. Fan, J. Liu, J. He, X. Qu, S. Dong, E. Wang, X. Yan, Nanozymes: A clear definition with fuzzy edges, *Nano Today* 40 (2021), 101269, <https://doi.org/10.1016/j.nantod.2021.101269>.
- [20] L. Gao, J. Zhuang, L. Nie, J. Zhang, Y. Zhang, N. Gu, T. Wang, J. Feng, D. Yang, S. Perrett, X. Yan, Intrinsic peroxidase-like activity of ferromagnetic nanoparticles, *Nat. Nanotechnol.* 2 (2007) 577–583, <https://doi.org/10.1038/nnano.2007.260>.
- [21] R. Zhang, K. Fan, X. Yan, Nanozymes: created by learning from nature, *Sci. China Life Sci.* 63 (2020) 1183–1200, <https://doi.org/10.1007/s11427-019-1570-7>.
- [22] R. Zhang, B. Xue, Y. Tao, H. Zhao, Z. Zhang, X. Wang, X. Zhou, B. Jiang, Z. Yang, X. Yan, K. Fan, Edge-Site Engineering of defective Fe–N<sub>4</sub> nanozymes with boosted catalase-like performance for retinal vasculopathies, *Adv. Mater.* 34 (2022) 2205324, <https://doi.org/10.1002/adma.202205324>.
- [23] G. Tosi, A. Vilella, R. Chhabra, M.J. Schmeisser, T.M. Boeckers, B. Ruozi, M. A. Vandelli, F. Forni, M. Zoli, A.M. Grabrucker, Insight on the fate of CNS-targeted nanoparticles. Part II: Intercellular neuronal cell-to-cell transport, *J. Control. Release* 177 (2014) 96–107, <https://doi.org/10.1016/j.jconrel.2014.01.004>.
- [24] W. Yang, X. Yang, L. Zhu, H. Chu, X. Li, W. Xu, Nanozymes: Activity origin, catalytic mechanism, and biological application, *Coord. Chem. Rev.* 448 (2021), 214170, <https://doi.org/10.1016/j.ccr.2021.214170>.
- [25] S. Zhao, H. Duan, Y. Yang, X. Yan, K. Fan, Fenozyme protects the integrity of the blood–brain barrier against experimental cerebral malaria, *Nano Lett.* 19 (2019) 8887–8895, <https://doi.org/10.1021/acs.nanolett.9b03774>.
- [26] Q. Zhu, Y. Huang, X. Zhu, L. Peng, H. Wang, S. Gao, Z. Yang, J. Zhang, X. Liu, Mannose-coated superparamagnetic iron oxide nanozyme for preventing postoperative cognitive dysfunction, *Materials Today Bio.* 19 (2023), 100568, <https://doi.org/10.1016/j.mtbio.2023.100568>.
- [27] N. Yin, Y. Wang, Y. Huang, Y. Cao, L. Jin, J. Liu, T. Zhang, S. Song, X. Liu, H. Zhang, Modulating nanozym-based nanomachines via microenvironmental feedback for differential photothermal therapy of orthotopic gliomas, *Adv. Sci.* 10 (2023) 2204937, <https://doi.org/10.1002/adv.202204937>.
- [28] J. Li, Z. Zhang, B. Zhang, X. Yan, K. Fan, Transferrin receptor 1 targeted nanomedicine for brain tumor therapy, *Biomater. Sci.* 11 (2023) 3394–3413, <https://doi.org/10.1039/D2BM02152H>.
- [29] P. Muhammad, S. Hanif, J. Li, A. Guller, F.U. Rehman, M. Ismail, D. Zhang, X. Yan, K. Fan, B. Shi, Carbon dots supported single Fe atom nanozyme for drug-resistant glioblastoma therapy by activating autophagy-lysosome pathway, *Nano Today* 45 (2022), 101530, <https://doi.org/10.1016/j.nantod.2022.101530>.
- [30] Q. Zhao, W. Du, L. Zhou, J. Wu, X. Zhang, X. Wei, S. Wang, Y. Huang, Y. Li, Transferrin-enabled blood-brain barrier crossing manganese-based nanozyme for rebalancing the reactive oxygen species level in ischemic stroke, *Pharmaceutics.* 14 (2022) 1122, <https://doi.org/10.3390/pharmaceutics14061122>.
- [31] Y. Guan, M. Li, K. Dong, N. Gao, J. Ren, Y. Zheng, X. Qu, Ceria/POMs hybrid nanoparticles as a mimicking metalloproteinase for treatment of neurotoxicity of amyloid- $\beta$  peptide, *Biomaterials* 98 (2016) 92–102, <https://doi.org/10.1016/j.biomaterials.2016.05.005>.
- [32] Y. Huang, J. Ren, X. Qu, Nanozymes: classification, catalytic mechanisms, activity regulation, and applications, *Chem. Rev.* 119 (2019) 4357–4412, <https://doi.org/10.1021/acs.chemrev.8b00672>.
- [33] P.K. Robinson, Enzymes: principles and biotechnological applications, *Essays Biochem.* 59 (2015) 1–41, <https://doi.org/10.1042/bse0590001>.
- [34] Y. Jiang, A.M. Brynskikh, S. Devika, A.V. Kabanov, SOD1 nanozyme salvages ischemic brain by locally protecting cerebral vasculature, *J. Control. Release* 213 (2015) 36–44, <https://doi.org/10.1016/j.jconrel.2015.06.021>.
- [35] D. Jiang, D. Ni, Z.T. Rosenkrans, P. Huang, X. Yan, W. Cai, Nanozyme: new horizons for responsive biomedical applications, *Chem. Soc. Rev.* 48 (2019) 3683–3704, <https://doi.org/10.1039/C8CS00718G>.
- [36] B. Srinivasan, A guide to the Michaelis-Menten equation: steady state and beyond, *FEBS J.* 289 (2022) 6086–6098, <https://doi.org/10.1111/febs.16124>.
- [37] M. Comotti, C. Della Pina, R. Matarrese, M. Rossi, The catalytic activity of “naked” gold particles, *Angew. Chem. Int. Ed.* 43 (2004) 5812–5815, <https://doi.org/10.1002/anie.200460446>.
- [38] M. Comotti, C. Della Pina, E. Falletta, M. Rossi, Aerobic oxidation of glucose with gold catalyst: hydrogen peroxide as intermediate and reagent, *Adv. Synth. Catal.* 348 (2006) 313–316, <https://doi.org/10.1002/adsc.200505389>.
- [39] J. Chen, Q. Ma, M. Li, D. Chao, L. Huang, W. Wu, Y. Fang, S. Dong, Glucose-oxidase like catalytic mechanism of noble metal nanozymes, *Nat. Commun.* 12 (2021) 3375, <https://doi.org/10.1038/s41467-021-23737-1>.
- [40] S. Zhao, H. Li, R. Liu, N.a. Tao, L. Deng, Q. Xu, J. Hou, J. Sheng, J. Zheng, L. Wang, W. Chen, S. Guo, Y.-N. Liu, Nitrogen-centered lactate oxidase nanozyme for tumor lactate modulation and microenvironment remodeling, *J. Am. Chem. Soc.* 145 (18) (2023) 10322–10332, <https://doi.org/10.1021/jacs.3c02005>.
- [41] M. Li, J. Chen, W. Wu, Y. Fang, S. Dong, Oxidase-like MOF-818 nanozyme with high specificity for catalysis of catechol oxidation, *J. Am. Chem. Soc.* 142 (2020) 15569–15574, <https://doi.org/10.1021/jacs.0c07273>.
- [42] Y. Li, P. Wang, L. Huang, C. Jia, X. Gao, S. Liu, S. Wang, P. Zhao, J. Sun, D. Zhang, M. Zhu, Y. Shen, J. Wang, Schiff-base chemistry-coupled catechol oxidase-like nanozyme reaction as a universal sensing mode for ultrasensitive biosensing, *Anal. Chem.* 95 (7) (2023) 3769–3778, <https://doi.org/10.1021/acs.analchem.2c04897>.
- [43] J. Liu, X. Hu, S. Hou, T. Wen, W. Liu, X. Zhu, J. Yin, X. Wu, Au@Pt core/shell nanorods with peroxidase- and ascorbate oxidase-like activities for improved detection of glucose, *Sens. Actuators B* 166–167 (2012) 708–714, <https://doi.org/10.1016/j.snb.2012.03.045>.
- [44] Y. Chen, T. Chen, X. Wu, G. Yang, Oxygen vacancy-engineered PEGylated MoO<sub>3-x</sub> Nanoparticles with Superior Sulfite Oxidase Mimetic Activity for Vitamin B1 Detection, *Small* 15 (2019) 1903153, <https://doi.org/10.1002/smll.201903153>.
- [45] D. Wu, J. Li, S. Xu, Q. Xie, Y. Pan, X. Liu, R. Ma, H. Zheng, M. Gao, W. Wang, J. Li, X. Cai, F. Jaouen, R. Li, Engineering Fe–N Doped Graphene to Mimic Biological Functions of NADPH Oxidase in Cells, *J. Am. Chem. Soc.* 142 (2020) 19602–19610, <https://doi.org/10.1021/jacs.0c08360>.
- [46] C. Zheng, W. Ke, T. Yin, X. An, Intrinsic peroxidase-like activity and the catalytic mechanism of gold@ carbon dots nanocomposites, *RSC Adv.* 6 (2016) 35280–35286, <https://doi.org/10.1039/C6RA01917J>.
- [47] H. Dong, W. Du, J. Dong, R. Che, F. Kong, W. Cheng, M. Ma, N. Gu, Y. Zhang, Depletible peroxidase-like activity of Fe<sub>3</sub>O<sub>4</sub> nanozymes accompanied with separate migration of electrons and iron ions, *Nat. Commun.* 13 (2022) 5365, <https://doi.org/10.1038/s41467-022-33098-y>.
- [48] Y. Huang, Z. Liu, C. Liu, Y. Zhang, J. Ren, X. Qu, Selenium-Based Nanozyme as Biomimetic Antioxidant Machinery, *Chem Eur J* 24 (2018) 10224–10230, <https://doi.org/10.1002/chem.201801725>.
- [49] D. Zhang, N. Shen, J. Zhang, J. Zhu, Y. Guo, L. Xu, A novel nanozyme based on selenopeptide-modified gold nanoparticles with a tunable glutathione peroxidase activity, *RSC Adv.* 10 (2020) 8685–8691, <https://doi.org/10.1039/C9RA10262K>.
- [50] V. Patel, M. Singh, E.L. Mayes, A. Martinez, V. Shutthanandan, V. Bansal, S. Singh, A.S. Karakoti, Ligand-mediated reversal of the oxidation state dependent ROS scavenging and enzyme mimicking activity of ceria nanoparticles, *Chem. Commun.* 54 (2018) 13973–13976, <https://doi.org/10.1039/C8CC08355J>.
- [51] Z. Wang, X. Shen, H. Gao, Y. Zhao, Simultaneous enzyme mimicking and chemical reduction mechanisms for nanoceria as a bio-antioxidant: a catalytic model bridging computations and experiments for nanozymes, *Nanoscale* 11 (2019) 13289–13299, <https://doi.org/10.1039/C9NR03473K>.
- [52] X. Shen, W. Liu, X. Gao, Z. Lu, X. Wu, X. Gao, Mechanisms of Oxidase and Superoxide Dismutation-like Activities of Gold, Silver, Platinum, and Palladium,

- and Their Alloys: A General Way to the Activation of Molecular Oxygen, *J. Am. Chem. Soc.* 137 (2015) 15882–15891, <https://doi.org/10.1021/jacs.5b10346>.
- [53] W. Gao, J. He, L. Chen, X. Meng, Y. Ma, L. Cheng, K. Tu, X. Gao, C. Liu, M. Zhang, K. Fan, D.W. Pang, X. Yan, Deciphering the catalytic mechanism of superoxide dismutase activity of carbon dot nanozyme, *Nat. Commun.* 14 (2023) 160, <https://doi.org/10.1038/s41467-023-35828-2>.
- [54] M. Xia, C. Zhuo, X. Ma, X. Zhang, H. Sun, Q. Zhai, Y. Zhang, Assembly of the active center of organophosphorus hydrolase in metal–organic frameworks via rational combination of functional ligands, *Chem. Commun.* 53 (2017) 11302–11305, <https://doi.org/10.1039/C7CC06270B>.
- [55] S. Li, Z. Zhou, Z. Tie, B. Wang, M. Ye, L. Du, R. Cui, W. Liu, C. Wan, Q. Liu, S. Zhao, Q. Wang, Y. Zhang, S. Zhang, H. Zhang, Y. Du, H. Wei, Data-informed discovery of hydrolytic nanozymes, *Nat. Commun.* 13 (2022) 827, <https://doi.org/10.1038/s41467-022-28344-2>.
- [56] S. Liang, X.L. Wu, M.H. Zong, W.Y. Lou, Construction of Zn-heptapeptide bionanozymes with intrinsic hydrolase-like activity for degradation of di(2-ethylhexyl) phthalate, *J. Colloid Interface Sci.* 622 (2022) 860–870, <https://doi.org/10.1016/j.jcis.2022.04.122>.
- [57] R. Walther, W. van den Akker, A.S. Fruergaard, A.N. Zelikin, Nanozymes and glucuronides: glucuronidase, esterase, and/or transferase activity, *Small* 16 (2020) 2004280, <https://doi.org/10.1002/sml.202004280>.
- [58] P. Nandhakumar, G. Kim, S. Park, S. Kim, J.K. Park, N.S. Lee, Y.H. Yoon, H. Yang, Metal Nanozyme with Ester Hydrolysis Activity in the Presence of Ammonia-Borane and its Use in a Sensitive Immunosensor, *Angew. Chem.* 132 (2020) 22605–22608, <https://doi.org/10.1002/ange.202009737>.
- [59] F. Li, H. Sun, J. Ren, B. Zhang, X. Hu, C. Fang, J. Lee, H. Gu, D. Ling, A nuclease-mimetic platinum nanozyme induces concurrent DNA platinumation and oxidative cleavage to overcome cancer drug resistance, *Nat. Commun.* 13 (2022) 7361, <https://doi.org/10.1038/s41467-022-35022-w>.
- [60] Z. Demirsoy, G. Gulseren, Self-assembled fullerene nanostructures for mimicking and understanding of natural enzymes, *ACS Appl. Nano Mater.* 5 (2022) 14285–14295, <https://doi.org/10.1021/acsnm.2c02194>.
- [61] J. Chen, L. Huang, Q. Wang, W. Wu, H. Zhang, Y. Fang, S. Dong, Bio-inspired nanozyme: a hydratase mimic in a zeolitic imidazolate framework, *Nanoscale* 11 (2019) 5960–5966, <https://doi.org/10.1039/c9nr01093a>.
- [62] Q. Zhou, T. Zhang, J. Jie, Y. Hou, Z. Hu, Z. Jiao, H. Su, TiO<sub>2</sub> as a nanozyme mimicking photolyase to repair DNA damage, *J. Phys. Chem. Lett.* 13 (2022) 10929–10935, <https://doi.org/10.1021/acs.jpclett.2c02717>.
- [63] Z. Tian, T. Yao, C. Qu, S. Zhang, X. Li, Y. Qu, Photolyase-like catalytic behavior of CeO<sub>2</sub>, *Nano Lett.* 19 (2019) 8270–8277, <https://doi.org/10.1021/acs.nanolett.9b03836>.
- [64] F. Li, S. Li, X. Guo, Y. Dong, C. Yao, Y. Liu, Y. Song, X. Tan, L. Gao, D. Yang, Chiral carbon dots mimicking topoisomerase I to mediate the topological rearrangement of supercoiled DNA enantioselectively, *Angew. Chem. Int. Ed.* 59 (2020) 11087–11092, <https://doi.org/10.1002/anie.202002904>.
- [65] A.A. Vernekar, G. Mughes, Hemin-functionalized reduced graphene oxide nanosheets reveal peroxynitrite reduction and isomerization activity, *Chem.–A Europ. J.* 18 (2012) 15122–15132, <https://doi.org/10.1002/chem.201202272>.
- [66] D. Trachootham, W. Lu, M.A. Ogasawara, R.D. Nilsa, P. Huang, Redox regulation of cell survival, *Antioxid. Redox Signal.* 10 (2008) 1343–1374, <https://doi.org/10.1089/ars.2007.1957>.
- [67] S. Liang, X. Tian, C. Wang, Nanozymes in the treatment of diseases caused by excessive reactive oxygen species, *J. Inflamm. Res.* 15 (2022) 6307–6328, <https://doi.org/10.2147/jir.S383239>.
- [68] Y. Zhang, W. Liu, X. Wang, Y. Liu, H. Wei, Nanozyme-enabled treatment of cardio- and cerebrovascular diseases, *Small* (Weinheim an der Bergstrasse Germany) (2022), e2204809, <https://doi.org/10.1002/sml.202204809>.
- [69] J. Liu, M. Wu, R. Zhang, Z.P. Xu, Oxygen-derived free radicals: Production, biological importance, bioimaging, and analytical detection with responsive luminescent nanopores, *View* 2 (2021) 20200139, <https://doi.org/10.1002/VIW.20200139>.
- [70] X. Wu, R. Zhang, X. Yan, K. Fan, Nanozyme: a new approach for anti-microbial infections, *J. Inorg. Mater.* 38 (2023) 43–54, <https://doi.org/10.15541/jim20220578>.
- [71] C. Quijano, M. Trujillo, L. Castro, A. Trostchansky, Interplay between oxidant species and energy metabolism, *Redox Biol.* 8 (2016) 28–42, <https://doi.org/10.1016/j.redox.2015.11.010>.
- [72] J.A. Bauer, M. Zámocká, J. Majtán, V. Bauerová-Hlinková, Glucose Oxidase, an Enzyme “Ferrari”: Its structure, function, production and properties in the light of various industrial and biotechnological applications, *Biomolecules* 12 (3) (2022) 472, <https://doi.org/10.3390/biom12030472>.
- [73] S.B. Bankar, M.V. Bule, R.S. Singhal, L. Ananthanarayan, Glucose oxidase — An overview, *Biotechnol. Adv.* 27 (2009) 489–501, <https://doi.org/10.1016/j.biotechadv.2009.04.003>.
- [74] Y. Chong, Q. Liu, C. Ge, Advances in oxidase-mimicking nanozymes: Classification, activity regulation and biomedical applications, *Nano Today* 37 (2021), 101076, <https://doi.org/10.1016/j.nantod.2021.101076>.
- [75] F. Taranto, A. Pasqualone, G. Mangini, P. Tripodi, M. Miazzi, S. Pavan, C. Montemurro, Polyphenol oxidases in crops: Biochemical, physiological and genetic Aspects, *Int. J. Mol. Sci.* 18 (2) (2017) 377, <https://doi.org/10.3390/ijms18020377>.
- [76] S. Jafarinejad, A. Bigdeli, M. Ghazi-Khansari, P. Sasanpour, M.R. Hormozi-nezhad, Identification of catecholamine neurotransmitters using a fluorescent electronic tongue, *ACS Chem. Neurosci.* 11 (2020) 25–33, <https://doi.org/10.1021/acscchemneuro.9b00537>.
- [77] B. Valderrama, M. Ayala, R. Vazquez-Duhalt, Suicide inactivation of peroxidases and the challenge of engineering more robust enzymes, *Chem. Biol.* 9 (5) (2002) 555–565, [https://doi.org/10.1016/S1074-5521\(02\)00149-7](https://doi.org/10.1016/S1074-5521(02)00149-7).
- [78] N.C. Veitch, Horseradish peroxidase: a modern view of a classic enzyme, *Phytochemistry* 65 (2004) 249–259, <https://doi.org/10.1016/j.phytochem.2003.10.022>.
- [79] F. Attar, M.G. Shahpar, B. Rasti, M. Sharifi, A.A. Saboury, S.M. Rezayat, M. Falahati, Nanozymes with intrinsic peroxidase-like activities, *J. Mol. Liq.* 278 (2019) 130–144, <https://doi.org/10.1016/j.molliq.2018.12.011>.
- [80] L. Zhang, H. Deng, F. Lin, X. Xu, S. Weng, A. Liu, X. Lin, X. Xia, W. Chen, In situ growth of porous platinum nanoparticles on graphene oxide for colorimetric detection of cancer cells, *Anal. Chem.* 86 (2014) 2711–2718, <https://doi.org/10.1021/ac404104j>.
- [81] O. Adeniyi, S. Sicwetsha, P. Mashazi, Nanomagnet-silica nanoparticles decorated with Au@Pd for enhanced peroxidase-like activity and colorimetric glucose sensing, *ACS Appl. Mater. Interfaces* 12 (2020) 1973–1987, <https://doi.org/10.1021/acscami.9b15123>.
- [82] R. Xiao, H. Yu, M. Liu, W. Xu, Y. Qin, R. Tan, Y. Chen, J. Wen, X. Peng, W. Gu, Selective chemical reactivity of non-fullerene acceptor for photoelectrochemical bioassay of urease activity, *Adv. Funct. Mater.* (2023) 2304915, <https://doi.org/10.1002/adfm.202304915>.
- [83] S. Ji, B. Jiang, H. Hao, Y. Chen, J. Dong, Y.u. Mao, Z. Zhang, R. Gao, W. Chen, R. Zhang, Q. Liang, H. Li, S. Liu, Y.u. Wang, Q. Zhang, L. Gu, D. Duan, M. Liang, D. Wang, X. Yan, Y. Li, Matching the kinetics of natural enzymes with a single-atom iron nanozyme, *Nat. Catal.* 4 (5) (2021) 407–417, <https://doi.org/10.1038/s41929-021-00609-x>.
- [84] R. Masuda, R. Kimura, T. Karasaki, S. Sase, K. Goto, Modeling the catalytic cycle of glutathione peroxidase by nuclear magnetic resonance spectroscopic analysis of selenocysteine selenenic acids, *J. Am. Chem. Soc.* 143 (2021) 6345–6350, <https://doi.org/10.1021/jacs.1c02383>.
- [85] X. Da, H. Ji, Z. Zhao, R. Lan, T. Li, J. Ma, Strongly prolonged hydroxyl radical production for Fenton-like reactions: The golden touch of Cu, *Sep. Purif. Technol.* 213 (2019) 500–506, <https://doi.org/10.1016/j.seppur.2018.12.060>.
- [86] J.W. Whittaker, Non-heme manganese catalase—the ‘other’ catalase, *Arch. Biochem. Biophys.* 525 (2012) 111–120, <https://doi.org/10.1016/j.abb.2011.12.008>.
- [87] D. Xu, L. Wu, H. Yao, L. Zhao, Catalase-Like Nanozymes: Classification, Catalytic Mechanisms, and Their Applications, *Small* 18 (2022) 2203400, <https://doi.org/10.1002/sml.202203400>.
- [88] S.I. Sun, H. Liu, Q.I. Xin, K.e. Chen, H. Ma, S. Liu, X. Mu, W. Hao, S. Liu, Y. Gao, Y. Wang, J. Pei, R. Zhao, S. Zhang, X. Zhang, H. Wang, Y. Li, X.-D. Zhang, Atomic engineering of clusterzyme for relieving acute neuroinflammation through lattice expansion, *Nano Lett.* 21 (6) (2021) 2562–2571, <https://doi.org/10.1021/acs.nanolett.0c05148>.
- [89] J. Ming, T. Zhu, J. Li, Z. Ye, C. Shi, Z. Guo, J. Wang, X. Chen, N. Zheng, A novel cascade nanoreactor integrating two-dimensional Pd-Ru nanozyme, uricase and red blood cell membrane for highly efficient hyperuricemia treatment, *Small* 17 (2021) 2103645, <https://doi.org/10.1002/sml.202103645>.
- [90] I. Celardo, J.Z. Pedersen, E. Traversa, L. Ghibelli, Pharmacological potential of cerium oxide nanoparticles, *Nanoscale* 3 (2011) 1411–1420, <https://doi.org/10.1039/C0NR00875C>.
- [91] H. Zhao, R. Zhang, X. Yan, K. Fan, Superoxide dismutase nanozymes: an emerging star for anti-oxidation, *J. Mater. Chem. B* 9 (2021) 6939–6957, <https://doi.org/10.1039/D1TB00720C>.
- [92] H. Younus, Therapeutic potentials of superoxide dismutase, *Int. J. Health Sci.* 12 (2018) 88–93, <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5969776/>.
- [93] H.W. Kroto, J.R. Heath, S.C. O'Brien, R.F. Curl, R.E. Smalley, C60: Buckminsterfullerene, *Nature* 318 (1985) 162–163, <https://doi.org/10.1038/318162a0>.
- [94] E.N. Efrementko, I.V. Lyagin, N.L. Klyachko, T. Bronich, N.V. Zavyalova, Y. Jiang, A.V. Kabanov, A simple and highly effective catalytic nanozyme scavenger for organophosphorus neurotoxins, *J. Control. Release* 247 (2017) 175–181, <https://doi.org/10.1016/j.jconrel.2016.12.037>.
- [95] P. Lin, M. Cao, F. Xia, H. Liao, H. Sun, Q. Wang, J. Lee, Y. Zhou, Y. Guan, C. Zhang, Z. Xu, F. Li, J. Wei, D. Ling, A Phosphatase-mimetic nano-stabilizer of mast cells for long-term prevention of allergic disease, *Adv. Sci.* 8 (2021) 2004115, <https://doi.org/10.1002/advs.202004115>.
- [96] T.L. Spire-Jones, J. Attems, D.R. Thal, Interactions of pathological proteins in neurodegenerative diseases, *Acta Neuropathol.* 134 (2017) 187–205, <https://doi.org/10.1007/s00401-017-1709-7>.
- [97] P. Sweeney, H. Park, M. Baumann, J. Dunlop, J. Frydman, R. Kopito, A. McCampbell, G. Leblanc, A. Venkateswaran, A. Nurmi, Protein misfolding in neurodegenerative diseases: implications and strategies, *Transl. Neurodegener.* 6 (2017) 1–13, <https://doi.org/10.1186/s40035-017-0077-5>.
- [98] F.A. Son, M.C. Wesson, T. Islamoglu, Z. Chen, X. Gong, S.L. Hanna, J. Lyu, X. Wang, K.B. Idrees, J.J. Mahle, G.W. Peterson, O.K. Farha, Uncovering the role of metal-organic framework topology on the capture and reactivity of chemical warfare agents, *Chem. Mater.* 32 (2020) 4609–4617, <https://doi.org/10.1021/acs.chemmater.0c00986>.
- [99] C.T. Supuran, Carbonic anhydrases: novel therapeutic applications for inhibitors and activators, *Nat. Rev. Drug Discov.* 7 (2008) 168–181, <https://doi.org/10.1038/nrd2467>.
- [100] J.K. Kim, C. Lee, S.W. Lim, A. Adhikari, J.T. Andring, R. McKenna, C.M. Ghim, C. U. Kim, Elucidating the role of metal ions in carbonic anhydrase catalysis, *Nat. Commun.* 11 (2020) 4557, <https://doi.org/10.1038/s41467-020-18425-5>.

- [101] R. Occhipinti, W.F. Boron, Role of carbonic anhydrases and inhibitors in acid–base physiology: insights from mathematical modeling, *Int. J. Mol. Sci.* 20 (2019) 3841, <https://doi.org/10.3390/ijms20153841>.
- [102] A. Angeli, F. Carta, C.T. Supuran, Carbonic anhydrases: versatile and useful biocatalysts in chemistry and biochemistry, *Catalysts* 10 (2020) 1008, <https://doi.org/10.3390/catal10091008>.
- [103] Z. Liu, L. Wang, D. Zhong, Dynamics and mechanisms of DNA repair by photolyase, *PCCP* 17 (2015) 11933–11949, <https://doi.org/10.1039/C4CP05286B>.
- [104] M. Zhang, L. Wang, D. Zhong, Photolyase: Dynamics and mechanisms of repair of sun-induced DNA damage, *Photochem. Photobiol.* 93 (2017) 78–92, <https://doi.org/10.1111/php.12695>.
- [105] S. Martínez Cuesta, S.A. Rahman, J.M. Thornton, Exploring the chemistry and evolution of the isomerases, *PNAS* 113 (2016) 1796–1801, <https://doi.org/10.1073/pnas.1509494113>.
- [106] O.A. Asojo, S.K. Nelson, S. Mootien, Y. Lee, W.C. Rezende, D.A. Hyman, M. M. Matsumoto, S. Reiling, A. Kelleher, M. Ledizet, R.A. Koski, K.G. Anthony, Structural and biochemical analyses of alanine racemase from the multidrug-resistant *Clostridium difficile* strain 630, *Acta Crystallogr. D.* 70 (7) (2014) 1922–1933, <https://doi.org/10.1107/S1399004714009419>.
- [107] N. Zhang, X.G. Meng, Y.Y. Wu, H.J. Song, H. Huang, F. Wang, J. Lv, Highly selective isomerization of glucose into fructose catalyzed by a mimic glucose isomerase, *ChemCatChem* 11 (2019) 2355–2361, <https://doi.org/10.1002/cctc.201900143>.
- [108] Y. Sun, C. Zhao, N. Gao, J. Ren, X. Qu, Stereoselective nanozyme based on ceria nanoparticles engineered with amino acids, *Chemistry–A, European Journal.* 23 (2017) 18146–18150, <https://doi.org/10.1002/chem.201704579>.
- [109] M. Omidvar, J. Zdzarta, S.B. Sigurdardóttir, M. Pinelo, Mimicking natural strategies to create multi-environment enzymatic reactors: From natural cell compartments to artificial polyelectrolyte reactors, *Biotechnol. Adv.* 54 (2022), 107798, <https://doi.org/10.1016/j.biotechadv.2021.107798>.
- [110] L. Su, S. Qin, Z. Xie, L. Wang, K. Khan, A.K. Tareen, D. Li, H. Zhang, Multi-enzyme activity nanozymes for biosensing and disease treatment, *Coord. Chem. Rev.* 473 (2022), 214784, <https://doi.org/10.1016/j.ccr.2022.214784>.
- [111] N. Singh, G.R. Sherin, G. Muges, Antioxidant and prooxidant nanozymes: From cellular redox regulation to next-generation therapeutics, *Angew. Chem.* 135 (2023), e202301232, <https://doi.org/10.1002/anie.202301232>.
- [112] N. Singh, M. Geethika, S.M. Eswarappa, G. Muges, Manganese-based nanozymes: Multienzyme redox activity and effect on the nitric oxide produced by endothelial nitric oxide synthase, *Chem. - Eur. J.* 24 (2018) 8393–8403, <https://doi.org/10.1002/chem.201800770>.
- [113] C. Wei, Y. Liu, X. Zhu, X. Chen, Y. Zhou, G. Yuan, Y. Gong, J. Liu, Iridium/ruthenium nanozyme reactors with cascade catalytic ability for synergistic oxidation therapy and starvation therapy in the treatment of breast cancer, *Biomaterials* 238 (2020), 119848, <https://doi.org/10.1016/j.biomaterials.2020.119848>.
- [114] X. Niu, B. Liu, P. Hu, H. Zhu, M. Wang, Nanozymes with multiple activities: prospects in analytical sensing, *Biosensors* 12 (2022) 251, <https://doi.org/10.3390/bios12040251>.
- [115] T. Wu, Z. Ma, P. Li, Q. Lu, M. Liu, H. Li, Y. Zhang, S. Yao, Bifunctional colorimetric biosensors via regulation of the dual nanoenzyme activity of carbonized FeCo-ZIF, *Sens. Actuators B* 290 (2019) 357–363, <https://doi.org/10.1016/j.snb.2019.03.130>.
- [116] F. Erbsloh, A. Bernsmeier, H. Hillesheim, [The glucose consumption of the brain & its dependence on the liver], *Arch. Psychiatr. Nervenkr. Z. Gesamte Neurol. Psychiatr.* 196 (1958) 611–626, <https://doi.org/10.1007/bf00344388>.
- [117] P. Mergenthaler, U. Lindauer, G.A. Dienel, A. Meisel, Sugar for the brain: the role of glucose in physiological and pathological brain function, *Trends Neurosci.* 36 (2013) 587–597, <https://doi.org/10.1016/j.tins.2013.07.001>.
- [118] M. Nimgampalle, H. Chakravarthy, V. Devanathan, Chapter 8 - Glucose metabolism in the brain: An update, in: B. Viswanath (Ed.) *R Recent Dev. Appl. Microbiol. Biochem. Academic Press*, 2021, pp. 77–88.
- [119] Q. Wang, H. Wei, Z. Zhang, E. Wang, S. Dong, Nanozyme: An emerging alternative to natural enzyme for biosensing and immunoassay, *TrAC-Trend Anal. Chem.* 105 (2018) 218–224, <https://doi.org/10.1016/j.trac.2018.05.012>.
- [120] H. Cheng, L. Zhang, J. He, W. Guo, Z. Zhou, X. Zhang, S. Nie, H. Wei, Integrated nanozymes with nanoscale proximity for in vivo neurochemical monitoring in living brains, *Anal. Chem.* 88 (2016) 5489–5497, <https://doi.org/10.1021/acs.analchem.6b00975>.
- [121] Y. Hu, H. Cheng, X. Zhao, J. Wu, F. Muhammad, S. Lin, J. He, L. Zhou, C. Zhang, Y. Deng, P. Wang, Z. Zhou, S. Nie, H. Wei, Surface-enhanced raman scattering active gold nanoparticles with enzyme-mimicking activities for measuring glucose and lactate in living tissues, *ACS Nano* 11 (2017) 5558–5566, <https://doi.org/10.1021/acsnano.7b00905>.
- [122] Y. Ding, G. Ren, G. Wang, M. Lu, J. Liu, K. Li, Y. Lin, V<sub>2</sub>O<sub>5</sub> nanobelts mimic tandem enzymes to achieve nonenzymatic online monitoring of glucose in living rat brain, *Anal. Chem.* 92 (2020) 4583–4591, <https://doi.org/10.1021/acs.analchem.9b05872>.
- [123] M.O. Klein, D.S. Battagello, A.R. Cardoso, D.N. Hauser, J.C. Bittencourt, R. G. Correa, Dopamine: functions, signaling, and association with neurological diseases, *Cell. Mol. Neurobiol.* 39 (2019) 31–59, <https://doi.org/10.1007/s10571-018-0632-3>.
- [124] R. Franco, I. Reyes-Resina, G. Navarro, Dopamine in Health and Disease: Much More Than a Neurotransmitter, *Biomedicines*. 9 (2) (2021) 109, <https://doi.org/10.3390/biomedicines9020109>.
- [125] K. Kang, B. Wang, X. Ji, Y. Liu, W. Zhao, Y. Du, Z. Guo, J. Ren, Hemin-doped metal–organic frameworks based nanozyme electrochemical sensor with high stability and sensitivity for dopamine detection, *RSC Adv.* 11 (4) (2021) 2446–2452, <https://doi.org/10.1039/D0RA08224D>.
- [126] J. Liu, W. Zhang, M. Peng, G. Ren, L. Guan, K. Li, Y. Lin, ZIF-67 as a template generating and tuning “Raisin Pudding”-type nanozymes with multiple enzyme-like activities: Toward online electrochemical detection of 3,4-dihydroxyphenylacetic acid in living brains, *ACS Appl. Mater. Interfaces* 12 (2020) 29631–29640, <https://doi.org/10.1021/acsami.0c05667>.
- [127] D.S. Goldstein, C. Holmes, G.J. Lopez, T. Wu, Y. Sharabi, Cerebrospinal fluid biomarkers of central dopamine deficiency predict Parkinson’s disease, *Parkinsonism Relat. d.* 50 (2018) 108–112, <https://doi.org/10.1016/j.parkrel.2018.02.023>.
- [128] G. Ren, F. Dong, Z. Zhao, K. Li, Y. Lin, Structure defect tuning of metal–organic frameworks as a nanozyme regulatory strategy for selective online electrochemical analysis of uric acid, *ACS Appl. Mater. Interfaces* 13 (2021) 52987–52997, <https://doi.org/10.1021/acsami.1c17974>.
- [129] C. Wang, M. Wang, W. Zhang, J. Liu, M. Lu, K. Li, Y. Lin, Integrating prussian blue analog-based nanozyme and online visible light absorption approach for continuous hydrogen sulfide monitoring in brains of living rats, *Anal. Chem.* 92 (2020) 662–667, <https://doi.org/10.1021/acs.analchem.9b04931>.
- [130] H.M. Albishri, D. Abd El Hady, Hyphenation of enzyme/graphene oxide-ionic liquid/glassy carbon biosensors with anodic differential pulse stripping voltammetry for reliable determination of choline and acetylcholine in human serum, *Talanta* 200 (2019) 107–114, <https://doi.org/10.1016/j.talanta.2019.03.028>.
- [131] R.A. Bekdash, Neuroprotective effects of choline and other methyl donors, *Nutrients* 11 (2019) 2995, <https://doi.org/10.3390/nu11122995>.
- [132] J. Guo, S. Wu, Y. Wang, M. Zhao, A label-free fluorescence biosensor based on a bifunctional MIL-101 (Fe) nanozyme for sensitive detection of choline and acetylcholine at nanomolar level, *Sens. Actuators B* 312 (2020), 128021, <https://doi.org/10.1016/j.snb.2020.128021>.
- [133] Y. Wu, W. Xu, L. Jiao, Y. Tang, Y. Chen, W. Gu, C. Zhu, Defect engineering in nanozymes, *Mater. Today* 52 (2022) 327–347, <https://doi.org/10.1016/j.mattod.2021.10.032>.
- [134] V.L. Feigin, M. Brainin, B. Norrving, S. Martins, R.L. Sacco, W. Hacke, M. Fisher, J. Pandian, P. Lindsay, World stroke organization (WSO): Global stroke fact sheet 2022, *Int. J. Stroke* 17 (2022) 18–29, <https://doi.org/10.1177/17474930211065917>.
- [135] J. Xu, Y. Zhang, G. Nie, Intelligent antithrombotic nanomedicines: Progress, opportunities, and challenges, *VIEW.* 2 (2021) 20200145, <https://doi.org/10.1002/VIW.20200145>.
- [136] U. Dirnagl, C. Iadecola, M.A. Moskowitz, Pathobiology of ischaemic stroke: an integrated view, *Trends Neurosci.* 22 (1999) 391–397, [https://doi.org/10.1016/S0166-2236\(99\)01401-0](https://doi.org/10.1016/S0166-2236(99)01401-0).
- [137] C. Tian, X. Cao, J. Wang, Recanalisation therapy in patients with acute ischaemic stroke caused by large artery occlusion: choice of therapeutic strategy according to underlying aetiological mechanism? *Stroke Vasc. Neurol.* 2 (4) (2017) 244–250, <https://doi.org/10.1136/svn-2017-000090>.
- [138] K. Zhang, M. Tu, W. Gao, X. Cai, F. Song, Z. Chen, Q. Zhang, J. Wang, C. Jin, J. Shi, X. Yang, Y. Zhu, W. Gu, B. Hu, Y. Zheng, H. Zhang, M. Tian, Hollow prussian blue nanozymes drive neuroprotection against ischemic stroke via attenuating oxidative stress, counteracting inflammation, and suppressing cell apoptosis, *Nano Lett.* 19 (2019) 2812–2823, <https://doi.org/10.1021/acs.nanolett.8b04729>.
- [139] A.K. Aranda-Rivera, A. Cruz-Grégorio, Y.L. Arancibia-Hernández, E. Y. Hernández-Cruz, J. Pedraza-Chaverri, RONS and Oxidative Stress: An Overview of Basic Concepts, *Oxygen* 2 (4) (2022) 437–478, <https://doi.org/10.3390/oxygen2040030>.
- [140] R. Kohen, A. Nyska, Oxidation of biological systems: oxidative stress phenomena, antioxidants, redox reactions, and methods for their quantification, *Toxicol. Pathol.* 30 (2002) 620–650, <https://doi.org/10.1080/01926230290166724>.
- [141] Y. Fu, Q. Liu, J. Anrather, F.D. Shi, Immune interventions in stroke, *Nature reviews, Neurology* 11 (2015) 524–535, <https://doi.org/10.1038/nrneuro.2015.144>.
- [142] S. Kobayashi, S. Fukuma, T. Ikenoue, S. Fukuhara, S. Kobayashi, Effect of edaravone on neurological symptoms in real-world patients with acute ischemic stroke, *Stroke* 50 (7) (2019) 1805–1811, <https://doi.org/10.1161/STROKEAHA.118.024351>.
- [143] C. Chen, M. Li, L. Lin, S. Chen, Y. Chen, L. Hong, Clinical effects and safety of edaravone in treatment of acute ischaemic stroke: A meta-analysis of randomized controlled trials, *J. Clin. Pharm. Ther.* 46 (2021) 907–917, <https://doi.org/10.1111/jcpt.13392>.
- [144] C. Xu, X. Qu, Cerium oxide nanoparticle: a remarkably versatile rare earth nanomaterial for biological applications, *NPJ Asia Mater.* 6 (2014) e90, <https://doi.org/10.1038/am.2013.88>.
- [145] Q. Wang, B. Wang, D. Shi, F. Li, D. Ling, Cerium oxide nanoparticles-based optical biosensors for biomedical applications, *Adv. Sensor Res.* 2 (2023) 2200065, <https://doi.org/10.1002/adsr.202200065>.
- [146] C.K. Kim, T. Kim, I.Y. Choi, M. Soh, D. Kim, Y.J. Kim, H. Jang, H.S. Yang, J. Y. Kim, H.K. Park, S.P. Park, S. Park, T. Yu, B.W. Yoon, S.H. Lee, T. Hyeon, Ceria nanoparticles that can protect against ischemic stroke, *Angew. Chem. Int. Ed.* 51 (2012) 11039–11043, <https://doi.org/10.1002/anie.201203780>.
- [147] Q. Bao, P. Hu, Y. Xu, T. Cheng, C. Wei, L. Pan, J. Shi, Simultaneous blood-brain barrier crossing and protection for stroke treatment based on edaravone-loaded

- ceria nanoparticles, *ACS Nano* 12 (2018) 6794–6805, <https://doi.org/10.1021/acsnano.8b01994>.
- [148] L. He, G. Huang, H. Liu, C. Sang, X. Liu, T. Chen, Highly bioactive zeolitic imidazolate framework-8-capped nanotherapeutics for efficient reversal of reperfusion-induced injury in ischemic stroke, *Sci. Adv.* 6 (2020) eaay9751, <https://doi.org/10.1126/sciadv.aay9751>.
- [149] B.H. Chen, B. Stephen Inbaraj, Various physicochemical and surface properties controlling the bioactivity of cerium oxide nanoparticles, *Crit. Rev. Biotechnol.* 38 (2018) 1003–1024, <https://doi.org/10.1080/07388551.2018.1426555>.
- [150] B.C. Yan, J. Cao, J. Liu, Y. Gu, Z. Xu, D. Li, L. Gao, Dietary Fe<sub>3</sub>O<sub>4</sub> nanozymes prevent the injury of neurons and blood-brain barrier integrity from cerebral ischemic stroke, *ACS Biomater. Sci. Eng.* 7 (2021) 299–310, <https://doi.org/10.1021/acsbomaterials.0c01312>.
- [151] Y. Liu, K. Ai, X. Ji, D. Askhatova, R. Du, L. Lu, J. Shi, Comprehensive insights into the multi-antioxidative mechanisms of melanin nanoparticles and their application to protect brain from injury in ischemic stroke, *J. Am. Chem. Soc.* 139 (2017) 856–862, <https://doi.org/10.1021/jacs.6b11013>.
- [152] L. Feng, C. Dou, Y. Xia, B. Li, M. Zhao, P. Yu, Y. Zheng, A.M. El Toni, N.F. Atta, A. Galal, Y. Cheng, X. Cai, Y. Wang, F. Zhang, Neutrophil-like cell-membrane-coated nanozyme therapy for ischemic brain damage and long-term neurological functional recovery, *ACS Nano* 15 (2021) 2263–2280, <https://doi.org/10.1021/acsnano.0c07973>.
- [153] Y. Liu, X. Wang, X. Li, S. Qiao, G. Huang, D.M. Hermann, T.R. Doeppner, M. Zeng, W. Liu, G. Xu, L. Ren, Y. Zhang, W. Liu, E. Casals, W. Li, Y.C. Wang, A Co-doped Fe<sub>3</sub>O<sub>4</sub> nanozyme shows enhanced reactive oxygen and nitrogen species scavenging activity and ameliorates the deleterious effects of ischemic stroke, *ACS Appl. Mater. Interfaces* 13 (2021) 46213–46224, <https://doi.org/10.1021/acsaami.1c06449>.
- [154] Z. Wang, Y. Zhao, Y. Hou, G. Tang, R. Zhang, Y. Yang, X. Yan, K. Fan, A thrombin-activated peptide-templated nanozyme for remedying ischemic stroke via thrombolytic and neuroprotective actions, *Adv. Mater.* (2023) 2210144, <https://doi.org/10.1002/adma.202210144>.
- [155] M.W. Ma, J. Wang, K.M. Dhandapani, R. Wang, D.W. Brann, NADPH oxidases in traumatic brain injury – Promising therapeutic targets? *Redox Biol.* 16 (2018) 285–293, <https://doi.org/10.1016/j.redox.2018.03.005>.
- [156] X. Mu, J. Wang, H. He, Q. Li, B. Yang, J. Wang, H. Liu, Y. Gao, L. Ouyang, S. Sun, Q. Ren, X. Shi, W. Hao, Q. Fei, J. Yang, L. Li, R. Vest, T. Wyss-Coray, J. Luo, X. D. Zhang, An oligomeric semiconducting nanozyme with ultrafast electron transfers alleviates acute brain injury, *Sci. Adv.* 7 (2021) eabk1210, <https://doi.org/10.1126/sciadv.abk1210>.
- [157] X. Mu, H. He, J. Wang, W. Long, Q. Li, H. Liu, Y. Gao, L. Ouyang, Q. Ren, S. Sun, J. Wang, J. Yang, Q. Liu, Y. Sun, C. Liu, X.D. Zhang, W. Hu, Carbogenic nanozyme with ultrahigh reactive nitrogen species selectivity for traumatic brain injury, *Nano Lett.* 19 (2019) 4527–4534, <https://doi.org/10.1021/acs.nanolett.9b01333>.
- [158] H. He, X. Shi, J. Wang, X. Wang, Q. Wang, D. Yu, B. Ge, X. Zhang, F. Huang, Reactive oxygen species-induced aggregation of nanozymes for neuron injury, *ACS Appl. Mater. Interfaces* 12 (2020) 209–216, <https://doi.org/10.1021/acsaami.9b17509>.
- [159] R. Yan, S. Sun, J. Yang, W. Long, J. Wang, X. Mu, Q. Li, W. Hao, S. Zhang, H. Liu, Y. Gao, L. Ouyang, J. Chen, S. Liu, X.D. Zhang, D. Ming, Nanozyme-based bandage with single-atom catalysis for brain trauma, *ACS Nano* 13 (2019) 11552–11560, <https://doi.org/10.1021/acsnano.9b05075>.
- [160] S. Zhang, Y. Liu, S. Sun, J. Wang, Q. Li, R. Yan, Y. Gao, H. Liu, S. Liu, W. Hao, H. Dai, C. Liu, Y. Sun, W. Long, X. Mu, X.D. Zhang, Catalytic patch with redox Cr/CeO<sub>2</sub> nanozyme of noninvasive intervention for brain trauma, *Theranostics* 11 (2021) 2806–2821, <https://doi.org/10.7150/thno.51912>.
- [161] X. Mu, J. Wang, Y. Li, F. Xu, W. Long, L. Ouyang, H. Liu, Y. Jing, J. Wang, H. Dai, Q. Liu, Y. Sun, C. Liu, X.D. Zhang, Redox Trimetallic nanozyme with neutral environment preference for brain injury, *ACS Nano* 13 (2019) 1870–1884, <https://doi.org/10.1021/acsnano.8b08045>.
- [162] J.C. Giugni, M.S. Okun, Treatment of advanced Parkinson's disease, *Curr. Opin. Neurol.* 27 (2014) 450–460, <https://doi.org/10.1097/wco.0000000000000118>.
- [163] A. Li, J. Tyson, S. Patel, M. Patel, S. Katakam, X. Mao, W. He, Emerging nanotechnology for treatment of Alzheimer's and Parkinson's disease, *Front. Bioeng. Biotechnol.* 9 (2021), 672594, <https://doi.org/10.3389/fbioe.2021.672594>.
- [164] L.F. Burbulla, P. Song, J.R. Mazzulli, E. Zampese, Y.C. Wong, S. Jeon, D.P. Santos, J. Blanz, C.D. Obermaier, C. Strojny, J.N. Savas, E. Kiskinis, X. Zhuang, R. Krüger, D.J. Surmeier, D. Krainc, Dopamine oxidation mediates mitochondrial and lysosomal dysfunction in Parkinson's disease, *Science* 357 (6357) (2017) 1255–1261, <https://doi.org/10.1126/science.aam9080>.
- [165] Y. Liu, Y. Mao, E. Xu, H. Jia, S. Zhang, V.L. Dawson, T.M. Dawson, Y. Li, Z. Zheng, W. He, X. Mao, Nanozyme scavenging ROS for prevention of pathologic  $\alpha$ -synuclein transmission in Parkinson's disease, *Nano Today* 36 (2021), 101027, <https://doi.org/10.1016/j.nantod.2020.101027>.
- [166] P.S. Whittom, Inflammation as a causative factor in the aetiology of Parkinson's disease, *British journal of pharmacology*, *Br. J. Pharmacol.* 150 (2007) 963–976, <https://doi.org/10.1038/sj.bjp.0707167>.
- [167] C. Hao, A. Qu, L. Xu, M. Sun, H. Zhang, C. Xu, H. Kuang, Chiral molecule-mediated porous Cu<sub>2</sub>O nanoparticle clusters with antioxidation activity for ameliorating Parkinson's disease, *J. Am. Chem. Soc.* 141 (2019) 1091–1099, <https://doi.org/10.1021/jacs.8b11856>.
- [168] L. Li, Y. Lu, X. Xu, X. Yang, L. Chen, C. Jiang, Y. Wang, W. Hu, X. Wei, Z. Yang, Catalytic-enhanced lactoferrin-functionalized Au-Bi<sub>2</sub>Se<sub>3</sub> nanodots for Parkinson's disease therapy via reactive oxygen attenuation and mitochondrial protection, *Adv. Healthc. Mater.* 10 (2021), e2100316, <https://doi.org/10.1002/adhm.202100316>.
- [169] H.J. Kwon, D. Kim, K. Seo, Y.G. Kim, S.I. Han, T. Kang, M. Soh, T. Hyeon, Ceria nanoparticle systems for selective scavenging of mitochondrial, intracellular, and extracellular reactive oxygen species in Parkinson's disease, *Angew. Chem. Int. Ed.* 57 (2018) 9408–9412, <https://doi.org/10.1002/anie.201805052>.
- [170] X. Ma, J. Hao, J. Wu, Y. Li, X. Cai, Y. Zheng, Prussian blue nanozyme as a pyroptosis inhibitor alleviates neurodegeneration, *Adv. Mater.* 34 (2022) e2106723, <https://doi.org/10.1002/adma.202106723>.
- [171] Q. Li, T. Wu, O.U. Akakuru, N. Song, W. Liu, W. Jiang, K. Fan, A dual synergetic nanoreactor for managing Parkinson's disease by regulating inflammation and mitigating oxidative damage, *Adv. Funct. Mater.* 33 (2023) 2214826, <https://doi.org/10.1002/adfm.202214826>.
- [172] M.A. DeTure, D.W. Dickson, The neuropathological diagnosis of Alzheimer's disease, *Mol. Neurodegener.* 14 (2019) 32, <https://doi.org/10.1186/s13024-019-0333-5>.
- [173] J. de Oliveira, E. Kucharska, M.L. Garcez, M.S. Rodrigues, J. Quevedo, I. Moreno-Gonzalez, J. Budni, Inflammatory cascade in Alzheimer's disease pathogenesis: A review of experimental findings, *Cells* 10 (10) (2021) 2581, <https://doi.org/10.3390/cells10102581>.
- [174] R.H. Swerdlow, J.M. Burns, S.M. Khan, X. Zhu, M.F. Beal, X. Wang, G. Perry, M. A. Smith, The Alzheimer's disease mitochondrial cascade hypothesis, *J. Alzheimer's Dis.* 20 (s2) (2010) S265–S279, <https://doi.org/10.3233/jad-2010-100339>.
- [175] N. Gao, K. Dong, A. Zhao, H. Sun, Y. Wang, J. Ren, X. Qu, Polyoxometalate-based nanozyme: Design of a multifunctional enzyme for multi-faceted treatment of Alzheimer's disease, *Nano Res.* 9 (2016) 1079–1090, <https://doi.org/10.1007/s12274-016-1000-6>.
- [176] M. Ma, Z. Liu, N. Gao, Z. Pi, X. Du, J. Ren, X. Qu, Self-protecting biomimetic nanozyme for selective and synergistic clearance of peripheral amyloid- $\beta$  in an Alzheimer's disease model, *J. Am. Chem. Soc.* 142 (2020) 21702–21711, <https://doi.org/10.1021/jacs.0c08395>.
- [177] Q. Chen, Y. Du, K. Zhang, Z. Liang, J. Li, H. Yu, R. Ren, J. Feng, Z. Jin, F. Li, J. Sun, M. Zhou, Q. He, X. Sun, H. Zhang, M. Tian, D. Ling, Tau-targeted multifunctional nanocomposite for combinational therapy of Alzheimer's disease, *ACS Nano* 12 (2018) 1321–1338, <https://doi.org/10.1021/acsnano.7b07625>.
- [178] Y. Gong, A. Huang, X. Guo, Z. Jia, X. Chen, X. Zhu, Y. Xia, J. Liu, Y. Xu, X. Qin, Selenium-core nanozymes dynamically regulates A $\beta$  & neuroinflammation circulation: Augmenting repair of nervous damage, *Chem. Eng. J.* 418 (2021), 129345, <https://doi.org/10.1016/j.cej.2021.129345>.
- [179] N.S. Caron, E. Dorsey, M.R. Hayden, Therapeutic approaches to Huntington disease: from the bench to the clinic, *Nat. Rev. Drug Discov.* 17 (2018) 729–750, <https://doi.org/10.1038/nrd.2018.133>.
- [180] A. Kumar, R.R. Ratan, Oxidative stress and Huntington's disease: The good, the bad, and the ugly, *J. Huntington's Dis.* 5 (2016) 217–237, <https://doi.org/10.3233/jhd-160205>.
- [181] W. Cong, R. Bai, Y.F. Li, L. Wang, C. Chen, Selenium nanoparticles as an efficient nanomedicine for the therapy of Huntington's disease, *ACS Appl. Mater. Interfaces* 11 (2019) 34725–34735, <https://doi.org/10.1021/acsaami.9b12319>.
- [182] K.A. Aly, M.T. Moutaoufik, M. Zilocchi, S. Phanse, M. Babu, Insights into SACS pathological attributes in autosomal recessive spastic ataxia of Charlevoix-Saguenay (ARSACS)☆, *Curr. Opin. Chem. Biol.* 71 (2022), 102211 <https://doi.org/10.1016/j.cbpa.2022.102211>.
- [183] M. Girard, R. Larivière, D.A. Parfitt, E.C. Deane, R. Gaudet, N. Nossova, F. Blondeau, G. Prenosil, E.G. Vermeulen, M.R. Duchon, A. Richter, E. A. Shoubridge, K. Gehring, R.A. McKinney, B. Brans, J.P. Chapple, P.S. McPherson, Mitochondrial dysfunction and Purkinje cell loss in autosomal recessive spastic ataxia of Charlevoix-Saguenay (ARSACS), *PNAS* 109 (2012) 1661–1666, <https://doi.org/10.1073/pnas.1113166109>.
- [184] C. Criscuolo, C. Procaccini, M.C. Meschini, A. Cianflone, R. Carbone, S. Doccini, D. Devos, C. Nesti, I. Vuillaume, M. Pellegrino, A. Filla, G. De Michele, G. Matarese, F.M. Santorelli, Powerhouse failure and oxidative damage in autosomal recessive spastic ataxia of Charlevoix-Saguenay, *J. Neurol.* 262 (2015) 2755–2763, <https://doi.org/10.1007/s00415-015-7911-4>.
- [185] M. Battaglioli, A. Carmignani, C. Martinelli, J. Colica, A. Marino, S. Doccini, V. Mollo, F. Santoro, M. Bartolucci, A. Petretto, F.M. Santorelli, G. Ciofani, In vitro study of polydopamine nanoparticles as protective antioxidant agents in fibroblasts derived from ARSACS patients, *Biomater. Sci.* 10 (14) (2022) 3770–3792, <https://doi.org/10.1039/D2BM00729K>.
- [186] M. Nehra, U. Uthappa, V. Kumar, R. Kumar, C. Dixit, N. Dilbaghi, Y.K. Mishra, S. Kumar, A. Kaushik, Nanobiotechnology-assisted therapies to manage brain cancer in personalized manner, *J. Control. Release* 338 (2021) 224–243, <https://doi.org/10.1016/j.jconrel.2021.08.027>.
- [187] A.A. Mansur, H.S. Mansur, S.M. Carvalho, Engineered hybrid nanozyme catalyst cascade based on polysaccharide-enzyme-magnetic iron oxide nanostructures for potential application in cancer therapy, *Catal. Today* 388 (2022) 187–198, <https://doi.org/10.1016/j.cattod.2020.06.083>.
- [188] N. Chakraborty, S. Gandhi, R. Verma, I. Roy, Emerging prospects of nanozymes for antibacterial and anticancer applications, *Biomedicines* 10 (2022) 1378, <https://doi.org/10.3390/biomedicines10061378>.
- [189] Y. Wang, F. Gao, X. Li, G. Niu, Y. Yang, H. Li, Y. Jiang, Tumor microenvironment-responsive fenton nanocatalysts for intensified anticancer treatment, *Nanobiotechnol.* 20 (2022) 1–33, <https://doi.org/10.1186/s12951-022-01278-z>.
- [190] A.A. Mansur, S.M. Carvalho, L.C.A. Oliveira, E.M. Souza-Fagundes, Z.I. Lobato, M. F. Leite, H.S. Mansur, Bioengineered carboxymethylcellulose-peptide hybrid

- nanozyme cascade for targeted intracellular biocatalytic–magnetothermal therapy of brain cancer cells, *Pharmaceutics*. 14 (2022) 2223, <https://doi.org/10.3390/pharmaceutics14102223>.
- [191] S.G. Sokolovski, E.U. Rafailov, A.Y. Abramov, P.R. Angelova, Medicine, Singlet oxygen stimulates mitochondrial bioenergetics in brain cells, *Free Radic. Biol. Med.* 163 (2021) 306–313, <https://doi.org/10.1016/j.freeradbiomed.2020.12.022>.
- [192] V. Sunil, J.H. Teoh, B.C. Mohan, A. Mozhi, C. Wang, Bioengineered immunomodulatory organelle targeted nanozymes for photodynamic immunometabolic therapy, *J. Control. Release* 350 (2022) 215–227, <https://doi.org/10.1016/j.jconrel.2022.08.025>.
- [193] C. Coban, M.S.J. Lee, K.J. Ishii, Tissue-specific immunopathology during malaria infection, *Nat. Rev. Immunol.* 18 (2018) 266–278, <https://doi.org/10.1038/nri.2017.138>.
- [194] K. Dorovini-Zis, K. Schmidt, H. Huynh, W. Fu, R.O. Whitten, D. Milner, S. Kamiza, M. Molyneux, T.E. Taylor, The neuropathology of fatal cerebral malaria in malawian children, *Am. J. Pathol.* 178 (2011) 2146–2158, <https://doi.org/10.1016/j.ajpath.2011.01.016>.
- [195] A. Hoffmann, J. Pfeil, J. Alfonso, F.T. Kurz, F. Sahm, S. Heiland, H. Monyer, M. Bendszus, A.-K. Mueller, X. Helluy, M. Pham, K.B. Seydel, Experimental cerebral malaria spreads along the rostral migratory stream, *PLoS Pathog.* 12 (3) (2016), e1005470, <https://doi.org/10.1371/journal.ppat.1005470>.
- [196] A. Pamplona, A. Ferreira, J. Balla, V. Jeney, G. Balla, S. Epiphonio, A. Chora, C. D. Rodrigues, I.P. Gregoire, M. Cunha-Rodrigues, S. Portugal, M.P. Soares, M. M. Mota, Heme oxygenase-1 and carbon monoxide suppress the pathogenesis of experimental cerebral malaria, *Nat. Med.* 13 (6) (2007) 703–710, <https://doi.org/10.1038/nm1586>.
- [197] G. Hasler, Pathophysiology of depression: do we have any solid evidence of interest to clinicians? *World Psychiatry* 9 (2010) 155, <https://doi.org/10.1002/j.2051-5545.2010.tb00298.x>.
- [198] R. Du, F. Wu, M. Lu, X. Shu, J. Ding, G. Wu, G. Hu, Uncoupling protein 2 modulation of the NLRP3 inflammasome in astrocytes and its implications in depression, *Redox Biol.* 9 (2016) 178–187, <https://doi.org/10.1016/j.redox.2016.08.006>.
- [199] Q. Ding, Y. Tian, X. Wang, P. Li, D. Su, C. Wu, W. Zhang, B. Tang, Oxidative damage of tryptophan hydroxylase-2 mediated by peroxisomal superoxide anion radical in brains of mouse with depression, *J. Am. Chem. Soc.* 142 (2020) 20735–20743, <https://doi.org/10.1021/jacs.0c09576>.
- [200] S. Fu, H. Chen, W. Yang, X. Xia, S. Zhao, X. Xu, P.u. Ai, Q. Cai, X. Li, Y.i. Wang, J. Zhu, B. Zhang, J.C. Zheng, ROS-Targeted Depression Therapy via BSA-Incubated Ceria Nanoclusters, *Nano Lett.* 22 (11) (2022) 4519–4527, <https://doi.org/10.1021/acs.nanolett.2c01334>.